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(74) Agent: TUSCAN, Michael, S.; Morgan, Lewis & Bockius LLP, 1800 M. Street, N.W., Washington, DC 20036-5869 (US).

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(71) Applicant (for all designated States except US): YALE UNIVERSITY [US/US]; Room 210, 155 Whitney Avenue, P.O. Box 208336, New Haven, CT 06520-8336 (US).

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(72) Inventors; and

(75) Inventors/Applicants (for US only): DIB-HAJJ, Sulayman [US/US]; Room 210, 155 Whitney Avenue, P.O. Box 208336, New Haven, CT 06520-8336 (US). WAXMAN, Stephen, G. [US/US]; 155 Whitney Avenue, P.O. Box 208336, New Haven, CT 06520-8336 (US).

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(54) Title: MODULATION OF HUMAN SODIUM CHANNELS IN DORSAL ROOT GANGLIA

(57) Abstract: A novel human tetrodotoxin resistant sodium channel is described, along with isolated nucleic acid molecules that encode this channel. Methods for identifying agents that modulate the Na<sup>+</sup> current through the channel are provided, as well as related therapeutic and diagnostic methods.

## MODULATION OF HUMAN SODIUM CHANNELS IN DORSAL ROOT GANGLIA

### FIELD OF THE INVENTION

The present invention relates to a novel human tetrodotoxin-resistant sodium channel and related nucleotides, as well as screening assays for identifying agents useful in treating acute or chronic pain or other hyperexcitability states. This application is related to U.S. Provisional Application 60/072,990, filed January 29, 1998; U.S. Provisional Application 60/109,402 filed November 20, 1998; U.S. Provisional Application 60/109,666 filed on November 20, 1998 ; PCT International Application PCT/US99/02008 filed January 29, 1999 and U.S. Patent Application 09/354,147 filed July 16, 1999, all of which are herein incorporated by reference in their entirety.

### BACKGROUND

#### A. Sodium Channels

Voltage-gated sodium channels are a class of specialized protein molecules that act as molecular batteries permitting excitable cells (neurons and muscle fibers) to produce and propagate electrical impulses. Voltage-gated Na<sup>+</sup> channels from rat brain are composed of three subunits, the pore-forming  $\alpha$  subunit (260 KDa) and two auxiliary subunits,  $\beta$ 1 (36 KDa) and  $\beta$ 2 (33 KDa) that may modulate the properties of the  $\alpha$ -subunit; the  $\alpha$  subunit is sufficient to form a functional channel that generates a Na current flow across the membrane (Catterall, (1993) Trends Neurosci. 16, 500-506; Isom *et al.*, (1994) Neuron 12, 1183-1194). Nine distinct  $\alpha$  subunits have been identified in vertebrates and are encoded by members of an expanding gene family (Goldin (1995) Handbook of receptors and channels (North, editor) CRC Press; Akopian *et al.*, (1996) Nature 379, 257-262; Akopian *et al.*, (1997) FEBS Lett. 400, 183-187; Sangameswaran *et al.*, (1996) J. Biol. Chem. 271, 5953-5956) and respective orthologues of a number of them have been cloned from various mammalian species including humans. Specific  $\alpha$  subunits are expressed in a tissue- and developmentally-specific manner (Beckh *et al.*, (1989) EMBO J. 8, 3611-3616; Mandel, (1992) J. Membr. Biol. 125, 193-205).

Aberrant expression patterns or mutations of voltage-gated sodium channel  $\alpha$ -subunits underlie a number of human and animal disorders (Roden & George, (1997) *Am. J. Physiol.* 273, H511-H525; Ptacek, (1997) *Neuromuscul. Disord.* 7, 250-255; Cannon, (1997) *Neuromuscul. Disord.* 7; 241-249; Cannon, (1996) *Trends Neurosci.* 19, 3-10); Rizzo *et al.*,  
5 (1996) *Eur. Neurol.* 36, 3-12).

Voltage-gated sodium channel  $\alpha$ -subunits consist of four domains (D1-4) of varying internal homology but of similar predicted structure, connected by three intracellular loops (L1-3). The four domains fold to form a channel that opens to both the cytoplasm and the extracellular space via a pore. The pore opens and closes depending upon the physiological  
10 state of the cell membrane.

Each domain consists of six transmembrane segments (S1-6) that allow the protein to weave through the membrane with intra- and extracellular linkers. The linkers of S5-S6 segments of the four domains contain sequences that line the pore of the channel, and a highly conserved subset of amino acids that acts as a filter to selectively allow sodium ions to  
15 traverse the channel pore into the cytoplasm, thus generating an electric current. The amphipathic S4 segment, in each of the four domains, rich in basic residues repeated every third amino acid, acts as a voltage sensor and undergoes a conformational change as a result of the change in the voltage difference across the cell membrane. This in turn triggers the conformational change of the protein to open its pore to the extracellular  $\text{Na}^+$  ion gradient.

20 In most of the known voltage-gated sodium channel  $\alpha$ -subunits the channels close and change into an inoperable state quickly (inactivate) within a few milliseconds after opening of the pore (activation); SNS-type channels, on the other hand, inactivate slowly and require a greater voltage change to activate. L3, the loop that links domains D3 and D4, contains a tripeptide which acts as an intracellular plug that closes the pore after activation, thus inducing  
25 the channel to enter the inactive state. After inactivation, these channels further undergo conformational change to restore their resting state and become available for activation. This period is referred to as recovery from inactivation (repriming). Different channels reprime at different rates, and repriming in SNS is relatively rapid.

Based on amino acid similarities, the voltage-gated sodium channel family has been further subdivided into two subfamilies (Felipe *et al.*, (1994) J. Biol. Chem. 269, 30125-30131). Eight of the nine cloned channels belong to subfamily 1. They share many structural features, particularly in their S4 transmembrane segments. However, some of them  
5 have been shown to have distinct kinetic properties of inactivation and repriming. Only a single channel of subfamily 2, also referred to as atypical channels, has been identified in human, rat and mouse tissues. This subfamily is primarily characterized by reduced numbers of basic residues in its S4 segments, and thus is predicted to have different voltage-dependence compared to subfamily 1. The physiological function of subfamily 2  
10 channels is currently unknown because its electrophysiological properties have not yet been elucidated.

The blocking of voltage-gated sodium channels by tetrodotoxin, a neurotoxin, has served to functionally classify these channels into sensitive (TTX-S) and resistant (TTX-R) phenotypes. Two mammalian TTX-R channels have so far been identified, one specific to the  
15 cardiac muscle and to very limited areas of the central nervous system (CNS) and the second, SNS, is restricted to peripheral neurons (PNS) of the dorsal root ganglia (DRG) and trigeminal ganglia. Specific amino acid residues that confer resistance or sensitivity to TTX have been localized to the ion selectivity filter of the channel pore. The SNS channel is also described in International Patent Application WO 97/01577.

## 20 **B. Role of Sodium Channels in Disease States**

Because different Na<sup>+</sup> channel  $\alpha$ -subunit isotypes exhibit different kinetics and voltage-dependence, the firing properties of excitable cells depend on the precise mixture of channel types that they express. Mutants of the cardiac and skeletal muscle  $\alpha$ -subunit have been shown to cause a number of muscle disorders. Some examples are as follows: A change  
25 of a single basic amino acid residue in the S4 of the skeletal muscle channel is sufficient to change the kinetic properties of this channel and induce a disease state in many patients. A tripeptide deletion in L3 of the cardiac channel, proximal to the inactivation gate, induces a cardiac disorder called Long QT syndrome. A single amino acid change in the S5-S6 linker of

domain 1 of Scn8a, the region lining the pore of the channel, causes the mouse mutant “jolting”. The total loss of this channel by a different mutation causes motor end plate “med” disease in mice. This mutation is characterized by loss of motor neuron stimulation of the innervated muscle.

5 **C. Sodium Channels and Pain**

Axonal injury (injury to nerve fibers, also called axons) can produce chronic pain (termed neuropathic pain). A number of studies have demonstrated altered excitability of the neuronal cell body and dendrites after axonal injury (Eccles *et al.*, (1958) J. Physiol. 143: 11-40; Gallego *et al.*, (1987) J. Physiol. (Lond) 391, 39-56; Kuno & Llinas, (1970) J. Physiol. 10 (Lond.) 210, 807-821), and there is evidence for a change in Na<sup>+</sup> channel density over the neuronal cell body and dendrites following axonal injury (Dodge & Cooley, (1973) IBM J. Res. Dev. 17, 219-229; Titmus & Faber (1986) J. Neurophysiol. 55, 1440-1454; Sernagor *et al.*, (1986) Proc. Natl. Acad. Sci. USA 83, 7966-7970). The expression of abnormal mixtures of different types of sodium channels in a neuronal cell can also lead to abnormal firing (Rizzo 15 *et al.*, (1996) Eur. Neurol. 36, 3-12), and can contribute to hyperexcitability, paresthesia or pain.

Recent studies on rat sensory DRG neurons have demonstrated a dramatic change in the expression profile of TTX-R and TTX-S currents and in a number of mRNA transcripts that could encode the channels responsible for these currents in DRG neurons following 20 various insults (Rizzo *et al.*, (1995) Neurobiol. Dis. 2: 87-96; Cummins *et al.*, (1997) J. Neurophysiol. 17, 3503-3514; Dib-Hajj *et al.*, (1996) Proc. Natl. Acad. Sci. USA 93, 14950-14954). For example, it has been shown an attenuation of the slowly inactivating, TTX-R current and simultaneous enhancement of the rapidly inactivating, TTX-S Na<sup>+</sup> currents in identified sensory cutaneous afferent neurons following axotomy (Rizzo *et al.*, (1995) 25 Neurobiol. Dis. 2, 87-96). A loss of TTX-S, slowly repriming current and TTX-R current and a gain in TTX-S, rapidly repriming current in nociceptive (pain) neurons following axotomy (Cummins & Waxman (1997) J. Neurophysiol. 17, 3503-3514), down-regulation of SNS transcripts and a simultaneous up-regulation of  $\alpha$ -III Transcripts has also been shown

(Dib-Hajj *et al.*, (1996) Proc. Natl. Acad. Sci. USA 93, 14950-14954). Also associated with axotomy is a moderate elevation in the levels of  $\alpha 1$  and  $\alpha II$  mRNAs (Waxman *et al.*, (1994) J. Neurophysiol. 72, 466-470). These changes in the sodium channel profile appear to contribute to abnormal firing that underlies neuropathic pain that patients suffer following axonal injury.

5 Inflammation, which is also associated with pain (termed inflammatory pain), also causes alteration in the sodium current profile in nociceptive DRG neurons. Inflammatory modulators up-regulate TTX-R current in small C-type nociceptive DRG neurons in culture (Gold *et al.*, (1996) Proc. Natl. Acad. Sci. USA 93, 1108-1112; England *et al.*, (1996) J. Physiol. 495, 429-440). The rapid action of these modulators suggests that their action  
10 include posttranslational modification of existing TTX-R channels. It has now been determined that inflammation also increases a TTX-R  $Na^+$  current and up-regulates SNS transcripts in C-type DRG neurons (Tanaka *et al.*, (1998) Neuroreport. 9, 967-972). This data suggests that changes in the sodium current profile contribute to inflammation evoked-pain.

#### **D. Therapies for Chronic Pain:**

15 A variety of classes of drugs (anticonvulsants such as phenytoin and carbamazepine; anti-arrhythmics such as mexitine; local anesthetics such as lidocaine) act on  $Na^+$  channels. Since the various  $Na^+$  channels produce sodium currents with different properties, selective blockade or activation (or other modulation) of specific channel subtypes is expected to be of significant therapeutic value. Moreover, the selective expression of certain  $\alpha$ -subunit isoforms  
20 (PN1, SNS, NaN) in specific types of neurons provides a means for selectively altering their behavior.

Nociceptive neurons of the DRG are the major source of the PNS TTX-R  $Na^+$  current. Thus, the  $Na^+$  channels producing TTX-R currents provide a relatively specific target for the manipulation of pain-producing neurons. The molecular structure of one TTX-R  
25 channel in these DRG neurons, SNS, has been identified but, prior to our research, it has not been determined whether there are other TTX-R channels in these neurons. If such channels could be identified, they would be ideal candidates as target molecules that are preferentially expressed in nociceptive neurons, and whose modulation would attenuate pain transmission.

## SUMMARY OF THE INVENTION

The present invention includes an isolated nucleic acid which encodes a voltage gated Na<sup>+</sup> channel that is preferentially expressed in dorsal root ganglia or trigeminal ganglia (the NaN channel). (In our preceding U.S. Provisional Application 60/072,990, this NaN channel  
5 was referred to by its previous name "NaX"). In a preferred embodiment, the isolated nucleic acid comprises the sequence shown in Figure 1 (SEQ ID NO: 1), Figure 7A (SEQ ID NO: 4), Figure 8A (SEQ ID NO: 6), Figure 11A (SEQ ID NO: 41), allelic variants of said sequences or nucleic acids that hybridize to the foregoing sequences under stringent conditions.

In another embodiment, the invention includes an expression vector comprising an  
10 isolated nucleic acid which encodes the voltage gated Na<sup>+</sup> channel that is preferentially expressed in dorsal root ganglia or trigeminal ganglia either alone or with appropriate regulatory and expression control elements. In a preferred embodiment, the expression vector comprises an isolated nucleic acid having the sequence shown in Figure 1 (SEQ ID NO: 1), Figure 7A (SEQ ID NO: 4), Figure 8A (SEQ ID NO: 6), Figure 11A (SEQ ID NO: 41), allelic  
15 variants of said sequences or nucleic acids that hybridize to the foregoing sequences under stringent conditions.

The present invention further includes a host cell transformed with an expression vector comprising an isolated nucleic acid which encodes a voltage gated Na<sup>+</sup> channel that is preferentially expressed in dorsal root ganglia or trigeminal ganglia with appropriate  
20 regulatory and expression control elements. In a preferred embodiment, the expression vector comprises an isolated nucleic acid having the sequence shown in Figure 1 (SEQ ID NO: 1), Figure 7A (SEQ ID NO: 4), Figure 8A (SEQ ID NO: 6), Figure 11A (SEQ ID NO: 41), allelic variants of said sequences or nucleic acids that hybridize to the foregoing sequences under stringent conditions.

25 The present invention also includes an isolated voltage gated Na<sup>+</sup> channel that is preferentially expressed in dorsal root ganglia or trigeminal ganglia. In a preferred embodiment, the channel has the amino acid sequence of Figure 2 (SEQ ID NO: 3), Figure 7B (SEQ ID NO: 5), Figure 8B (SEQ ID NO: 8) or Figure 11B (SEQ ID NO: 42), or is encoded by a nucleic acid having the sequence shown in Figure 2 (SEQ ID NO: 3), Figure 7B (SEQ ID

NO: 5), Figure 8B (SEQ ID NO: 8) or Figure 11B (SEQ ID NO: 42), allelic variants of said sequences or nucleic acids that hybridize to the foregoing sequences under stringent conditions. Peptide fragments of the channel are also included.

Another aspect of the invention is a method to identify an agent that modulates the  
5 activity of the NaN channel, comprising the steps of bringing the agent into contact with a cell that expresses the Na<sup>+</sup> channel on its surface and measuring depolarization, or any resultant changes in the sodium current. The measuring step may be accomplished with voltage clamp measurements, by measuring depolarization, the level of intracellular sodium or by measuring sodium influx.

10 Another aspect of the invention is a method to identify an agent that modulates the transcription or translation of mRNA encoding the NaN channel. The method comprises the steps of bringing the agent into contact with a cell that expresses the Na<sup>+</sup> channel on its surface and measuring the resultant level of expression of the Na<sup>+</sup> channel.

The invention also includes a method to treat pain, paraesthesia and hyperexcitability  
15 phenomena in an animal or human subject by administering an effective amount of an agent capable of modulating, such as by inhibiting or enhancing, Na<sup>+</sup> current flow through NaN channels in DRG or trigeminal neurons. The method may include administering an effective amount of an agent capable of modulating the transcription or translation of mRNA encoding the NaN channel.

20 Another aspect of the invention is an isolated nucleic acid that is antisense to the nucleic acids described above. In a preferred embodiment, the antisense nucleic acids are of sufficient length to modulate the expression of NaN channel mRNA in a cell containing the mRNA.

Another aspect of the invention is a scintigraphic method to image the loci of pain  
25 generation or provide a measure the level of pain associated with DRG or trigeminal neuron mediated hyperexcitability in an animal or human subject by administering labeled monoclonal antibodies or other labeled ligands specific for the NaN Na<sup>+</sup> channel.



Another aspect of the invention is a method to identify tissues, cells and cell types that express the NaN sodium channel. This method comprises the step of detecting NaN on the cell surface, or en route to the cell surface, or the presence of NaN encoding mRNA.

The present invention further includes a method of producing a transformed cell that  
5 expresses an exogenous NaN encoding nucleic acid, comprising the step of transforming the cell with an expression vector comprising an isolated nucleic acid having the sequence shown in Figures 1, 7A, 8A or 11A, allelic variants of said sequences or nucleic acids that hybridize to the foregoing sequences under stringent conditions, together with appropriate regulatory and expression control elements. The invention also includes a method of producing  
10 recombinant NaN protein, comprising the step of culturing the transformed host under conditions in which the NaN sodium channel or protein is expressed, and recovering the NaN protein.

The invention also includes an isolated antibody specific for the *NaN* channel or polypeptide fragment thereof. The isolated antibody may be labeled.

15 Another aspect of the invention includes a therapeutic composition comprising an effective amount of an agent capable of decreasing rapidly repriming sodium current flow in axotomized, inflamed or otherwise injured DRG neurons or in normal DRG neurons that are being driven to fire at high frequency. The invention also includes a method to treat acute pain or acute or chronic neuropathic or inflammatory pain and hyperexcitability phenomena in  
20 an animal or a human patient by administering the therapeutic composition.

The present invention also includes a method to screen candidate compounds for use in treating pain and hyperexcitability phenomena by testing their ability to alter the expression or activity of an NaN channel mRNA or protein in axotomized, inflamed or otherwise injured DRG neurons.

## BRIEF DESCRIPTION OF THE DRAWING FIGURES

Figure 1 shows the sequence of the rat *NaN* cDNA (SEQ ID NO: 3).

Figure 2 shows the putative amino acid sequence of the rat *NaN* cDNA (SEQ ID NO: 3). Predicted transmembrane segments of domains I - IV are underlined. The amino acid  
5 serine "S" in DI-SS2, implicated in the TTX-R phenotype, is in bold face type.

Figure 3 presents a schematic diagram of predicted secondary structure of the *NaN*  $\alpha$ -subunit.

Figure 4 shows the results of RT-PCR analysis for  $\alpha$ -*NaN* in extracts of various tissues using *NaN*-specific primers. *NaN* is abundantly expressed in dorsal root and  
10 trigeminal ganglia. Low levels of *NaN* are detected in cerebral hemisphere and retina tissues. No detectable *NaN* signal is seen in cerebellum, optic nerve, spinal cord, sciatic nerve, superior cervical ganglia, skeletal muscle, cardiac muscle, adrenal gland, uterus, liver and kidney.

Figure 5 shows the tissue distribution of  $\alpha$ -*NaN* by *in situ* hybridization. Trigeminal  
15 ganglion neurons show moderate-to-high hybridization signal (A). Dorsal root ganglion neurons show moderate-to-high hybridization signal in small neurons (B). Hybridization signal is attenuated in large neurons (arrow). (C) Sense probe shows no signal in DRG neurons. No hybridization signal is seen in spinal cord, cerebellum and liver (D-F). All tissues are from adult Sprague-Dawley rat (scale bars = 50 micrometer).

20 Figure 6 shows the predicted lengths of domain I amplification products of rat  $\alpha$ -subunits and their subunit-specific restriction enzyme profile.

Figure 7 sets forth the (A) nucleotide (SEQ ID NO: 4) and (B) amino acid (SEQ ID NO: 5) sequences of the murine NaN.

Figure 8 is a partial (A) nucleotide sequence (SEQ ID NO: 6) of the human *NaN* and  
25 partial (B) amino acid sequence (SEQ ID NO: 8) of the human NaN protein.

Figure 9 shows cultures of DRG neurons obtained from L4/5 ganglia of adult rats that were reacted with antibody to NaN and then processed for immunofluorescent localization. (A-B) NaN immunostaining is prominent within the cell bodies of DRG neurons. (C) NaN is

present in the neuritic outgrowths, as well as the cell bodies, of DRG neurons. Nomarski (D) and fluorescent (D') images of a neuron that does not express NaN protein.

Figure 10 shows the location of *Scn1 la* and related genes on distal mouse chromosome 9. (A) Haplotypes from the Jackson BSS backcross. Black boxes represent C57BL/6J alleles and white boxes represent SPRET/Ei alleles. The number of animals with each haplotype is given below each column. Missing data was inferred from adjacent data when typing was ambiguous. (B) Map of distal chromosome 9 based on data in (A). Positions of *Scn5a* and *Scn10a* from the MGD consensus map and the locations of the human orthologs are indicated. Numbers are cM positions on the consensus map  
(<http://www.informatics.jax.org/bin/ccr/index>).

Figure 11 shows the (A) cDNA nucleotide sequence (SEQ ID NO: 41) of the human *NaN* gene spanning the complete open reading frame and (B) sets forth the amino acid sequence (SEQ ID NO: 42) of the full length human NaN protein.

## DETAILED DESCRIPTION

The present invention relates to a novel gene that Applicants have discovered, called *NaN*. *NaN* encodes a previously unidentified protein, referred to herein as NaN, that belongs to the  $\alpha$ -subunit voltage-gated sodium channel protein family and that produces a TTX-R sodium current. Such channels underlie the generation and propagation of impulses in excitable cells like neurons and muscle fibers. *NaN* is a novel sodium channel, with a sequence distinct from other, previously identified, channels. The preferential expression of *NaN* on sensory, but not other neurons, makes it a very useful target for diagnostic and/or therapeutic uses in relation to acute and/or chronic pain pathologies..

### A. Definitions

This specification uses several technical terms and phrases which are intended to have the following meanings:

The phrase "modulate" or "alter" refers to up- or down-regulating the level or activity of a particular receptor, ligand or current flow. For example an agent might modulate  $\text{Na}^+$  current flow by inhibiting (decreasing) or enhancing (increasing)  $\text{Na}^+$  current flow. Similarly,

an agent might modulate the level of expression of the NaN sodium channel or the activity of the NaN channels that are expressed.

The phrase "sodium current" or "Na<sup>+</sup> current" means the flow of sodium ions across a cell membrane, often through channels (specialized protein molecules) that are specifically permeable to certain ions, in this case sodium ions.

The phrase "voltage gated" means that the ion channel opens when the cell membrane is in a particular voltage range. Voltage-sensitive sodium channels open when the membrane is depolarized. They then permit Na<sup>+</sup> ions to flow into the cell, producing further depolarization. This permits the cell to generate electrical impulses (also known as "action potentials").

The phrase "rapidly repriming" means that the currents recover from inactivation more rapidly than do such currents in most other voltage gated sodium channel family members.

The terms "TTX-R" and "TTX-S" means that the flow of current through a cell membrane is, respectively, resistant or sensitive to tetrodotoxin (a neurotoxin produced in certain species) at a concentration of about 100 nM.

The phrase "peripheral nervous system (PNS)" means the part of the nervous system outside of the brain and spinal cord, *i.e.*, the spinal roots and associated ganglia such as dorsal root ganglia (DRG) and trigeminal ganglia, and the peripheral nerves.

The phrase "inhibits Na<sup>+</sup> current flow" means that an agent has decreased such current flow relative to a control cell not exposed to that agent. A preferred inhibitor will selectively inhibit such current flow, without affecting the current flow of other sodium channels; or it will inhibit Na<sup>+</sup> current in the channel of interest to a much larger extent than in other channels.

The phrase "enhances Na<sup>+</sup> current flow" means that an agent has increased such current flow relative to a control cell not exposed to that agent. A preferred agent will selectively increase such current flow, without affecting the current flow of other sodium channels; or it will increase Na<sup>+</sup> current in the channel of interest to a much larger extent than in other channels.

The phrase “specifically hybridizes” refers to nucleic acids which hybridize under highly stringent or moderately stringent conditions to the nucleic acids encoding the NaN sodium channel, such as the DNA sequence of SEQ ID NO: 1, 4, 6 or 41.

The phrase “isolated nucleic acid” refers to nucleic acids that have been separated from  
5 or substantially purified relative to contaminant nucleic acids encoding other polypeptides. “Nucleic acids” refers to all forms of DNA and RNA, including cDNA molecules and antisense RNA molecules.

The phrase “RT-PCR” refers to the process of reverse transcription of RNA (RT) using the enzyme reverse transcriptase, followed by the amplification of certain cDNA templates  
10 using the polymerase chain reaction (PCR); PCR requires generic or gene-specific primers and thermostable DNA polymerase, for example, *Taq* DNA polymerase.

The phrase “preferentially expressed” means that voltage gated Na<sup>+</sup> channel is expressed in the defined tissues in detectably greater quantities than in other tissues. For instance, a voltage gated Na<sup>+</sup> channel that is preferentially expressed in dorsal root ganglia or  
15 trigeminal ganglia is found in detectably greater quantities in dorsal root ganglia or trigeminal ganglia when compared to other tissues or cell types. The quantity of the voltage gated Na<sup>+</sup> channel may be detected by any available means, including the detection of specific RNA levels and detection of the channel protein with specific antibodies.

#### **B. Characterization of the NaN Sodium Channel**

20 The present invention relates to a previously unidentified, voltage-gated sodium channel  $\alpha$ -subunit (*NaN*), predicted to be TTX-R, voltage-gated, and preferentially expressed in sensory neurons innervating the body (dorsal root ganglia or DRG) and the face (trigeminal ganglia). The predicted open reading frame (ORF), the part of the sequence coding for the NaN protein molecule, has been determined with the putative amino acid sequence from  
25 different species (rat, mouse, human) presented in Figures 2 (SEQ ID NO: 3), 7B (SEQ ID NO: 5), 8B (SEQ ID NO: 8) or 11B (SEQ ID NO: 42).

All of the relevant landmark sequences of voltage-gated sodium channels are present in NaN at the predicted positions, indicating that *NaN* belongs to the sodium channel family.

But NaN is distinct from all other previously identified Na channels, sharing a sequence identity of less than 53% with each one of them. *NaN* is distinct from *SNS*, the only other TTX-R Na<sup>+</sup> channel subunit that has been identified, until our discovery, in PNS. We have identified and cloned *NaN* without using any primers or probes that are based upon or specific to *SNS*. Moreover, *NaN* and *SNS* share only 47% similarity of their predicted open reading frame (ORF), comparable to the limited similarity of *NaN* to all subfamily 1 members.

The low sequence similarity to existing  $\alpha$ -subunits clearly identifies *NaN* as a novel gene, not simply a variant of an existing channel. Sequence variations compared to the other voltage-gated channels indicate that *NaN* may be the prototype of a novel and previously unidentified, third class of TTX-R channels that may possess distinct properties compared to *SNS*. *NaN* and *SNS*, which are present in nociceptive DRG and trigeminal neurons, may respond to pharmacological interventions in different ways. The preferential expression of *NaN* in sensory DRG and trigeminal neurons provides a target for selectively modifying the behavior of these nerve cells while not affecting other nerve cells in the brain and spinal cord. A further elucidation of the properties of *NaN* channels will be important to understand more fully the effects of drugs designed to modulate the function of the "TTX-R" currents which are characteristic of DRG nociceptive neurons and which contribute to the transmission of pain messages, and to abnormal firing patterns after nerve injury and in other painful conditions..

### C. NaN Nucleic Acids

Nucleic acid molecules of the invention include the nucleotide sequences set forth in Figures 1, 7A, 8A and 11A as well as nucleotide sequences that encode the amino acid sequences of Figures 2, 7B, 8B and 11B. Nucleic acids of the claimed invention also include nucleic acids which specifically hybridize to nucleic acids comprising the nucleotide sequences set forth in Figures 1, 7A, 8A and 11A, or nucleotide sequences which encode the amino acid sequences of Figures 2, 7B, 8B and 11B. A nucleic acid which specifically hybridizes to a nucleic acid comprising that sequence remains stably bound to said nucleic acid under highly stringent or moderately stringent conditions. Stringent and moderately stringent conditions are those commonly defined and available, such as those defined by

Sambrook *et al.*, (1989) Molecular Cloning - A Laboratory Approach, Cold Spring Harbor Press or Ausubel *et al.*, (1995) Current Protocols in Molecular Biology, Greene Publishing. The precise level of stringency is not important, rather, conditions should be selected that provide a clear, detectable signal when specific hybridization has occurred.

5        Hybridization is a function of sequence identity (homology), G+C content of the sequence, buffer salt content, sequence length and duplex melt temperature ( $T_m$ ) among other variables (see Maniatis *et al.*, (1982) Molecular Cloning, Cold Spring Harbor Press). With similar sequence lengths, the buffer salt concentration and temperature provide useful variables for assessing sequence identity (homology) by hybridization techniques. For  
10        example, where there is at least 90 percent homology, hybridization is commonly carried out at 68°C in a buffer salt such as 6×SCC diluted from 20×SSC (see Sambrook *et al.*, (1989) Molecular Cloning - A Laboratory Approach, Cold Spring Harbor Press). The buffer salt utilized for final Southern blot washes can be used at a low concentration, *e.g.*, 0.1×SSC and at a relatively high temperature, *e.g.*, 68°C, and two sequences will form a hybrid duplex  
15        (hybridize). Use of the above hybridization and washing conditions together are defined as conditions of high stringency or highly stringent conditions. Moderately stringent conditions can be utilized for hybridization where two sequences share at least about 80 percent homology. Here, hybridization is carried out using 6×SSC at a temperature of about 50-55°C. A final wash salt concentration of about 1-3×SSC and at a temperature of about  
20        60-68°C are used. These hybridization and washing conditions define moderately stringent conditions.

      In particular, specific hybridization occurs under conditions in which a high degree of complementarity exists between two nucleic acid molecules. With specific hybridization, complementarity will generally be at least about 70%, 75%, 80%, 85%, preferably about  
25        90-100%, or most preferably about 95-100%. When referring the human NaN sequence of SEQ ID NO:41 and 42, preferred homologous sequences will typically encode an NaN protein exhibiting at least about 81% amino acid sequence similarity or at least about 75% or 76% sequence identity to SEQ ID NO: 42. A more preferred human NaN sequence will contain a positively changed residue at amino acid 670, preferably an arginine residue.

As used herein, homology or identity is determined by **BLAST** (Basic Local Alignment Search Tool) analysis using the algorithm employed by the programs **blastp**, **blastn**, **blastx**, **tblastn** and **tblastx** (Karlin *et al.*, (1990) Proc. Natl. Acad. Sci. USA 87, 2264-2268 and Altschul, (1993) J. Mol. Evol. 36, 290-300, both of which are herein  
5 incorporated by reference) which are tailored for sequence similarity searching. The approach used by the **BLAST** program is to first consider similar segments between a query sequence and a database sequence, then to evaluate the statistical significance of all matches that are identified and finally to summarize only those matches which satisfy a preselected threshold of significance. For a discussion of basic issues in similarity searching of sequence databases  
10 (see Altschul *et al.*, Nat. Genet. (1994) 6, 119-129) which is herein incorporated by reference. The search parameters for **histogram**, **descriptions**, **alignments**, **expect** (*i.e.*, the statistical significance threshold for reporting matches against database sequences), **cutoff**, **matrix** and **filter** are at the default settings. The default scoring matrix used by **blastp**, **blastx**, **tblastn**, and **tblastx** is the **BLOSUM62** matrix (Henikoff *et al.*, (1992) Proc. Natl. Acad. Sci. USA 89,  
15 10915-10919, herein incorporated by reference). For **blastn**, the scoring matrix is set by the ratios of **M** (*i.e.*, the reward score for a pair of matching residues) to **N** (*i.e.*, the penalty score for mismatching residues), wherein the default values for **M** and **N** are 5 and -4, respectively.

The nucleic acids of the present invention can be used in a variety of ways in accordance with the present invention. For example, they can be used as nucleic acid probes  
20 to screen other cDNA and genomic DNA libraries so as to select by hybridization other DNA sequences that encode homologous *NaN* sequences. Contemplated nucleic acid probes could be RNA or DNA labeled with radioactive nucleotides or by non-radioactive methods (for example, biotin). Screening may be done at various stringencies (through manipulation of the hybridization  $T_m$ , usually using a combination of ionic strength, temperature and/or presence  
25 of formamide) to isolate close or distantly related homologs. The nucleic acids may also be used to generate primers to amplify cDNA or genomic DNA using polymerase chain reaction (PCR) techniques. The nucleic acid sequences of the present invention can also be used to identify adjacent sequences in the genome, for example, flanking sequences and regulatory elements of *NaN*. The nucleic acids may also be used to generate antisense primers or



constructs that could be used to modulate the level of gene expression of NaN. The amino acid sequence may be used to design and produce antibodies specific to NaN that could be used to localize NaN to specific cells and to modulate the function of NaN channels expressed on the surface of cells.

5 Nucleic acids of the invention also include recombinantly prepared altered NaN sequences. For instance, fusion proteins may be prepared with the open reading frames herein disclosed, or functional fragments thereof, and any available fusion protein. Nucleic acid molecules may also be prepared that encode chimeric NaN proteins, for instance, chimeric proteins comprising individual domains from different species. Such chimeric proteins  
10 include, but are not limited to, human NaN chimeras containing mouse or rat domains, or mouse or rat chimeras containing human domains. Preferred chimeras include human NaN with a rat or mouse domain surrounding the residue equivalent to amino acid 670 of human NaN.

#### **D. Vectors and Transformed Host Cells**

15 The present invention also comprises recombinant vectors containing and capable of replicating and directing the expression of nucleic acids encoding a NaN sodium channel in a compatible host cell. For example, the insertion of a DNA in accordance with the present invention into a vector using enzymes such as T4 DNA ligase, may be performed by any conventional means. Such an insertion is easily accomplished when both the DNA and the  
20 desired vector have been cut with the same restriction enzyme or enzymes, since complementary DNA termini are thereby produced. If this cannot be accomplished, it may be necessary to modify the cut ends that are produced by digesting back single-stranded DNA to produce blunt ends, or by achieving the same result by filling in the single-stranded termini with an appropriate DNA polymerase. In this way, blunt-end ligation may be carried out.  
25 Alternatively, any site desired may be produced by ligating nucleotide sequences (linkers) onto the DNA termini. Such linkers may comprise specific oligonucleotide sequences that encode restriction site recognition sequences.

Any available vectors and the appropriate compatible host cells may be used (Sambrook *et al.*, (1989) Molecular Cloning - A Laboratory Approach, Cold Spring Harbor Press; Ausubel *et al.*, (1995) Current Protocols in Molecular Biology, Greene Publishing). Commercially available vectors, for instance, those available from New England Biolabs,  
5 Promega, Stratagene or other commercial sources are included.

The transformation of appropriate cell hosts with an rDNA (recombinant DNA) molecule of the present invention is accomplished by well known methods that typically depend on the type of vector used and host system employed. Frog oocytes can be injected with RNA and will express channels, but in general, expression in a mammalian cell line  
10 (such as HEK293 or CHO cells) is preferred. With regard to transformation of prokaryotic host cells, electroporation and salt treatment methods are typically employed (see, for example, Cohen *et al.*, (1972) Proc. Natl. Acad. Sci. USA 69, 2110-2114; and Maniatis *et al.*, (1982) Molecular Cloning - A Laboratory Manual, Cold Spring Harbor Press). With regard to transformation of vertebrate cells with vectors containing rDNAs, electroporation, cationic  
15 lipid or salt treatment methods are typically employed (Graham *et al.*, (1973) Virology 52, 456-467; Wigler *et al.*, Proc. Natl. Acad. Sci. USA (1979) 76, 1373-1376).

Successfully transformed cells, *i.e.*, cells that contain an rDNA molecule of the present invention, can be identified by well known techniques. For example, cells resulting from the introduction of an rDNA of the present invention can be cloned to produce single colonies.  
20 Cells from those colonies can be harvested, lysed and their DNA content examined for the presence of the rDNA using conventional methods (Southern, (1975) J. Mol. Biol. 98, 503-517) or the proteins produced from the cell assayed via an immunological method. If tags such as green fluorescent protein are employed in the construction of the recombinant DNA, the transfected cells may also be detected *in vivo* by the fluorescence of such molecules by cell  
25 sorting.

For transient expression of recombinant channels, transformed host cells for the measurement of Na<sup>+</sup> current or intracellular Na<sup>+</sup> levels are typically prepared by co-transfecting constructs into cells such as HEK293 cells with a fluorescent reporter plasmid (such as pGreen Lantern-1, Life Technologies) using the calcium-phosphate precipitation

technique (Ukomadu *et al.*, (1992) Neuron 8, 663-676). HEK293 cells are typically grown in high glucose DMEM (Life Technologies) supplemented with 10% fetal calf serum (Life Technologies). After forty-eight hours, cells with green fluorescence are selected for recording (Dib-Hajj *et al.*, (1997) FEBS Lett. 416, 11-14).

- 5        For preparation of cell lines continuously expressing recombinant channels, the *NaV* construct is cloned into other vectors that carry a selectable marker in mammalian cells. Transfections are carried out using the calcium phosphate precipitation technique (Ukomadu *et al.*, (1992) Neuron 8, 663-676). Human embryonic kidney (HEK-293), chinese hamster ovary (CHO) cells, derivatives of either or other suitable cell lines are grown under standard
- 10    tissue culture conditions in Dulbecco's modified Eagle's medium supplemented with 10% fetal bovine serum. The calcium phosphate-DNA mixture is added to the cell culture medium and left for 15-20 hours, after which time the cells are washed with fresh medium. After 48 hours, antibiotic (G418) is added to select for cells which have acquired neomycin resistance. After 2-3 weeks in G418, 10-20 isolated cell colonies are harvested using sterile 10 ml pipette
- 15    tips. Colonies are grown for another 4-7 days, split and subsequently tested for channel expression using whole-cell patch-clamp recording techniques and RT-PCR.

#### **E. Method of Measuring Na<sup>+</sup> Current Flow**

- Na<sup>+</sup> currents are measured using patch clamp methods (Hamill *et al.*, (1981) Pflügers Arch. 391, 85-100), as described by Rizzo *et al.*, (1994) J. Neurophysiol. 72, 2796-2815 and
- 20    Dib-Hajj *et al.*, (1997) FEBS Lett. 416, 11-14. For these recordings data are acquired on a MacIntosh Quadra 950 or similar computer, using a program such as Pulse (v 7.52, HEKA, German). Fire polished electrodes typically (0.8-1.5 MW) are fabricated from capillary glass using a Sutter P-87 puller or a similar instrument. In the most rigorous analyses, cells are usually only considered for analysis if initial seal resistance is <5 Gohm, they have high
- 25    leakage currents (holding current <0.1 nA at -80 mV), membrane blebs, and an access resistance <5 Mohm. Access resistance is usually monitored throughout the experiment and data are not used if resistance changes occur. Voltage errors are minimized using series resistance compensation and the capacitance artifact is canceled using computer controlled

amplifier circuitry or other similar methods. For comparisons of the voltage dependence of activation and inactivation, cells with a maximum voltage error of  $\pm 10$  mV after compensation are used. Linear leak subtraction is usually used for voltage clamp recordings. Membrane currents are typically filtered at 5 KHz and sampled at 20 KHz. The pipette solution contains  
5 a standard solution such as: 140 mM CsF, 2 mM MgCl, 1 mM EGTA, and 10 mM Na-HEPES (pH 7.3). The standard bathing solution is usually 140 mM NaCl, 3 mM KCl, 2 mM MgCl<sub>2</sub>, 1 mM CaCl<sub>2</sub>, 10 mM HEPES, and 10 mM glucose (pH 7.3).

Voltage clamp studies on transformed cells or DRG neurons, using methods such as intracellular patch-clamp recordings, can provide a quantitative measure of the sodium current  
10 density (and thus the number of sodium channels in a cell), and channel physiological properties. These techniques, which measure the currents that flow through ion channels such as sodium channels, are described in Rizzo *et al.*, (1995) Neurobiol. Dis. 2, 87-96. Alternatively, the blockage or enhancement of sodium channel function can be measured using optical imaging with sodium-sensitive dyes or with isotopically labeled Na. These  
15 methods which are described in Rose *et al.*, (1997) J. Neurophysiol. 78, 3249-3258 and by (Kimelberg & Waltz, (1988) The Neuronal Microenvironment (Boulton *et al.*, editors) Humana Press), measure the increase in intracellular concentration of sodium ions that occurs when sodium channels are open.

#### **F. Measurement of Intracellular Sodium [Na<sup>+</sup>]**

20 The effects of various agents on cells that express Na<sup>+</sup> can be determined using ratiometric imaging of [Na<sup>+</sup>]<sub>i</sub> using SBFI or other similar ion-sensitive dyes. In this method, as described by Sontheimer *et al.*, (1994) J. Neurosci. 14, 2464-2475, cytosolic-free Na<sup>+</sup> is measured using an indicator for Na<sup>+</sup>, such as SBFI (sodium-binding benzofuran isophthalate (Harootunian *et al.*, (1989) J. Biol. Chem. 264, 19458-19467)); or a similar dye. Cells are first loaded with the  
25 membrane-permeable acetoxymethyl ester form of the dye (which is dissolved in dimethyl sulfoxide (DMSO) at a stock concentration of 10 mM). Recordings are obtained on the stage of a microscope using a ratiometric imaging setup (*e.g.*, from Georgia Instruments). Excitation light is provided at appropriate wavelengths (*e.g.*, 340:385 nm). Excitation light is

passed to the cells through a dichroic reflector (400 nm) and emitted light above 450 nm is collected. Fluorescence signals are amplified, *e.g.*, by an image intensifier (GenIISyS) and collected with a CCD camera, or similar device, interfaced to a frame grabber. To account for fluorescence rundown, the fluorescence ratio 340:385 is used to assay cytosolic-free  $\text{Na}^+$ .

- 5 For calibration of SBFI's fluorescence, cells are perfused with calibration solutions containing known  $\text{Na}^+$  concentrations (typically 0 and 30 mM, or 0, 30, and 50 mM  $[\text{Na}^+]$ ), and with ionophores such as gramicidin and monensin (see above) after each experiment. As reported by Rose & Ransom, (1996) *J. Physiol. (Lond)* 491, 291-305, the 345/390 nm fluorescence ratio of intracellular SBFI changes monotonically with changes in  $[\text{Na}^+]_i$ .
- 10 Experiments are typically repeated on multiple (typically at least four) different coverslips, providing statistically significant measurements of intracellular sodium in control cells, and in cells exposed to various concentrations of agents that may block, inhibit or enhance  $\text{Na}^+$ .

#### **G. Method to Measure $\text{Na}^+$ Influx via Measuring $^{22}\text{Na}$ or $^{86}\text{Rb}$**

- $^{22}\text{Na}$  is a gamma emitter and can be used to measure  $\text{Na}^+$  flux (Kimelberg & Waltz, 15 (1988) *The Neuronal Microenvironment* (Boulton *et al.*, editors) Humana Press), and  $^{86}\text{Rb}^+$  can be used to measure  $\text{Na}^+/\text{K}^+$ -ATPase activity (Sontheimer *et al.*, (1994) *J. Neurosci.* 14, 2464-2475).  $^{86}\text{Rb}^+$  ions are taken up by the  $\text{Na}^+/\text{K}^+$ -ATPase-like  $\text{K}^+$  ions, but have the advantage of a much longer half-life than  $^{42}\text{K}^+$  (Kimelberg & Mayhew (1975) *J. Biol. Chem.* 250, 100-104). Thus, measurement of the unidirectional ouabain-sensitive  $^{86}\text{Rb}^+$  uptake 20 provides a quantitative method for assaying  $\text{Na}^+/\text{K}^+$ -ATPase activity which provides another indicator of the electrical firing of nerve cells. Following incubation of cells expressing *NaN* with the isotope  $^{22}\text{Na}^+$ , the cellular content of the isotope is measured by liquid scintillation counting or a similar method, and cell protein is determined using a method such as the bicinchoninic acid protein assay (Smith *et al.*, (1985) *Anal. Biochem.* 150, 76-85) following 25 the modifications described by Goldschmidt & Kimelberg (1989) *Anal. Biochem.* 177, 41-45 for cultured cells.  $^{22}\text{Na}$  and  $^{86}\text{Rb}^+$  fluxes are determined in the presence and absence of agents that may block, inhibit, or enhance *NaN*. This permits determination of the actions of these agents on *NaN*.

#### **H. Method to Identify Agents that Modulate NaN-Mediated Current**

Several approaches can be used to identify agents that are able to modulate (*i.e.*, block or augment) the Na<sup>+</sup> current through the NaN sodium channel. In general, to identify such agents, a model cultured cell line that expresses the NaN sodium channel is utilized, and  
5 one or more conventional assays are used to measure Na<sup>+</sup> current. Such conventional assays include, for example, patch clamp methods, the ratiometric imaging of [Na<sup>+</sup>], and the use of <sup>22</sup>Na and <sup>86</sup>Rb as described above.

In one embodiment of the present invention, to evaluate the activity of a candidate compound to modulate Na<sup>+</sup> current, an agent is brought into contact with a suitable  
10 transformed host cell that expresses *NaN*. After mixing or appropriate incubation time, the Na<sup>+</sup> current is measured to determine if the agent inhibited or enhanced Na<sup>+</sup> current flow.

Agents that inhibit or enhance Na<sup>+</sup> current are thereby identified. A skilled artisan can readily employ a variety of art-recognized techniques for determining whether a particular agent modulates the Na<sup>+</sup> current flow.

15 Because Na<sup>+</sup> is preferentially expressed in pain-signaling cells, one can also design agents that block, inhibit, or enhance Na<sup>+</sup> channel function by measuring the response of laboratory animals, treated with these agents, to acute, inflammatory or chronic pain. In one embodiment of this aspect of the invention, laboratory animals such as rats are treated with an agent for instance, an agent that blocks or inhibits (or is thought to block or inhibit) NaN. The  
20 response to various painful stimuli are then measured using tests such as the tail-flick test and limb withdrawal reflex, and are compared to untreated controls. These methods are described by Dubner, (1994) Textbook of Pain (Wall & Melzack, editors) Churchill Livingstone Publishers. In another embodiment of this aspect of the invention, laboratory animals such as rats are subjected to localized injection of pain-producing inflammatory agents such as  
25 formalin (Dubuisson & Dennis (1977) Pain 4, 161-74), Freund's adjuvant (Iadarola *et al.*, (1988) Pain 35, 313-326) or carageenan, or are subjected to nerve constriction (Bennett & Xie, (1988) Pain 33, 87-107; Kim & Chung (1992) Pain 50, 355-363) or nerve transection (Seltzer *et al.*, (1990) Pain 43, 205-218) which produce persistent pain. The response to various normal and painful stimuli are then measured, for example, by measuring the latency to

withdrawal from a warm or hot stimulus (Dubner, (1994) Textbook of Pain (Wall & Melzack, editors) Churchill Livingstone Publishers) so as to compare control animals and animals treated with agents that are thought to modify NaN.

The preferred inhibitors and enhancers of NaN preferably will be selective for the NaN  
5 Na<sup>+</sup> channel. They may be totally specific (like tetrodotoxin, TTX, which inhibits sodium channels but does not bind to or directly effect any other channels or receptors), or relatively specific (such as lidocaine which binds to and blocks several types of ion channels, but has a predilection for sodium channels). Total specificity is not required for an inhibitor or  
10 enhancer to be efficacious. The ratio of its effect on sodium channels vs. other channels and receptors, may often determine its effect and effects on several channels, in addition to the targeted one, may be efficacious (Stys *et al.*, (1992) J. Neurophysiol. 67, 236-240).  
Modulators of NaN may be combined with or coadministered with agents that modulate other channels expressed in primary sensory neurons, including but not limited to PN1/hNE and SNS/PN3 (Waxman (1999) Pain Supplement 6:S133-140).

15 It is contemplated that modulating agents of the present invention can be, as examples, peptides, small molecules, naturally occurring and other toxins and vitamin derivatives, as well as carbohydrates. A skilled artisan can readily recognize that there is no limit as to the structural nature of the modulating agents of the present invention. Screening of libraries of molecules may reveal agents that modulate NaN or current flow through it. Similarly,  
20 naturally occurring toxins (such as those produced by certain fish, amphibians and invertebrates) can be screened. Such agents can be routinely identified by exposing a transformed host cell or other cell which expresses a sodium channel to these agents and measuring any resultant changes in Na<sup>+</sup> current.

#### **I. Recombinant Protein Expression, Synthesis and Purification**

25 Recombinant NaN proteins can be expressed, for example, in *E. coli* strains HB101, DH5a or the protease deficient strain such as CAG-456 and purified by conventional techniques.

The peptide agents of the invention can be prepared using standard solid phase (or solution phase) peptide synthesis methods, as is known in the art. In addition, the DNA encoding these peptides may be synthesized using commercially available oligonucleotide synthesis instrumentation and produced recombinantly using standard recombinant production systems. The production using solid phase peptide synthesis is necessitated if non-gene-encoded amino acids are to be included.

#### **J. Antibodies and Immunodetection**

Another class of agents of the present invention are antibodies immunoreactive with the Na<sup>+</sup> channel. These antibodies may block, inhibit, or enhance the Na<sup>+</sup> current flow through the channel. Antibodies can be obtained by immunization of suitable mammalian subjects with peptides, containing as antigenic regions, those portions of NaN, particularly (but not necessarily) those that are exposed extracellularly on the cell surface. Such immunological agents also can be used in competitive binding studies to identify second generation inhibitory agents. The antibodies may also be useful in imaging studies, once appropriately labeled by conventional techniques.

#### **K. Production of Transgenic Animals**

Transgenic animals containing and mutant, knock-out or modified *NaN* genes are also included in the invention. Transgenic animals wherein both *NaN* and the *SNS/PN3* gene are modified, disrupted or in some form modified are also included in the present invention. Transgenic animals are genetically modified animals into which recombinant, exogenous or cloned genetic material has been experimentally transferred. Such genetic material is often referred to as a "transgene". The nucleic acid sequence of the transgene, in this case a form of *NaN*, may be integrated either at a locus of a genome where that particular nucleic acid sequence is not otherwise normally found or at the normal locus for the transgene. The transgene may consist of nucleic acid sequences derived from the genome of the same species or of a different species than the species of the target animal.

The term "germ cell line transgenic animal" refers to a transgenic animal in which the genetic alteration or genetic information was introduced into a germ line cell, thereby



conferring the ability of the transgenic animal to transfer the genetic information to offspring. If such offspring in fact possess some or all of that alteration or genetic information, then they too are transgenic animals.

The alteration or genetic information may be foreign to the species of animal to which  
5 the recipient belongs, foreign only to the particular individual recipient, or may be genetic information already possessed by the recipient. In the last case, the altered or introduced gene may be expressed differently than the native gene.

Transgenic animals can be produced by a variety of different methods including transfection, electroporation, microinjection, gene targeting in embryonic stem cells and  
10 recombinant viral and retroviral infection (see, *e.g.*, U.S. Patent No. 4,736,866; U.S. Patent No. 5,602,307; Mullins *et al.*, (1993) Hypertension 22, 630-633; Brenin *et al.*, (1997) Surg. Oncol. 6, 99-110; Tuan (1997) Recombinant Gene Expression Protocols, Humana Press).

A number of recombinant or transgenic mice have been produced, including those which express an activated oncogene sequence (U.S. Patent No. 4,736,866); express simian  
15 SV 40 T-antigen (U.S. Patent No. 5,728,915); lack the expression of interferon regulatory factor 1 (IRF-1) (U.S. Patent No. 5,731,490); exhibit dopaminergic dysfunction (U.S. Patent No. 5,723,719); express at least one human gene which participates in blood pressure control (U.S. Patent No. 5,731,489); display greater similarity to the conditions existing in naturally occurring Alzheimer's disease (U.S. Patent No. 5,720,936); have a reduced capacity to  
20 mediate cellular adhesion (U.S. Patent No. 5,602,307); possess a bovine growth hormone gene (Clutter *et al.*, (1996) Genetics 143, 1753-1760); or, are capable of generating a fully human antibody response (McCarthy, (1997) Lancet 349, 405).

While mice and rats remain the animals of choice for most transgenic experimentation, in some instances it is preferable or even necessary to use alternative animal species.

25 Transgenic procedures have been successfully utilized in a variety of non-murine animals, including sheep, goats, pigs, dogs, cats, monkeys, chimpanzees, hamsters, rabbits, cows and guinea pigs (see Kim *et al.*, (1997) Mol. Reprod. Dev. 46, 515-526; Houdebine, (1995) Reprod. Nutr. Dev. 35, 609-617; Petters (1994) Reprod. Fertil. Dev. 6, 643-645; Schnieke *et al.*, (1997) Science 278, 2130-2133; Amoah, (1997) J. Animal Sci. 75, 578-585).

The method of introduction of nucleic acid fragments into recombination competent mammalian cells can be by any method which favors co-transformation of multiple nucleic acid molecules. Detailed procedures for producing transgenic animals are readily available to one skilled in the art, including the disclosures in U.S. Patent No. 5,489,743 and U.S. Patent  
5 No. 5,602,307.

The specific examples presented below are illustrative only and are not intended to limit the scope of the invention.

## EXAMPLES

### Example 1: Cloning and Characterization of the Rat *NaN* Coding Sequence

#### 10 a. RNA Preparation

Dorsal root ganglia (DRG) from the lumbar region (L4-L5) were dissected from adult Sprague-Dawley rats and total cellular RNA was isolated by the single step guanidinium isothiocyanate-acid phenol procedure (Chomczynski, (1987) Anal. Biochem. 162, 156-159). For analytical applications, DRG tissues were dissected from a few animals at a time. The  
15 quality and relative yield of the RNA was assessed by electrophoresis in a 1% agarose gel. Because of the limited starting material (four DRGs weigh on average 10 mg), quantifying the RNA yield was not attempted. PolyA+ RNA was purified from about 300 mg of total DRG RNA (28 animals) using the PolyATract isolation system according to the manufacturers recommendations (Promega). Half of the purified RNA was used for the preparation of  
20 Marathon cDNA (see below) without further quantification.

#### b. Reverse Transcription

For analytical applications, first strand cDNA was synthesized essentially as previously described (Dib-Hajj *et al.*, (1996) FEBS Lett. 384, 78-82). Briefly, total RNA was reverse transcribed in a 25 ml final volume using 1mM random hexamer (Boehringer  
25 Mannheim) and 500 units SuperScript II reverse transcriptase (Life Technologies) in the presence of 100 units of RNase Inhibitor (Boehringer Mannheim). The reaction buffer

consisted of 50 mM Tris-HCl (pH 8.3), 75 mM KCl, 3 mM MgCl<sub>2</sub>, 10 mM DTT and 125 mM dNTP. The reaction was allowed to proceed at 37°C for 90 minutes, 42°C for 30 minutes, then terminated by heating to 65°C for 10 minutes.

c. First-Strand cDNA Synthesis

5 The Marathon cDNA synthesis protocol was followed according to the manufacturer's instruction as summarized below (all buffers and enzymes are purchased from the manufacturer (Clontech):

Combine the following reagents in a sterile 0.5-ml microcentrifuge tube: 1 mg (1-4 ml) PolyA<sup>+</sup> RNA sample, one ml cDNA Synthesis Primer (10 mM) and sterile water to a final  
10 volume of 5 ml. Mix contents and spin the tube briefly in a microcentrifuge. Incubate the mixture at 70°C for two minutes, then immediately quench the tube on ice for two minutes. Touch-spin the tube briefly to collect the condensation. Add the following to each reaction tube: 2 ml 5× First-Strand Buffer, 1 ml dNTP Mix (10 mM), 1 ml [ $\alpha$ -<sup>32</sup>P]dCTP (1 mCi/ml), 1 ml AMV Reverse Transcriptase (20 units/ml) for a 10 ml volume. The radiolabeled dCTP is  
15 optional (used to determine yield of cDNA) and is replaced by sterile H<sub>2</sub>O if not used. Mix the contents of the tube by gently pipetting and touch-spin the tube to collect the contents at the bottom. Incubate the mixture at 42°C for one hour in an air incubator to reduce condensation and enhance the yield of the first strand cDNA. Place the tube on ice to terminate first-strand synthesis.

20 d. Second-Strand cDNA Synthesis

Combine the following components in the reaction tube from above: 48.4 ml Sterile water, 16 ml 5× Second-Strand Buffer, 1.6 ml dNTP Mix (10 mM), 4 ml 20× Second-Strand Enzyme Cocktail for an 80 ml total volume. Mix the contents thoroughly with gentle pipetting and spin the tube briefly in a microcentrifuge. Incubate the mixture at 16°C for 1.5  
25 hours then add 2 ml (10 units) of T4 DNA Polymerase, mix thoroughly with gentle pipetting and incubate the mixture at 16°C for 45 minutes. Add 4 ml of the EDTA/Glycogen mix to terminate second-strand synthesis. Extract the mixture with an equal volume of buffer-

saturated (pH 7.5) phenol:chloroform:isoamyl alcohol (25:24:1). Mix the contents thoroughly by vortexing and spin the tube in a microcentrifuge at maximum speed (up to 14,000 rpm or 13000×g), 4°C for ten minutes to separate layers. Carefully transfer the top aqueous layer to a clean 0.5-ml tube. Extract the aqueous layer with 100 ml of chloroform:isoamyl alcohol  
5 (24:1), vortex, and spin the tube as before to separate the layers. Collect the top layer into a clean 0.5-ml microcentrifuge tube. Ethanol precipitate the double-stranded cDNA by adding one-half volume of 4 M Ammonium Acetate and 2.5 volumes of room-temperature 95% ethanol. Mix thoroughly by vortexing and spin the tube immediately in a microcentrifuge at top speed, room temperature for twenty minutes. Remove the supernatant carefully and wash  
10 the pellet with 300 ml of 80% ethanol. Spin the tube as before for 10 minutes and carefully remove the supernatant. Air dry the pellet for up to 10 minutes and dissolve the cDNA in 10 ml of sterile H<sub>2</sub>O and store at -20°C. Analyze the yield and size of cDNA by running 2 ml of the cDNA solution on a 1.2% agarose/EtBr gel with suitable DNA size markers (for example, the 1 kilobp ladder, Gibco-BRL). If EtBr staining does not show a signal and [ $\alpha$ -<sup>32</sup>P]dCTP  
15 was included in the reaction, dry the agarose gel on a vacuum gel drying system and expose an x-ray film to the gel overnight at -70°C.

e. Adaptor Ligation

Combine these reagents in a 0.5-ml microcentrifuge test tube, at room temperature, and in the following order: 5 ml double-stranded cDNA, 2 ml Marathon cDNA Adaptor (10  
20 mM), 2 ml 5× DNA Ligation Buffer, 1 ml T4 DNA Ligase (1 unit/ml) for a 10 ml final volume. Mix the contents thoroughly with gentle pipetting and spin the tube briefly in a microcentrifuge. Incubate at either: 16°C overnight; or room temperature (19-23°C) for three to four hours. Inactivate the ligase enzyme by heating the mixture at 70°C for five minutes. Dilute 1 ml of this reaction mixture with 250 ml of Tricine-EDTA buffer and use for RACE  
25 protocols. Store the undiluted adaptor-ligated cDNA at -20°C for future use.

f. PCR

For the initial discovery of *NaN*, we used generic primers designed against highly conserved sequences in domain 1 (D1) of  $\alpha$ -subunits I, II and III and later added more primers to accommodate the new  $\alpha$ -subunits that were discovered. Thus, generic primers were used that recognize conserved sequences in all known  $\text{Na}^+$  channels. The middle of the amplified region shows significant sequence and length polymorphism (Figure 6) and (Gu *et al.*, (1997) J. Neurophysiol. 77, 236-246; Fjell *et al.*, (1997) Mol. Brain Res. 50, 197-204). Due to codon degeneracy, 4 forward primers (F1-F4) and 3 reverse primers (R1-R3) were designed to ensure efficient priming from all templates that might have been present in the cDNA pool (Table 1); however, any of these primers may bind to multiple templates depending on the stringency of the reaction. Forward primer F1 matches subunits  $\alpha\text{I}$ ,  $\alpha\text{III}$ ;  $\alpha\text{Na6}$ ;  $\alpha\text{PN1}$ ;  $\alpha\text{m1}$ ,  $\alpha\text{rH1}$  and  $\alpha\text{SNS/PN3}$ . Sequences of individual subunits show 1 or 2 mismatches to this primer: T to C at position 16 and A to G at position 18 ( $\alpha\text{Na6}$ ); C to R at position 6 ( $\alpha\text{m1}$ ); A to G at position 18 ( $\alpha\text{rH1}$ ) and T to C at position 3 ( $\alpha\text{SNS}$ ). Forward primer F2 matches subunit  $\alpha\text{II}$ . Forward primer F3 perfectly matches  $\alpha\text{Na6}$  and also matches  $\alpha\text{rH1}$  with a single mismatch of C to T at position 16. Reverse primer R1 matches subunits  $\alpha\text{I}$ ,  $\alpha\text{II}$ ,  $\alpha\text{III}$ ,  $\alpha\text{Na6}$ ,  $\alpha\text{PN1}$ ,  $\alpha\text{m1}$  and  $\alpha\text{rH1}$ . This primer has mismatches compared to 4 subunits: G to A at position 3, A to G at position 4 and T to G at position 7 ( $\alpha\text{I}$ ); T to C at position 1 and A to G at position 19 ( $\alpha\text{PN1}$ ); G to A at position 3 and A to G at position 7 ( $\alpha\text{m1}$ ); an extra G after position 3, GC to CT at positions 14-15, and A to T at position 21 ( $\alpha\text{rH1}$ ). Reverse primer R2 matches subunit  $\alpha\text{SNS/PN3}$ .

**Table 1:** Generic and *NaV*-specific primers used for the identification and cloning of *NaV*. All primers except the marathon primers, were synthesized at the department of Pathology, Program for Critical Technologies in Molecular Medicine, Yale University.

Forward Primers	Reverse Primers
5 1. GACCCRTGGAATTGGTTGGA (SEQ ID NO: 9)	1. CAAGAAGGCCCCAGCTGAAGGTGTC (SEQ ID NO: 15)
2. AATCCCTGGAATTGGTTGGA (SEQ ID NO: 10)	2. GAGGAATGCCACGCAAAGGAATC (SEQ ID NO: 16)
10 3. GACCCGTGGAAGTGGTTAGA (SEQ ID NO: 11)	3. AAGAAGGGACCAGCCAAAGTTGTC (SEQ ID NO: 17)
4. GATCTTTGGAAGTGGCTTGA (SEQ ID NO: 12)	4. ACYTCCATRCANWCCCACAT (SEQ ID NO: 18)
5. AACATAGTGCTGGAGTTCAGG (SEQ ID NO: 13)	5. AGRAARTCNAGCCARCACCA (SEQ ID NO: 19)
15 6. GTGGCCTTTGGATTCCGGAGG (SEQ ID NO: 14)	6. TCTGCTGCCGAGCCAGGTA (SEQ ID NO: 20)
	7. CTGAGATAACTGAAATCGCC (SEQ ID NO: 21)
Marathon AP-1 CCATCCTAATACGACTCACTATAGGGC (SEQ ID NO: 22)	
Marathon AP-2 ACTCACTATAGGGCTCGAGCGGC (SEQ ID NO: 23)	

The respective mouse atypical sodium channel mNa<sub>v</sub>2.3 sequence was used to design forward  
 20 primer F4 and reverse primer R3 to amplify the analogous sequence from aNaG, the presumed  
 rat homolog of mNa<sub>v</sub>2.3 (Felipe *et al.*, (1994) J. Biol. Chem. 269, 30125-301231). The  
 amplified sequence was cloned into the *Srf*I site of the vector pCR-Script (Stratagene). The  
 nucleotide sequence of this fragment shows 88% identity to the respective sequence of  
 mNa<sub>v</sub>2.3 (Dib-Hajj & Waxman, unpublished). The restriction enzyme *Xba*I was found to be  
 25 unique to this subunit. Recently, the sequence of a full length cDNA clone of putative sodium  
 channel, NaG-like (SCL-11:Y09164), subunit was published (Akopian *et al.*, (1997) FEBS

Lett. 400, 183-187). The published sequence is 99% identical to our sequence and confirms the size and restriction enzyme polymorphism of the NaG PCR product.

The predicted lengths of amplified products and subunit-specific restriction enzyme recognition sites are shown in Figure 6. All subunit sequences are based on Genbank database  
5 (accession numbers:  $\alpha$ I: X03638;  $\alpha$ II: X03639;  $\alpha$ III: Y00766;  $\alpha$ Na6: L39018;  $\alpha$ hNE-Na: X82835;  $\alpha$ m1 M26643;  $\alpha$ rH1 M27902 and  $\alpha$ SNS X92184; mNa 2.3 L36719).

Subsequently, amplification of NaN sequences 3' terminal to the aforementioned fragment was achieved using NaN-specific primers and two generic reverse primers, R4 and R5. The sequence of the R4 primer was based on the amino acid sequence MWV/DCMEV (SEQ ID  
10 NO: 38) located just N-terminal to domain II S6 segment (see schematic diagram of Figure 3 of voltage-gated sodium channel  $\alpha$ -subunits for reference). The sequence of the R5 primer is based on the amino acid sequence AWCWLDFL (SEQ ID NO: 43) which forms the N-terminal portion of domain III S3 segment.

Amplification was typically performed in 60  $\mu$ l volume using one  $\mu$ l of the first  
15 strand cDNA, 0.8mM of each primer and 1.75 units of Expand Long Template DNA polymerase enzyme mixture (Boehringer Mannheim). Compared to conventional and thermostable DNA polymerases, Expand Long Template enzyme mixture increases the yield of the PCR products without an increase in non-specific amplification (Barnes, (1994) Proc. Natl. Acad. Sci. USA 91, 2216-2220; Cheng *et al.*, (1994) Proc. Natl. Acad. Sci. USA 91,  
20 5695-5699). The PCR reaction buffer consisted of 50 mM Tris-HCl (pH 9.2), 16 mM  $(\text{NH}_4)_2\text{SO}_4$ , 2.25 mM MgCl<sub>2</sub>, 2% (v/v) DMSO and 0.1% Tween 20. As described previously (Dib-Hajj *et al.*, (1996) FEBS Lett. 384, 78-82), amplification was carried out in two stages using a programmable thermal cycler (PTC-200, MJ Research). First, a denaturation step at 94°C for four minutes, an annealing step at 60°C for two minutes and an elongation step at  
25 72°C for 90 seconds. Second, a denaturation step at 94°C for one minute, an annealing step at 60°C for one minute and an elongation step at 72°C for 90 seconds. The second stage was repeated 33 times for a total of 35 cycles, with the elongation step in the last cycle extended to ten minutes

Primary RACE amplification was performed in 50 ml final volume using 4 ml diluted DRG marathon cDNA template, 0.2 mM marathon AP-1 and NaN-specific primers, 3.5 units Expand Long Template enzyme mixture. Extension period was adjusted at 1 minute per 800 base pairs based on the expected product. 5' and 3' RACE amplification was performed using primer pairs marathon AP-1/*NaN*-specific R6 and *NaN*-specific F5/marathon AP-1, respectively. The PCR reaction buffer consisted of 50 mM Tris-HCl (pH 9.2), 16 mM (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 3.0 mM MgCl<sub>2</sub>, 2% (v/v) DMSO and 0.1% Tween 20. Amplification in three stages was performed in a programmable thermal cycler (PTC-200, MJ Research). An initial denaturation step at 92°C was carried out for two minutes. This was followed by 35 cycles consisting of denaturation at 92°C for 20 seconds, annealing step at 60°C for one minute, and an elongation step at 68°C. Finally, an elongation step at 68°C was carried out for five minutes. Nested amplification was performed using 2 ml of a 1/500 diluted primary RACE product in a final volume of 50 ml under similar conditions to the primary RACE reactions. Primer pairs AP-2/*NaN*-specific R7 and *NaN*-specific F6/marathon AP-2 were used for nested 5' and 3' RACE, respectively. Secondary RACE products were band isolated from 1% agarose gels and purified using Qiaex gel extraction kit (Qiagen).

A schematic diagram of the putative structure of *NaN* is shown in Figure 3. The length of the intracellular loops is highly variable both in sequence and length among the various subunits. The exception is the loop between domains III and IV.

20 Example 2: Determination of the Putative Rat Amino Acid Sequence for the NaN Channel

*NaN*-related clones and secondary RACE fragments were sequenced at the W. M. Keck Foundation Biotechnology Resource Lab, DNA sequencing group at Yale University. Sequence analysis including determination of the predicted amino acid sequence was performed using commercial softwares, Lasergene (DNASTar) and GCG. The putative amino acid sequence of *NaN* is shown in Figure 2. Predicted transmembrane segments of domains I - IV are underlined.



### Example 3: Determination of the Murine NaN Sequence

Total RNA extraction from trigeminal ganglia of mice, purification of polyA<sup>+</sup> RNA, and Marathon cDNA construction were done as previously described for the rat. The initial amplification was performed using rat *NaN* primers. The forward primer corresponds to  
5 nucleotides 765-787 of the rat sequence (5' CCCTGCTGCGCTCGGTGAAGAAG 3') (SEQ ID NO: 24), and the reverse primer corresponds to nucleotides 1156-1137 (negative strand) of the rat sequence (5' GACAAAGTAGATCCCAGAGG 3') (SEQ ID NO: 25). The amplification produced a fragment of the expected size. The sequence of this fragment demonstrated high similarity to rat *NaN*. Other fragments were amplified using different rat  
10 primers and primers designed based on the new mouse *NaN* sequence that was being produced. Finally, longer fragments were amplified using mouse Marathon cDNA template and mouse *NaN*-specific primers in combination with adaptor primers that were introduced during the Marathon cDNA synthesis. These fragments were sequenced using primer walking and assembled into Figure 7A.

15 Mouse *NaN* nucleotide sequence, like rat *NaN*, lacks the out-of-frame ATG at the -8 position relative to the translation initiation codon ATG at position 41 (Figure 7A). Translation termination codon TGA is at position 5314. A polyadenylation signal (AATAAA) is present at position 5789 and a putative 23 nucleotide polyA tail is present beginning at position 5800. The sequence encodes an ORF of 1765 amino acids (Figure 7B), which is 90%  
20 similar to rat *NaN*. The gene encoding *NaN* has been named *Scn11a*.

### Chromosomal localization of mouse NaN

A genetic polymorphism between strains C57BL/6J and SPRET/Ei was identified by SSCP analysis of a 274 bp fragment from the 3'UTR of *Scn11a*. Genotyping of 94 animals from the BSS backcross panel (Rowe *et al.*, (1994) Mamm. Genome 5, 253-274)  
25 demonstrated linkage of *Scn11a* with markers on distal chromosome 9 (Figure 10). No recombinants were observed between *Scn11a* and the microsatellite marker *D9Mit19*. Comparison of our data with the MGD consensus map of mouse chromosome 9 revealed close linkage of *Scn11a* with the two other TTX-R voltage-gated sodium channels, *Scn5a* (George

*et al.*, (1995) Cytogenet. Cell. Genet. 68, 67-70) and *Scn10a* (Kozak & Sangameswaran, (1996) Mamm. Genome 7, 787-788; Souslova *et al.*, (1997) Genomics 41, 201-209).

**Example 4: Determination of a Partial and Complete Human NaN Coding Sequence**

Human DRG tissue was obtained from a transplant donor. Total RNA extraction and  
5 cDNA synthesis were performed as described previously.

Forward primer corresponds to sequence 310-294 (minus strand) of EST AA446878. The sequence of the primer is 5' CTCAGTAGTTGGCATGC 3' (SEQ ID NO: 26). Reverse primer corresponds to sequence 270-247 (minus strand) of EST AA88521 1. The sequence of the primer is 5' GGAAAGAAGCACGACCACACAGTC 3' (SEQ ID NO: 27). Amplification  
10 was performed as previously described. PCR amplification was successful and a 2.1 kbp fragment was obtained. This fragment was gel purified and sent for sequencing by primer walking, similar to what is done for mouse *NaN*. The sequence of the ESTs is extended in both directions; the additional sequence shows highest similarity to rat and mouse *NaN*, compared to the rest of the subunits.

15 The sequence of a human 2.1 kbp fragment was obtained using the PCR forward and reverse primers for sequencing from both ends of the fragment. Two additional primers were used to cover the rest of the sequence. The sequence was then extended in the 5' direction using forward primer 1 (above) and human *NaN* reverse primer (5'-GTGCCGTAAACATGAGACTGTCG3') (SEQ ID NO: 44) near the 5' end of the 2.1 kbp  
20 fragment. The partial amino acid sequence is set forth in Figure 8B.

The partial ORF of the human *NaN* consists 1241 amino acids. The sequence is 64% identical to the corresponding sequence of rat *NaN* (73% similar, allowing for conservative substitutions) using the advanced BLAST program available at the National Center for Biotechnology Information. Using the Clustal method of alignment (Lasergene software,  
25 DNASTar) the human *NaN* is 68% and 69% similar to mouse and rat *NaN*, respectively. The respective mouse and rat sequences are 88% similar.

Further sequencing revealed the cDNA sequence spanning the full length open reading frame for the human *NaN* gene. This extended sequence is shown in Figure 11A (SEQ ID

NO: 41). In addition to the features noted with regard to the partial cDNA sequence (Figure 8A), notable features of the extended sequence include a translation start codon (ATG) at position 31 and a translation termination codon at position 5400. A recognizable polyadenylation signal has not been observed and presumably is located 3' of the disclosed sequence. The putative amino acid sequence of the human Nan protein is set forth in Figure 11B (SEQ ID NO: 42).

**Example 5: Isolation of an Alternative Splicing Variant of Rat NaN**

A rat NaN cDNA that encodes a C-terminal truncated version of the full-length rat NaN in Figures 1 and 2 was isolated by sequencing the insert of a rat cDNA clone. The variant NaN cDNA encodes an NaN protein lacking the 387 C-terminal amino acids of the full length NaN and containing a novel 94 amino acid stretch at the C-terminal end. The new sequence arises from the use of a cryptic donor splice site in exon 23 and a novel exon 23', which is located in intron 23. The novel C terminal amino acids are: AAGQAMRKQG DILGPNIHQF SQSSETPFLG CPQQRTCVSF VRPQRVLRVP WFPWRTVTF LSRPRSSESS AWLGLVESSG WSGLPGESGP SSSL (SEQ ID NO: 28). The N-terminal amino acids of the truncated variant are identical to amino acids 1-1378 of the full length rat NaN of Figure 2. The alternative exon and the splicing pattern was confirmed by comparing the cDNA sequence and the genomic sequence in the respective region.

**Example 6: Methods to Isolate Other NaN Sequences**

**a. Isolation of NaN sequences from genomic DNA**

The genomic structure of three voltage-gated Na<sup>+</sup> channel  $\alpha$ -subunits have already been determined (George *et al.*, (1993) Genomics 15, 598-606; Souslova *et al.*, (1997) Genomics 41, 201-209; McClatchey *et al.*, (1992) Hum. Mol. Genet. 1, 521-527; Wang *et al.*, (1996) Genomics 34, 9-16). These genes bear remarkable similarity in their organization and provide a predictable map of most of the exon/intron boundaries. Based on the available rat, mouse and human cDNA sequence of NaN, disclosed herein, PCR primers are designed to amplify NaN homologous sequences from other species using standard PCR protocols.

Alternatively, commercially available genomic DNA libraries are screened with *NaN*-specific probes (based on the rat, mouse, or more preferably, the human sequence) using standard library screening procedures (Sambrook *et al.*, (1989) Molecular Cloning - A Laboratory Approach, Cold Spring Harbor Press; Ausubel *et al.*, (1995) Current Protocols in Molecular Biology, Greene Publishing). This strategy yields genomic DNA isolates that can then be sequenced and the exon/intron boundaries determined by homology to the rat, mouse or human cDNA sequence.

b. Isolation of full length *NaN* sequences allelic variants from autopsy or biopsy tissues

For isolation of human ganglia total RNA, a full length *NaN* human cDNA homologue is isolated from human dorsal root ganglia or trigeminal ganglia or other cranial ganglia from post-mortem human material, foetuses or biopsy or surgical tissues. Total ribonucleic acid (RNA) is isolated from these tissues by extraction in guanidinium isothiocyanate (Saiki *et al.*, (1985) Science 230, 1350-1354) as described in Example 1.

For Determination of the full length transcript size of the human homologue of the rat *NaN* sodium channel cDNA, the method of determining transcript size is as described in Example 9.

Example 7: Production of human DRG cDNA library

A cDNA library from human DRG or trigeminal ganglia polyA<sup>+</sup> RNA was prepared in Example 4 using standard molecular biology techniques (Sambrook *et al.*, (1989) Molecular Cloning - A Laboratory Approach, Cold Spring Harbor Press; Ausubel *et al.*, (1995) Current Protocols in Molecular Biology, Greene Publishing).

PolyA<sup>+</sup> mRNA is hybridized to an oligo(dT) primer and the RNA is copied by reverse transcriptase into single strand cDNA. Then, the RNA in the RNA-DNA hybrid is fragmented by RNase H as *E. coli* DNA polymerase I synthesizes the second-strand fragment. The ends of the double stranded cDNA are repaired, linkers carrying specific restriction enzyme site (for example, Eco RI) are ligated to the ends using *E. coli* DNA ligase. The pool

of the cDNA insert is then ligated into one of a variety of bacteriophage vectors that are commercially available like Lambda-Zap (Stratagene). The procedures are summarized in more detail as follows:

a. First strand cDNA Synthesis

- 5 Dissolve 10 mg poly(A) + RNA at a concentration of 1 mg/ml in sterile water. Heat the RNA for two to five minutes at 65-70°C then quench immediately on ice. In a separate tube add in the following order (180 ml total) : 20 ml of 5 mM dNTPs (500 µM final each), 40 ml 5× RT buffer (1×final), 10 ml 200 mM DTT (10 mM final), 20 ml 0.5 mg/ml oligo (dT)<sub>12-18</sub> (50 mg/ml final), 60 ml deionized water, 10 ml (10 units) RNasin (50 units/ml
- 10 final). Mix by vortexing, briefly microcentrifuge, and add the mixture to the tube containing the RNA. Add 20 ml (200 U) AMV or MMLV reverse transcriptase for a final concentration of 1000 units/ml in 200 ml. Mix by pipetting up and down several times and remove 10 ml to a separate tube containing 1 ml of  $\alpha^{32}\text{P}$  dCTP. Typically, incubate both tubes at room temperature for five minutes, then place both tubes at 42°C for one and a half hours. This
- 15 radiolabeled aliquot is removed to determine incorporation and permit an estimation of recovery; this reaction is stopped by adding 1 ml of 0.5 M EDTA (pH 8.0) and stored frozen at -20°C. The radiolabeled reaction will be used later to estimate the yield and average size of the cDNA inserts. The main reaction is stopped by adding 4 ml of 0.5 M EDTA (pH 8.0) and 200 ml buffered phenol. The mixture is vortexed well, microcentrifuged at room temperature
- 20 for one minute to separate phases, and the upper aqueous layer is transferred to a fresh tube. Back extract the phenol layer with 1×TE buffer (10 mM Tris, 1 mM EDTA, pH 7.5) and pool the aqueous layers from the two extractions. This back extraction of the phenol layer improves the yield. The cDNA is ethanol precipitated using 7.5 M ammonium acetate (final concentration 2.0 to 2.5 M) and 95% ethanol. Place in dry ice/ethanol bath fifteen minutes,
- 25 warm to 4°C, and microcentrifuge at ten minutes at full speed at 4°C to pellet nucleic acids. The small, yellow-white pellet is then washed with ice-cold 70% ethanol, and microcentrifuged for three minutes at full speed, 4°C. Again, the supernatant is removed and the pellet briefly dried.

b. Second strand synthesis

Typically, the pellet from the first-strand synthesis is resuspended in 284 ml water and these reagents are added in the following order (400 ml total): Four ml of 5 mM dNTPs (50  $\mu$ M final each), 80 ml 5 $\times$  second-strand buffer (1 $\times$ final), 12 ml 5 mM  $\beta$ -NAD (150  $\mu$ M final), 2 ml 10 uCi/ml  $\alpha$ -<sup>32</sup>P dCTP (50  $\mu$ Ci/ml final). Mix by vortexing, briefly microcentrifuge, and add: 4 ml (4 units) RNase H (10 units/ml final), 4 ml (20 units) *E. coli* DNA ligase (50 units/ml final), and 10 ml (100 units) *E. coli* DNA polymerase I (250 units/ml final). Mix by pipetting up and down, briefly microcentrifuge, and incubate twelve to sixteen hours at 14°C. After second-strand synthesis, remove 4 ml of the reaction to determine the yield from the incorporation of the radiolabel into acid-insoluble material. Extract the second-strand synthesis reaction with 400 ml buffered phenol and back extract the phenol phase with 200 ml TE buffer (pH 7.5) as described above. The double stranded cDNA is then ethanol precipitated as described above.

To complete the second-strand synthesis the double-stranded cDNA ends are rendered blunt using a mixture of enzymes. Resuspend the pellet in 42 ml water then add these reagents in the following order (80 ml total): 5 ml 5 mM dNTPs (310  $\mu$ M final each), 16 ml 5 $\times$ TA buffer (1 $\times$ final), 1 ml 5 mM  $\beta$ -NAD (62  $\mu$ M final). Mix by vortexing, microcentrifuge briefly, and add: 4 ml of 2 mg/ml RNase A (100 ng/ml final), 4 ml (4 units) RNase H (50 units/ml final), 4 ml (20 units) *E. coli* DNA ligase (250 units/ml final) and 4 ml (8 units) T4 DNA polymerase (100 units/ml final). Mix as above and incubate forty-five minutes at 37°C. Add 120 ml TE buffer (pH 7.5) and 1 ml of 10 mg/ml tRNA. Extract with 200 ml buffered phenol and back extract the phenol layer with 100 ml TE buffer as described above. Pool the two aqueous layers and ethanol precipitate as described above.

c. Addition of linkers to double stranded cDNA

Combine these reagents in a 0.5 ml microcentrifuge test tube, at room temperature, and in the following order: 100 ng double stranded cDNA, 2 ml linkers/adaptors (10 mM), 2 ml 5 $\times$  DNA Ligation Buffer, 1 ml T4 DNA Ligase (unit/ml) for a 10 ml final volume. Mix the contents thoroughly with gentle pipetting and spin the tube briefly in a microcentrifuge.

Incubate at either: 16°C overnight; or room temperature (19-23°C) for three to four hours. Inactivate the ligase enzyme by heating the mixture at 70°C for five minutes. This cDNA is typically digested by *Eco* RI to prepare the cohesive ends of the cDNA for ligation into the vector and to cleave linker concatemers. Typically this reaction consists of the 10 ml of the  
5 cDNA, 2 ml of 10× *Eco* RI buffer (depending on the company of source), 2 ml of *Eco* RI (10 units/ml) and sterile water to a final volume of 20 ml. The mixture is incubated at 37°C for two to four hours.

d. Size fractionation of cDNA

Size exclusion columns are typically used to remove linker molecules and short  
10 cDNA fragments (350 bp). For example, a 1-ml Sepharose CL-4B column is prepared in a plastic column plugged with a small piece of sterilized glass wool (a 5 ml plastic pipet will work fine). The column is equilibrated with 0.1 M sodium chloride in 1×TE (10mM Tris, 1 mM EDTA, pH 7.5). The cDNA is then loaded onto the column and 200 µl fractions are collected. 2 µl aliquots of each fraction are analyzed by gel electrophoresis and  
15 autoradiography to determine the peak of cDNA elution. Typically, fractions containing the first half of the peak are pooled and purified by ethanol precipitation and resuspending in 10 µl distilled water.

e. Cloning of cDNA into bacteriophage vector

Bacteriophage vectors designed for the cloning and propagation of cDNA are  
20 provided ready-digested with *Eco* RI and with phosphatased ends from commercial sources (e.g., lambda gt10 from Stratagene). The prepared cDNA is ligated into lambda vectors following manufacturer's instructions. Ligated vector/cDNA molecules are packaged into phage particles using packaging extracts available commercially.

**Example 8: Screening of Human cDNA Library**

**a. Labeling of cDNA fragments (probes) for library screening**

An RNA probe is used that recognizes nucleotide sequences that are specific to *NaN*, such as 1371-1751 of *NaN*. Other nucleotide sequences can be developed on the basis of the

5 *NaN* sequence (Figures 2, 7 & 8) such as nucleotides 765-1160 of the human nucleotide sequence. A *Hind* III/*Bam* HI fragment of *NaN* was inserted in pBluescript (SK+) vector (Stratagene). The sequence of the resulting construct was verified by sequencing. The orientation of the insert is such that the 5' and 3' ends of the construct delineated by the *Hind* III and *Bam* HI restriction enzyme sites, respectively, are proximal to T7 and T3 RNA

10 polymerase promoters, respectively. Digoxigenin-labeled Sense (linearized at the *Hind* III site and transcribed by T7 RNA polymerase) and antisense (linearized at the *Bam* HI site and transcribed by T3 RNA polymerase) transcripts were prepared in vitro using MEGAscript transcription kit (Ambion) according to manufacturer specifications. Briefly, 1 µg linearized template was transcribed with the respective RNA polymerase in a 20 µl final volume

15 containing the following reagents: 1× enzyme mixture containing the respective RNA polymerase and RNase inhibitor and reaction buffer (Ambion), 7.5 mM ATP, GTP and CTP nucleotides, 5.625 mM UTP and 1.725 mM Dig-11UTP (Boehringer Mannheim). *In vitro* transcription was carried out at 37°C for three hours in a water bath. DNA template was removed by adding 1 µl of RNase-free DNase I (2 units/µl) to each reaction and incubating

20 further at 37°C for fifteen minutes. The reaction was then stopped by adding 30 µl nuclease-free water) and 25 µl of LiCl precipitation solution (7.5 M Lithium Chloride, 50 mM EDTA).

The mixture was incubated at -20°C for thirty minutes. The RNA transcripts were pelleted in a microfuge at 13000×g, 4°C for fifteen minutes. The supernatant was removed and the pellet washed once with 100 µl of 75% ethanol. The mixture was re-centrifuged at

25 13000×g, room temperature for five minutes. The pellet was then air-dried in a closed chamber and subsequently dissolved in 100 µl of RNase-free water. The transcript yield and integrity were determined by comparison to a control DIG-labeled RNA on agarose-formaldehyde gel as described in the DIG/Genius kit according to manufacturer



recommendations (Boehringer Mannheim). Alternatively, a skilled artisan can design radioactive probes for autoradiographic analysis.

Other regions of the rat, mouse or human *NaN* sodium channel cDNA, like 3' untranslated sequences, can also be used as probes in a similar fashion for cDNA library screening or Northern blot analysis. Specifically, a probe is made using commercially available kits, such as the Pharmacia oligo labeling kit, or Genius kit (Boehringer Mannheim).

b. cDNA library screening

Recombinant plaques containing full length human homologues of the *NaN* sodium channel are detected using moderate stringency hybridization washes (50-60°C, 5×SSC, thirty minutes), using non-radioactive (see above) or radiolabeled DNA or cRNA *NaN*-specific probes derived from the 3' untranslated or other regions as described above. Libraries are screened using standard protocols (Sambrook *et al.*, (1989) Molecular Cloning - A Laboratory Approach, Cold Spring Harbor Press; Ausubel *et al.*, (1995) Current Protocols in Molecular Biology, Greene Publishing) involving the production of nitrocellulose or nylon membrane filters carrying recombinant phages. The recombinant DNA is then hybridized to *NaN*-specific probes (see above). Moderate stringency washes are carried out.

Plaques which are positive on duplicate filters (*i.e.*, not artefacts or background) are selected for further purification. One or more rounds of screening after dilution to separate the phage are typically performed. Resulting plaques are then purified, DNA is extracted and the insert sizes of these clones characterized. The clones are cross-hybridized to each other using standard techniques (Sambrook *et al.*, (1989) Molecular Cloning - A Laboratory Approach, Cold Spring Harbor Press) and distinct positive clones identified.

Typically, overlapping clones that encode the channel are isolated. Standard cloning techniques are then used to produce a full length cDNA construct that contains any 5' untranslated sequence, the start codon ATG, the coding sequence, a stop codon and any 3' untranslated sequence, a poly A consensus sequence and possibly a poly A run. If overlapping clones do not produce sufficient fragments to assemble a full length cDNA clone, alternative

methods like RACE (PCR-based) could be used to generate the missing pieces or a full length clone.

c. Characterization of a human homologue full-length clone

- A *NaV*-specific cDNA sequence from a full-length clone is used as a probe in
- 5 Northern blot analysis to determine the messenger RNA size in human tissue for comparison with the rat and mouse messenger RNA size. Confirmation of biological activity of the cloned cDNA is carried out using methods similar to those described for the rat *NaV*.

Example 9: Polymerase chain reaction (PCR) approaches to clone other full length human *NaV* sodium channels using DNA sequences derived from rat, mouse or human amino acid

10 sequences

Total RNA and poly A<sup>+</sup> RNA is isolated from human dorsal root ganglia or trigeminal ganglia or other cranial ganglia from post-mortem human material or foetuses or biopsy/surgical tissues as described above. Preparation of cDNA and PCR-based methods are then used as described previously in Example 1.

- 15 Using degenerate PCR primers derived from the rat, mouse or human *NaV*-specific coding sequence (see Figures 2 (SEQ ID NO: 3), 7B (SEQ ID NO: 5), 8B (SEQ ID NO: 8) and 11B (SEQ ID NO: 41)), the cDNA is amplified using the polymerase chain reaction (Saiki *et al.*, (1985) Science 230, 1350-1354). A skilled artisan could utilize the many variables which can be manipulated in a PCR reaction to derive the homologous sequences required.
- 20 These include, but are not limited to, varying cycle and step temperatures, cycle and step times, number of cycles, thermostable polymerase, and Mg<sup>2+</sup> concentration. A greater specificity can be achieved using nested primers derived from further conserved sequences from the *NaV* sodium channel.

- Amplification is typically performed in 60 µl volume using 1 µl of the first strand
- 25 cDNA, 0.8 mM of each primer and 1.75 units of Expand Long Template DNA polymerase enzyme mixture (Boehringer Mannheim). Compared to conventional and thermostable DNA polymerases, Expand Long Template enzyme mixture increases the yield of the PCR products

without an increase in non-specific amplification (Barnes, (1994) Proc. Natl. Acad. Sci. USA 91, 2216-2220; Cheng *et al.*, (1994) Proc. Natl. Acad. Sci. USA 91, 5695-5699). The PCR reaction buffer consists of 50 mM Tris-HCl (pH 9.2), 16 mM (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 2.25 mM MgCl<sub>2</sub>, 2% (v/v) DMSO and 0.1% Tween 20. As described previously (Dib-Hajj *et al.*, (1996) FEBS Lett. 384, 78-82), amplification is carried out in two stages using a programmable thermal cycler (PTC-200, MJ Research). First, a denaturation step at 94°C for four minutes, an annealing step at 60°C for two minutes and an elongation step at 72°C for ninety seconds. Second, a denaturation step at 94°C for one minute, an annealing step at 60°C for one minute and an elongation step at 72°C for ninety seconds. The second stage is repeated 33 times for a total of 35 cycles, with the elongation step in the last cycle extended to 10 minutes. In addition, control reactions are performed alongside the samples. These should be: (1) all components without cDNA, (negative control) and (2) all reaction components with primers for constitutively expressed product, *e.g.*, GAPDH.

The products of the PCR reactions are examined on 1-1.6% (w/v) agarose gels. Bands on the gel (visualized by staining with ethidium bromide and viewing under UV light) representing amplification products of the approximate predicted size are then cut from the gel and the DNA purified.

The resulting DNA may be sequenced directly or is ligated into suitable vectors such as, but not limited to, pCR II (Invitrogen) or pGEMT (Promega). Clones are then sequenced to identify those containing sequence with similarity to the rat, mouse or partial human *NaV* sodium channel sequence.

#### Example 10: Clone analysis

Candidate clones from Example 9 are further characterized by conventional techniques. The biological activity of expression products is also confirmed using conventional techniques.

**Example 11: Isolation of full length *NaN* sequences from human fetal tissues**

Commercially available human fetal cDNA libraries and/or cDNA pools are screened with *NaN*-specific primers (by PCR) or probes (library screening) using PCR standard PCR protocols and standard library screening procedures as described above.

**5    Example 12: Northern Blot of rat DRG or Trigeminal Neurons with Fragments of Rat *NaN***

10-30 µg total DRG and/or RNA from DRG or trigeminal (for positive tissues) and cerebral hemisphere, cerebellum and liver (for negative tissues) is electrophoresed in denaturing 1% agarose-formaldehyde gel or agarose-glyoxal gel, and then is transferred to a nylon membrane as described in achieved in multiple steps, as detailed in standard molecular biology manuals (Sambrook *et al.*, (1989) Molecular Cloning - A Laboratory Approach, Cold Spring Harbor Press; Ausubel *et al.*, (1995) Current Protocols in Molecular Biology, Greene Publishing). Radiolabeled (specific activity of  $>10^8$  dpm/µg) or Digoxigenin-labeled RNA probes are typically used for Northern analysis. An antisense RNA probe (see Example 20, which describes *in situ* hybridization with a *NaN*-specific probe) is created by *in vitro* synthesis from a sense DNA fragment. The membrane carrying the immobilized RNA in wetted with 6×SSC, and the membrane is placed RNA-side-up in a hybridization tube. One ml formamide prehybridization/hybridization solution per 10 cm<sup>2</sup> of membrane is added. Prehybridization and hybridization are usually carried out in glass tubes in a commercial hybridization oven. The tubes are place in a hybridization oven and incubated, with rotation, at 60°C for fifteen minutes to one hour. The desired volume of probe is pipeted into the hybridization tube, and the incubation is continued with rotation overnight at 60°C. The probe concentration in the hybridization solution should be 10 ng/ml if the specific activity is  $10^8$  dpm/µg or 2 ng/ml if the specific activity is  $10^9$  dpm/µg (use 2-10 ng/ml of Digoxigenin labeled probe).

25        The hybridization solution is poured off and an equal volume of 2×SSC/0.1% SDS is added. Incubation with rotation for 5 minutes at room temperature is carried out. The wash solution is changed, and this step is repeated. To reduce background, it may be beneficial to double the volume of the wash solutions. The wash solution is replaced with an equal volume

of 0.2×SSC/0.1% SDS and the tube is incubated for five minutes with rotation at room temperature. The wash solution is changed and this step is repeated (this is a low-stringency wash). For moderate or high stringency conditions, further washes are done with wash solutions pre-warmed to moderate (42°C) or high (68°C) temperatures. The final wash  
5 solution is removed and the membrane rinsed in 2×SSC at room temperature. Autoradiography is then performed for up to one week. Alternatively, signal is detected using chemiluminescence technology (Amersham) if non-radioactive probes are used. The transcript size is calculated from the signal from the gel in comparison with gel molecular weight standard markers.

10 Example 13: Tissue specific distribution of *NaN* by RT-PCR

*NaN*-specific forward (5' CCCTGCTGCGCTCGGTGAAGAA 3') (SEQ ID NO: 39) and reverse primer (5' GACAAAGTAGATCCCAGAGG 3') (SEQ ID NO: 25), were used in RT-PCR assays using cDNA template prepared from multiple rat. These primers amplify *NaN* sequence between nucleotides 765 and 1156 (392 bp) and are *NaN*-specific as judged by  
15 lack of similarity to sequences in the database (using programs like BLASTN from the National Center for Biotechnology Information). Amplification was typically performed in a 60 µl volume using 1 µl of the first strand of cDNA, 0.8 µM of each primer and 1.75 units of Expand Long Template DNA polymerase enzyme mixture (Boehringer Mannheim). Compared to conventional and thermostable DNA polymerases, Expand Long Template  
20 enzyme mixture increases the yield of the PCR products without an increase in non-specific amplification (Barnes, (1994) Proc. Natl. Acad. Sci. USA 91, 2216-2220; Cheng *et al.*, (1994) Proc. Natl. Acad. Sci. USA 91, 5695-5699). The PCR reaction buffer consisted of 50 mM Tris-HCl (pH 9.2), 16mM (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 2.25 mM MgCl<sub>2</sub>, 2% (v/v) DMSO and 0.1% Tween 20. As described previously (Dib-Hajj *et al.*, (1996) FEBS Lett. 384, 78-82), amplification was  
25 carried out in two stages using a programmable thermal cycler (PTC-200, MJ Research). First, a denaturation step is performed at 94°C for four minutes, followed by an annealing step at 60°C for two minutes, and then an elongation step at 72°C for ninety seconds. Second, a denaturation step is performed at 94°C for one minute, followed by an annealing step at 60°C

for one minute, and then an elongation step at 72°C for ninety seconds. The second stage was repeated 33 times for a total of 25-35 cycles, with the elongation step in the last cycle extended to ten minutes.

Glyceraldehyde-3-phosphate dehydrogenase (GAPDH) was used as an internal  
5 control to ensure that a lack of *NaN* signals in different tissues was not due to degraded templates or presence of PCR inhibitors. Rat GAPDH sequences were co-amplified using primers which amplify a 66 bp product that corresponds to nucleotides 328-994 (accession number: M17701). The amplified product spans multiple exon/intron splice sites, based on the structure of the human gene (Benham *et al.*, (1987) Nature 328, 275-278). DnaseI  
10 treatment was routinely performed prior to reverse transcription to prevent amplification of GAPDH sequences from genomic processed pseudogenes that may have contaminated the total RNA preparation (Ercolani *et al.*, (1988) J. Biol. Chem. 263, 15335-15341).

*NaN* is primarily and preferentially expressed in DRG and trigeminal ganglia neurons. Figure 4 shows the results of screening by RT-PCR for the expression of *NaN* in  
15 various neuronal and non-neuronal tissues. Lanes 1, 2, 4, 9 and 16 show a single amplification product co-migrating with the 400 bp marker, consistent with *NaN*-specific product of 392 bp. Lanes 1, 2, 4, 9 and 16 contain products using DRG, cerebral hemisphere, retina, and trigeminal ganglia, respectively. Using this assay, *NaN* was not detected in cerebellum, optic nerve, spinal cord, sciatic nerve, superior cervical ganglia, skeletal muscle,  
20 cardiac muscle, adrenal gland, uterus, liver or kidney (lanes 3, 5-8, and 10-15, respectively). The attenuated *NaN* signal in cerebral hemisphere and retina, and the absence of this signal in the remaining tissues is not due to degraded RNA or the presence of PCR inhibitors in the cDNA templates as comparable GAPDH amplification products were obtained in a parallel set of PCR reaction (data not shown).

25 **Example 14: Transformation of a Host Cell with the *NaN* Coding Sequence**

Transformed host cells for the measurement of Na<sup>+</sup> current or intracellular Na<sup>+</sup> levels are usually prepared by co-transfecting constructs into cells such as HEK293 cells with a fluorescent reporter plasmid (pGreen Lantern-1, Life Technologies, Inc.) using the

calcium-phosphate precipitation technique (Ukomadu *et al.*, (1992) Neuron 8, 663-676).

HEK293 cells are typically grown in high glucose DMEM (Life Technologies) supplemented with 10% fetal calf serum (Life Technologies). After 48 hours, cells with green fluorescence are selected for recording (Dib-Hajj *et al.*, (1997) FEBS Lett. 416, 11-14).

5 For preparation of cell lines continuously expressing recombinant channels, the *NaN* construct is cloned into other vectors that carry a selectable marker in mammalian cells. Transfections are carried out using the calcium phosphate precipitation technique (Ukomadu *et al.*, (1992) Neuron 8, 663-676). Human embryonic kidney (HEK-293), chinese hamster ovary (CHO) cells, or other suitable cell lines are grown under standard tissue culture  
10 conditions in Dulbeccos's modified Eagle's medium supplemented with 10% fetal bovine serum. The calcium phosphate-DNA mixture is added to the cell culture medium and left for fifteen to twenty hours, after which time the cells are washed with fresh medium. After forty-eight hours, antibiotic (G418) is added to select for cells which have acquired neomycin resistance. After two weeks in G418, 10-20 isolated cell colonies are harvested using sterile  
15 10ml pipette tips. Colonies are grown for another four to seven days, split and subsequently tested for channel expression using whole-cell patch-clamp recording techniques and RT-PCR.

#### Example 15: Production of NaN specific Antibodies

Antibodies specific to the rat, mouse or human NaN are produced with an  
20 immunogenic NaN-specific peptide by raising polyclonal antibodies in rabbits. In one example, the peptide **CGPNPASNKDCCFEKEKDSED** (rat amino acids 285-304) (SEQ ID NO: 40) was selected because it fits the criteria for immunogenecity and surface accessibility. This peptide sequence does not match any peptide in the public databases. The underlined cysteine (C) residue was changed to Alanine (A) to prevent disulfide bond formation. This  
25 amino acid change is not expected to significantly affect the specificity of the antibodies.

Peptide synthesis, rabbit immunization, and affinity purification of the antipeptide antibodies were performed using standard procedures. Purified antibodies were then tested on

DRG neurons in culture. Immunostaining procedures using these antibodies before and after blocking with excess peptide were performed according to standard procedures.

DRG neurons after sixteen to twenty-four hours in culture were processed for immunocytochemical detection of NaN protein as follows. Coverslips were washed with  
5 complete saline solution (137 mM NaCl, 5.3 mM KCl, 1 ITIM M902 25 mM sorbitol, 10 mM HEPES, 3 mM CaCl<sub>2</sub> (pH 7.2)), fixed with 4% paraformaldehyde in 0.14 M phosphate buffer for ten minutes at 4°C, washed with three five minutes with phosphate-buffered saline (PBS), and blocked with PBS containing 20% normal goat serum, 1% bovine serum albumin and 0.1  
10 % Triton X-100 for fifteen minutes. The coverslips were incubated in anti-NaN antibody (1:100 dilution) at 4°C overnight. Following overnight incubation, coverslips were washed extensively in PBS and then incubated with goat anti-rabbit IgG-conjugated to Cy3 (1:3000; Amersham) for two hours at room temperature. The coverslips were rinsed with PBS and mounted onto glass slides with Aqua-poly-mount. The neurons were examined with a Leitz Aristoplan light microscope equipped with epifluorescence and images were captured with a  
15 Dage DC330T color camera and Scion CG-7 color PCI frame grabber (see Figure 7).

**Example 16: NaN expression is altered in a neuropathic pain model**

The CCI model of neuropathic pain was used to study the plasticity of sodium channel expression in DRG neurons. Twenty two adult, female Sprague-Dawley rats, weighing 240-260 g were anesthetized with pentobarbital sodium (50 mg/kg ip) and the right sciatic nerve  
20 exposed at the mid-thigh. Four chromic gut (4-0) ligatures were tied loosely around the nerve as described by Bennett & Xie, (1988) Pain 33, 87-107. The incision site was closed in layers and a bacteriostatic agent administered intramuscularly.

Previous studies have shown that transection of the sciatic nerve induces dramatic changes in sodium currents of axotomized DRG neurons, which is paralleled by significant  
25 changes to transcripts of various sodium channels expressed in these neurons. Sodium currents that are TTX-R and the transcripts of two TTX-R sodium channels (SNS/PN3 and NaN) are significantly attenuated while a rapidly repriming silent TTX-S current emerges and the transcript of  $\alpha$ -III sodium channel, which produces a TTX-S current, is up-regulated. We



have discovered that CCI-induced changes in DRG neurons, fourteen days post-surgery, mirror those of axotomy. Transcripts of NaN and SNS, the two sensory neuron-specific TTX-R channels, are significantly down-regulated as is the TTX-R sodium current, while transcripts of the TTX-S  $\alpha$ -III sodium channel are up-regulated, in small diameter DRG neurons. These changes may be partly responsible for making DRG neurons hyperexcitable, that contributes to the hyperalgesia that results from this injury.

Example 17: Assays for agents which modulate the activity of the NaN channel using patch clamp methods

Cells lines expressing the cloned Na<sup>+</sup> channel are used to assay for agents which modulate the activity of the NaN channel, *e.g.*, agents which block or inhibit the channel or enhance channel opening. Since the channel activation is voltage dependent, depolarizing conditions may be used for observation of baseline activity that is modified by the agent to be tested. Depolarization may be achieved by any means available, for example, by raising the extracellular potassium ion concentration to about 20 to 40 nM, or by repeated electrical pulses.

The agent to be tested is incubated with HEK 293 or other transformed cells that express the Na<sup>+</sup> channel (Dib-Hajj *et al.*, (1997) FEBS Lett. 416, 11-14). After incubation for a sufficient period of time, the agent induced changes in Na<sup>+</sup> channel activity can be measured by patch-clamp methods (Hamill *et al.*, (1981) Pflügers Arch. 391, 85-100). Data for these measurements are acquired on a MacIntosh Quadra 950, or similar computer, using a program such as Pulse (v 7.52, HEKA, German). Fire-polished electrodes (0.8-1.5 MW) are fabricated from capillary glass using a Sutter P-87 puller or a similar instrument. Cells are usually only considered for analysis if initial seal resistance is <5 Gohm, they have high leakage currents (holding current <0.1 nA at -80 mV), membrane blebs, and an access resistance <5 Mohm. Access resistance is monitored and data is not used if resistance changes occur. Voltage errors are minimized using series resistance compensation and the capacitance artifact will be canceled as necessary using computer-controlled amplifier circuitry or other similar methods.

For comparisons of the voltage dependence of activation and inactivation, cells with a maximum voltage error of <10 mV after compensation are usually used. Linear leak subtraction is used for voltage clamp recordings. Membrane currents are typically filtered at 5 KHz and sampled at 20 KHz. The pipette solution contains a standard solution such as: 140 mM CsF, 2 mM MgCl<sub>2</sub>, 1 mM EGTA, and 10 mM Na-HEPES (pH 7.3). The standard bathing solution is a standard solution such as 140 mM NaCl, 3 mM KCl, 2 mM MgCl<sub>2</sub>, 1 mM CaCl<sub>2</sub>, 10 mM HEPES, and 10 mM glucose (pH 7.3).

Tetrodotoxin (TTX)-resistant and TTX-sensitive Na<sup>+</sup> currents are measured by exposure to appropriate concentrations of TTX and/or by pre-pulse protocols which distinguish between TTX-sensitive and TTX-resistant currents on the basis of their distinct steady-state inactivation properties (Cummins & Waxman (1997) J. Neurophysiol. 17, 3503-3514; Sontheimer & Waxman, (1992) J. Neurophysiol. 68, 1001-1011).

Data are collected using standard pulse protocols and are analyzed to measure sodium current properties that include voltage-dependence, steady-state characteristics, kinetics, and re-priming. Measurements of current amplitude and cell capacitance provides an estimate of Na<sup>+</sup> current density, thereby permitting comparisons of channel density under different conditions (Cummins & Waxman (1997) J. Neurophysiol. 17, 3503-3514,30). Cells are studied in the current clamp mode to study patterns of spontaneous and evoked action potential generation, threshold for firing, frequency response characteristics, and response to de- and hyperpolarization, and other aspects of electrogenesis (Sontheimer & Waxman, (1992) J. Neurophysiol. 68, 1001-1011). These measurements are carried out both in control cells expressing *NaV* and in cells expressing *NaN* that also have been exposed to the agent to be tested.

Example 18: Assays for agents which modulate the activity of the NaN channel by the measurement of Intracellular Sodium [Na<sup>+</sup>]

The agent to be tested is incubated with cells exhibiting NaN channel activity. After incubation for a sufficient period of time, the agent induced changes in Na<sup>+</sup> channel are measured by ratiometric imaging of [Na<sup>+</sup>]<sub>i</sub> using SBFI. In this method, cytosolic-free Na<sup>+</sup> is

measured using an indicator for  $\text{Na}^+$ , such as SBFI (sodium-binding benzofuran isophthalate (Harootunian *et al.*, (1989) J. Biol. Chem. 264, 19458-19467)); or a similar dye. Cells are first loaded with the membrane-permeable acetoxymethyl ester form of SBFI (SBFI/AM) or a similar dye (usually dissolved in dimethyl sulfoxide (DMSO) at a stock concentration of 10 mM). Recordings are obtained on the stage of a microscope using a commercially available ratiometric imaging setup (*e.g.*, from Georgia Instruments). Excitation light is provided at appropriate wavelengths (*e.g.*, 340:385 nm). Excitation light is passed to the cells through a dichroic reflector (400 nm) and emitted light above 450 nm was collected. Fluorescence signals are amplified, *e.g.*, by an image intensifier (GenIISyS) and collected with a CCD camera, or similar device, interfaced to a frame grabber. To account for fluorescence rundown, the fluorescence ratio 340:385 is used to assay cytosolic-free  $\text{Na}^+$ .

For calibration of SBFI's fluorescence, cells are perfused with calibration solutions containing known  $\text{Na}^+$  concentrations (typically 0 and 30 mM, or 0, 30, and 50 mM  $[\text{Na}^+]$ , and gramicidin and monensin. As reported by Rose and Ransom (Rose & Ransom, (1996) J. Physiol. (Lond) 491, 291-305), the 345/390 nm fluorescence ratio of intracellular SBFI changes monotonically with changes in  $[\text{Na}^+]$ . Experiments are repeated on multiple (typically at least four) different coverslips, providing statistically significant measurements of intracellular sodium in control cells, and in cells exposed to various concentrations of agents that may block, inhibit or enhance the activity of the channel. Use of this method is illustrated in Sontheimer *et al.*, (1994) J. Neurosci. 14, 2464-2475.

Example 19: Assays for agents which modulate the activity of the  $\text{Na}^+$  channel by scintigraphic imaging

Cells lines expressing the cloned  $\text{Na}^+$  channel are used to assay for agents which modulate the activity of the  $\text{Na}^+$  channel, *e.g.*, agents which block the channel or enhance channel opening. For example, the agent to be tested is incubated with HEK 293 or other transformed cells that express the  $\text{Na}^+$  channel (Dib-Hajj *et al.*, (1997) FEBS Lett. 416, 11-14). After incubation for a sufficient period of time, the agent induced changes in  $\text{Na}^+$  channel activity are detected by the measurement of  $\text{Na}^+$  influx by isotopic methods.  $^{22}\text{Na}$  is a

gamma emitter and can be used to measure  $\text{Na}^+$  flux (Kimelberg & Waltz, (1988) The Neuronal Microenvironment (Boulton *et al.*, editors) Humana Press) and  $^{86}\text{Rb}^+$  can be used to measure  $\text{Na}^+/\text{K}^+$ ATPase activity which provides a measure of Na channel activity (Sontheimer *et al.*, (1994) J. Neurosci. 14, 2464-2475)  $^{86}\text{Rb}^+$  ions are taken up by the  $\text{Na}^+/\text{K}^+$ ATPase like  
5  $\text{K}^+$  ions, but have the advantage of a much longer half-life than  $^{42}\text{K}^+$  (Kimelberg & Mayhew (1975) J. Biol. Chem. 250, 100-104). Thus, measurement of the unidirectional ouabain-sensitive  $^{86}\text{Rb}^+$  uptake provides a quantitative method for assaying  $\text{Na}^+/\text{K}^+$ -ATPase activity which follows action potentials.

Following incubation of cell expressing *NaN* to the isotope, the cellular content of the  
10 isotope is measured by liquid scintillation counting or a similar method, and cell protein is determined using a method such as the bicinchoninic acid protein assay (Smith *et al.*, (1985) Anal. Biochem. 150, 76-85) following the modifications (Goldschmidt & Kimelberg (1989) Anal. Biochem. 177, 41-45) for cultured cells.  $^{22}\text{Na}$  and  $^{86}\text{Rb}^+$  fluxes are determined in the presence and absence of agents that may block, inhibit, or enhance  $\text{Na}^+$ . This permits  
15 determination of the actions of these agents on *NaN*.

#### Example 20: In situ hybridization

##### a. Probes

Probes are prepared as described above in Example 5.

##### b. DRG Neuron Culture

20 Cultures of DRG neurons from adult rats were established as described previously (Rizzo *et al.*, (1994) J. Neurophysiol. 72, 2796-2815). Briefly, lumbar ganglia (L4, L5) from adult Sprague Dawley female rats were freed from their connective sheaths and incubated sequentially in enzyme solutions containing collagenase and then papain. The tissue was triturated in culture medium containing 1:1 Dulbecco's modified Eagle's medium (DMEM)  
25 and Hank's F12 medium and 10% fetal calf serum, 1.5 mg/ml trypsin inhibitor, 1.5 mg/ml bovine serum albumin, 100 units/ml penicillin and 0.1 mg/ml streptomycin and plated at a density of 500-1000 cells/mm<sup>2</sup> on polyornithine/laminin coated coverslips. The cells were

maintained at 37°C in a humidified 95% air/5% CO<sub>2</sub> incubator overnight and then processed for *in situ* hybridization cytochemistry as described previously (Black *et al.*, (1994) Brain Res. Mol. Brain Res. 23, 235-245; Zur *et al.*, (1995) Brain Res. Mol. Brain Res. 30, 97-105). Trigeminal ganglia can be cultured by a skilled artisan using similar methods.

5       c. Tissue Preparation

Adult female Sprague Dawley rats were deeply anesthetized, *e.g.*, with chloral hydrate and perfused through the heart, first with a phosphate-buffered saline (PBS) solution and then with a 4% paraformaldehyde in 0.14 M Sorensen's phosphate buffer (pH 7.4) at 4°C. Following perfusion fixation, dorsal root ganglia at levels L4 and L5 and trigeminal ganglia  
10 were collected and placed in fresh fixative at 4°C. After two to four hours, the tissue was transferred to a solution containing 4% paraformaldehyde and 30% sucrose in 0.14 M phosphate buffer and stored overnight at 4°C. Fifteen µm sections were cut and placed on poly-L-lysine-coated slides. The slides were processed for *in situ* hybridization cytochemistry as previously described (Waxman *et al.*, (1994) J. Neurophysiol. 72, 466-470; Black *et al.*,  
15 (1994) Brain Res. Mol. Brain Res. 23, 235-245). Following *in situ* hybridization cytochemistry, the slides were dehydrated, cleared and mounted with Permount. The results are shown in Figure 5.

Sections of DRG hybridized with *NaV* sense riboprobe showed no specific labeling (panel C, Figure 5). In DRG (panel A, Figure 5) and trigeminal (panel B) sections hybridized  
20 with a *NaV* antisense riboprobe, with the *NaV* signal present in most small (<30 µm diam.) neurons; in contrast, most large (>30 µm diam.) neurons did not exhibit *NaV* hybridization signal. Sections of spinal cord, cerebellum and liver hybridized with an antisense *NaV* riboprobe showed no specific signal (panels D, E and F respectively).

Example 21: Microsatellite Sequences

25       The following are the murine intronic microsatellite sequences. These microsatellites may be polymorphic in the human SCN11a gene and could be used as markers to screen for

AGTTTAATGTTGAGTGAATTGTGGTGGTGATTTCCCACTTGAGGCCTTTGTGTAA  
5 AGCCCAATGT  
GTGTGGTTGGGGGGTGGTGGCAGAGTCTGGTATTGGTAAGGTGAGAGCAATCCCA  
GAACGTCCACCTGCTCTTCCATTTTATTAATCAGGCAGGCCTCT

[illegible]

15 TGTGCATGCTTGATTCCCAGCTCCTATGGTCTGATTACTCGGTCCTTAGGAGCAAG  
GCCAGACTGTCCACCCTGACACACACACACACACACACACACACACACACAC  
ACACACACACACAGTGTAGAGAATTACCTCATTCTTGGAGTTTCTCTGGAAAAGG  
AATGTCTCAAAGCCAAGTTCACAGAGCAACAGCTG

[illegible]



It should be understood that the foregoing discussion and examples merely present a detailed description of certain preferred embodiments. It will be apparent to those of ordinary skill in the art that various modification and equivalents can be made without departing from the spirit and scope of the invention. The documents cited and referred to in this patent  
5 specification are hereby incorporated by reference in their entirety.



**CLAIMS**

1. An isolated nucleic acid molecule selected from the group consisting of a nucleic acid molecule comprising SEQ ID NO: 41, a nucleic acid molecule which encodes the amino acid sequence of SEQ ID NO: 42, a nucleic acid molecule which encodes an allelic variant of  
5 SEQ ID NO:42, a nucleic acid molecule which encodes a human protein exhibiting at least about 76% amino acid sequence identity to SEQ ID NO:42 and a nucleic acid molecule that hybridizes to one of the foregoing sequences under stringent conditions.
2. The isolated nucleic acid of claim 1, wherein the nucleic acid encodes a voltage gated Na<sup>+</sup> channel that is preferentially expressed in dorsal root ganglia or trigeminal ganglia.
3. The isolated nucleic acid of claim 2, wherein the nucleic acid encodes the human NaN sodium channel.
4. An expression vector comprising the isolated nucleic acid of any one of claims 1 to 3, alone or together with appropriate regulatory and expression control elements.
5. A host cell transformed with the expression vector of claim 4.
6. A Na<sup>+</sup> channel encoded by an isolated nucleic acid molecule of any one of claims 1 to 3.
7. The Na<sup>+</sup> channel of claim 6, comprising the amino acid sequence of SEQ ID NO:42.
8. An isolated protein consisting of the amino acid sequence of SEQ ID NO: 42 or a peptide fragment thereof.

9. A protein comprised within a membrane fragment isolated from the host cell of claim 5.

10. A method to identify an agent that modulates the activity of the Na<sup>+</sup> channel of claim 6, comprising the steps of bringing the agent into contact with a cell that expresses the Na<sup>+</sup> channel on its surface and measuring any resultant changes in the sodium current, resultant change in membrane potential or change in intracellular Na<sup>+</sup>.

11. The method of claim 10, wherein the measuring step is accomplished by voltage clamp measurements or measurement of membrane potential.

12. The method of claim 10, wherein the measuring step is accomplished by measuring the level of intracellular sodium.

13. The method of claim 10, wherein the measuring step is accomplished by measuring sodium influx.

14. The method of claim 13, wherein the sodium influx is measured using <sup>22</sup>Na or <sup>86</sup>Rb.

15. A method to identify an agent that modulates the transcription or translation of mRNA encoding the Na<sup>+</sup> channel of claim 6, comprising the steps of bringing the agent into contact with a cell that expresses the Na<sup>+</sup> channel and measuring the resultant level of expression of the Na<sup>+</sup> channel.

16. A method to treat pain, paraesthesia and/or hyperexcitability phenomena in an animal or human subject by administering an effective amount of an agent that alters Na<sup>+</sup> current flow through Na<sup>+</sup> channels in DRG or trigeminal neurons.

17. A method to treat pain, paraesthesia and/or hyperexcitability phenomena in an animal or human subject by administering an effective amount of an agent that modulates the transcription or translation of mRNA encoding the Na<sup>+</sup> channel of claim 6.

18. An isolated nucleic acid that is antisense to the nucleic acid of claim 1 and of sufficient length to modulate the expression of NaN channel in a cell containing the mRNA.

19. A scintigraphic method to image the loci of pain generation or provide a measure of the level of pain associated with DRG or trigeminal neuron mediated hyperexcitability in an animal or human subject by administering labeled monoclonal antibodies or other labeled ligands specific for the human NaN Na<sup>+</sup> channel.

20. A method to identify tissues, cells or cell types that express the human NaN sodium channel, comprising the step of detecting human NaN on the cell surface or intracellularly.

21. A method to identify tissues, cells or cell types that express human NaN comprising the step of detecting the presence therein of human NaN encoding mRNA.

22. A method of producing a transformed cell that expresses an exogenous NaN encoding nucleic acid, comprising the step of transforming the cell with an expression vector comprising the isolated nucleic acid of any of claims 1 to 3, together with appropriate regulatory or expression control elements.

23. An isolated antibody specific for the human NaN channel or polypeptide fragment thereof.

24. The isolated antibody of claim 23, wherein the antibody is labeled.

25. A method of producing recombinant NaN protein, comprising the step of culturing the transformed host of claim 5 under conditions in which the NaN sodium channel or protein is expressed.

26. A therapeutic composition comprising an effective amount of an agent capable of altering, such as by increasing or decreasing, the rapidly repriming current flow in axotomized, inflamed or otherwise injured DRG neurons.

27. A method to treat acute pain or acute or chronic neuropathic or inflammatory pain and hyperexcitability phenomena in an animal or a human patient by administering the therapeutic composition of claim 26.

28. A method to screen candidate compounds for use in treating pain and hyperexcitability phenomena by testing their ability to upregulate or downregulate the NaN channel mRNA in axotomized, inflamed or otherwise injured DRG neurons.

29. A chimeric NaN channel.

30. A chimeric channel of claim 29, wherein at least one human domain has been substituted with the corresponding domain from the NaN channel of another species.

31. A chimeric channel of claim 30, wherein the species is rat or mouse.

32. A nucleic acid molecule encoding a chimeric NaN channel of any one of claims 29 through 31.

33. An NaN channel protein comprising a positively charged amino acid at a position corresponding to residue 670 of SEQ ID NO:42.

34. An NaN channel protein of claim 33, wherein the positively charged amino acid is arginine.

35. An isolated nucleic acid molecule encoding a channel protein of either of claims 33 or 34.

36. A therapeutic composition of claim 26, further comprising at least one second agent that modulates a channel in primary sensory neurons.

37. A therapeutic composition of claim 36, wherein the composition comprises agents which modulate NaN and at least one channel selected from the group consisting of PN1/hNE and SNS/PN3.

## SEQUENCE LISTING

<110> Yale University  
 Dib-Hajj, Sulayman  
 Waxman, Stephen G.

<120> Modulation of Sodium Channels in Dorsal Root Ganglia

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<150> US 09/354,147

<151> 1999-07-16

<160> 44

<170> PatentIn Ver. 2.1

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<222> (41)..(5335)

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<221> unsure

<222> (1996)..(4042)

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Lys Lys Ser Lys Asp Lys Ala Ala Ala Glu Pro Gln Pro Arg Pro Gln
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Leu Asp Leu Lys Ala Ser Arg Lys Leu Pro Lys Leu Tyr Gly Asp Ile
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7

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&lt;210&gt; 2

&lt;211&gt; 1765

&lt;212&gt; PRT

&lt;213&gt; Rattus norvegicus

&lt;400&gt; 2

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His His Asn Met Asp Asp Asn Leu Lys Thr Ile Leu Lys Ile Gly Asn  
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 Leu Cys Gly Glu Trp Ile Glu Asn Met Trp Gly Cys Met Gln Asp Met  
 755 760 765  
 Asp Gly Ser Pro Leu Cys Ile Ile Val Phe Val Leu Ile Met Val Ile  
 770 775 780  
 Gly Lys Leu Val Val Leu Asn Leu Phe Ile Ala Leu Leu Leu Asn Ser  
 785 790 795 800  
 Phe Ser Asn Glu Glu Lys Asp Gly Ser Leu Glu Gly Glu Thr Arg Lys  
 805 810 815  
 Thr Lys Val Gln Leu Ala Leu Asp Arg Phe Arg Arg Ala Phe Ser Phe  
 820 825 830  
 Met Leu His Ala Leu Gln Ser Phe Cys Cys Lys Lys Cys Arg Arg Lys  
 835 840 845  
 Asn Ser Pro Lys Pro Lys Glu Thr Thr Glu Ser Phe Ala Gly Glu Asn  
 850 855 860  
 Lys Asp Ser Ile Leu Pro Asp Ala Arg Pro Trp Lys Glu Tyr Asp Thr  
 865 870 875 880  
 Asp Met Ala Leu Tyr Thr Gly Gln Ala Gly Ala Pro Leu Ala Pro Leu  
 885 890 895

Ala Glu Val Glu Asp Asp Val Glu Tyr Cys Gly Glu Gly Gly Ala Leu  
                     900                    905                    910

Pro Thr Ser Gln His Ser Ala Gly Val Gln Ala Gly Asp Leu Pro Pro  
                     915                    920                    925

Glu Thr Lys Gln Leu Thr Ser Pro Asp Asp Gln Gly Val Glu Met Glu  
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Val Phe Ser Glu Glu Asp Leu His Leu Ser Ile Gln Ser Pro Arg Lys  
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Lys Ser Asp Ala Val Ser Met Leu Ser Glu Cys Ser Thr Ile Asp Leu  
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Asn Asp Ile Phe Arg Asn Leu Gln Lys Thr Val Ser Pro Lys Lys Gln  
                     980                    985                    990

Pro Asp Arg Cys Phe Pro Lys Gly Leu Ser Cys His Phe Leu Cys His  
                     995                    1000                    1005

Lys Thr Asp Lys Arg Lys Ser Pro Trp Val Leu Trp Trp Asn Ile Arg  
                     1010                    1015                    1020

Lys Thr Cys Tyr Gln Ile Val Lys His Ser Trp Phe Glu Ser Phe Ile  
                     1025                    1030                    1035                    1040

Ile Phe Val Ile Leu Leu Ser Ser Gly Ala Leu Ile Phe Glu Asp Val  
                     1045                    1050                    1055

Asn Leu Pro Ser Arg Pro Gln Val Glu Lys Leu Leu Arg Cys Thr Asp  
                     1060                    1065                    1070

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                     1075                    1080                    1085

Ala Phe Gly Phe Arg Arg Tyr Phe Thr Ser Ala Trp Cys Trp Leu Asp  
                     1090                    1095                    1100

Phe Leu Ile Val Val Val Ser Val Leu Ser Leu Met Asn Leu Pro Ser  
                     1105                    1110                    1115                    1120

Leu Lys Ser Phe Arg Thr Leu Arg Ala Leu Arg Pro Leu Arg Ala Leu  
                     1125                    1130                    1135

Ser Gln Phe Glu Gly Met Lys Val Val Val Tyr Ala Leu Ile Ser Ala  
                     1140                    1145                    1150

Ile Pro Ala Ile Leu Asn Val Leu Leu Val Cys Leu Ile Phe Trp Leu  
                     1155                    1160                    1165

Val Phe Cys Ile Leu Gly Val Asn Leu Phe Ser Gly Lys Phe Gly Arg  
                     1170                    1175                    1180

Cys Ile Asn Gly Thr Asp Ile Asn Met Tyr Leu Asp Phe Thr Glu Val  
                     1185                    1190                    1195                    1200

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Ser Gly Ile Asp Asp Ile Phe Asn Phe Glu Thr Phe Thr Gly Ser Met  
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 <212> PRT

&lt;213&gt; Rattus norvegicus

&lt;220&gt;

&lt;223&gt; putative amino acid seq. of rat NaN

&lt;400&gt; 3

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Ile Gln Lys Glu Arg Lys Lys Ser Lys Asp Lys Ala Ala Ala Glu Pro  
 35 40 45

Gln Pro Arg Pro Gln Leu Asp Leu Lys Ala Ser Arg Lys Leu Pro Lys  
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Leu Tyr Gly Asp Ile Pro Pro Glu Leu Val Ala Lys Pro Leu Glu Asp  
 65 70 75 80

Leu Asp Pro Phe Tyr Lys Asp His Lys Thr Phe Met Val Leu Asn Lys  
 85 90 95

Lys Arg Thr Ile Tyr Arg Phe Ser Ala Lys Arg Ala Leu Phe Ile Leu  
 100 105 110

Gly Pro Phe Asn Pro Leu Arg Ser Leu Met Ile Arg Ile Ser Val His  
 115 120 125

Ser Val Phe Ser Met Phe Ile Ile Cys Thr Val Ile Ile Asn Cys Met  
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Phe Met Ala Asn Ser Met Glu Arg Ser Phe Asp Asn Asp Ile Pro Glu  
 145 150 155 160

Tyr Val Phe Ile Gly Ile Tyr Ile Leu Glu Ala Val Ile Lys Ile Leu  
 165 170 175

Ala Arg Gly Phe Ile Val Asp Glu Phe Ser Phe Leu Arg Asp Pro Trp  
 180 185 190

Asn Trp Leu Asp Phe Ile Val Ile Gly Thr Ala Ile Ala Thr Cys Phe  
 195 200 205

Pro Gly Ser Gln Val Asn Leu Ser Ala Leu Arg Thr Phe Arg Val Phe  
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Arg Ala Leu Lys Ala Ile Ser Val Ile Ser Gly Leu Lys Val Ile Val  
 225 230 235 240

Gly Ala Leu Leu Arg Ser Val Lys Lys Leu Val Asp Val Met Val Leu  
 245 250 255

Thr Leu Phe Cys Leu Ser Ile Phe Ala Leu Val Gly Gln Gln Leu Phe  
 260 265 270

Met Gly Ile Leu Asn Gln Lys Cys Ile Lys His Asn Cys Gly Pro Asn  
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 Pro Ala Ser Asn Lys Asp Cys Phe Glu Lys Glu Lys Asp Ser Glu Asp  
 290 295 300  
 Phe Ile Met Cys Gly Thr Trp Leu Gly Ser Arg Pro Cys Pro Asn Gly  
 305 310 315 320  
 Ser Thr Cys Asp Lys Thr Thr Leu Asn Pro Asp Asn Asn Tyr Thr Lys  
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 485 490 495  
 Ser Gln Asn Leu Pro Val Asp Leu Phe Asp Glu His Val Asp Pro Leu  
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 His Arg Gln Arg Ala Leu Ser Ala Val Ser Ile Leu Thr Ile Thr Met  
 515 520 525  
 Gln Glu Gln Glu Lys Phe Gln Glu Pro Cys Phe Pro Cys Gly Lys Asn  
 530 535 540  
 Leu Ala Ser Lys Tyr Leu Val Trp Asp Cys Ser Pro Gln Trp Leu Cys  
 545 550 555 560  
 Ile Lys Lys Val Leu Arg Thr Ile Met Thr Asp Pro Phe Thr Glu Leu  
 565 570 575

Ala Ile Thr Ile Cys Ile Ile Ile Asn Thr Val Phe Leu Ala Val Glu  
 580 585 590  
 His His Asn Met Asp Asp Asn Leu Lys Thr Ile Leu Lys Ile Gly Asn  
 595 600 605  
 Trp Val Phe Thr Gly Ile Phe Ile Ala Glu Met Cys Leu Lys Ile Ile  
 610 615 620  
 Ala Leu Asp Pro Tyr His Tyr Phe Arg His Gly Trp Asn Val Phe Asp  
 625 630 635 640  
 Ser Ile Val Ala Leu Leu Ser Leu Ala Asp Val Leu Tyr Asn Thr Leu  
 645 650 655  
 Ser Asp Asn Asn Arg Ser Phe Leu Ala Ser Leu Arg Val Leu Arg Val  
 660 665 670  
 Phe Lys Leu Ala Lys Ser Trp Pro Thr Leu Asn Thr Leu Ile Lys Ile  
 675 680 685  
 Ile Gly His Ser Val Gly Ala Leu Gly Asn Leu Thr Val Val Leu Thr  
 690 695 700  
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 Trp His Met Asp Asn Phe Tyr His Ser Phe Leu Val Val Phe Arg Ile  
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 Leu Cys Gly Glu Trp Ile Glu Asn Met Trp Gly Cys Met Gln Asp Met  
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 Asp Gly Ser Pro Leu Cys Ile Ile Val Phe Val Leu Ile Met Val Ile  
 770 775 780  
 Gly Lys Leu Val Val Leu Asn Leu Phe Ile Ala Leu Leu Leu Asn Ser  
 785 790 795 800  
 Phe Ser Asn Glu Glu Lys Asp Gly Ser Leu Glu Gly Glu Thr Arg Lys  
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 Thr Lys Val Gln Leu Ala Leu Asp Arg Phe Arg Arg Ala Phe Ser Phe  
 820 825 830  
 Met Leu His Ala Leu Gln Ser Phe Cys Cys Lys Lys Cys Arg Arg Lys  
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 865 870 875 880

Asp Met Ala Leu Tyr Thr Gly Gln Ala Gly Ala Pro Leu Ala Pro Leu  
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 1170 1175 1180



Cys Ile Asn Gly Thr Asp Ile Asn Met Tyr Leu Asp Phe Thr Glu Val  
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 Pro Asn Arg Ser Gln Cys Asn Ile Ser Asn Tyr Ser Trp Lys Val Pro  
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&lt;211&gt; 5822

&lt;212&gt; DNA

&lt;213&gt; Mus musculus

&lt;220&gt;

&lt;221&gt; CDS

&lt;222&gt; (19) .. (5313)

&lt;220&gt;

&lt;221&gt; unsure

&lt;222&gt; (5804)

&lt;223&gt; cDNA sequence of mouse NaN, n = a or c or g or t

&lt;400&gt; 4

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Glu Lys Arg Ile Thr Ile Gln Lys Glu Lys Lys Lys Ser Lys Asp Lys
      30             35             40

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Ala Ala Thr Glu Pro Gln Pro Arg Pro Gln Leu Asp Leu Lys Ala Ser
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Arg Lys Leu Pro Lys Leu Tyr Gly Asp Val Pro Pro Asp Leu Ile Ala
      60             65             70             75

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Lys Pro Leu Glu Asp Leu Asp Pro Phe Tyr Lys Asp His Lys Thr Phe
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Met Val Leu Asn Lys Lys Arg Thr Ile Tyr Arg Phe Ser Ala Lys Arg
      95             100            105

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Ala Leu Phe Ile Leu Gly Pro Phe Asn Pro Ile Arg Ser Phe Met Ile
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cgc atc tct gtc cat tca gtc ttc agc atg ttc att atc tgc aca gtg 435
Arg Ile Ser Val His Ser Val Phe Ser Met Phe Ile Ile Cys Thr Val
      125            130            135

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Ile Ile Asn Cys Met Phe Met Ala Asn Asn Ser Ser Val Asp Ser Arg
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cct agc agt aac att ccc gaa tac gtc ttc att ggg att tat gtt tta 531
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Thr Glu Ala Lys Glu Lys Met Phe Gln Glu Ala Gln Gln Leu Leu Arg				
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Leu Leu Glu Gln Thr Lys Arg Leu Ser Gln Asn Leu Pro Val Glu Leu				
	495	500	505	
ttt gat gag cac gtg gac ccc ctc cat agg cag aga gcg ctg agt gcc				1587
Phe Asp Glu His Val Asp Pro Leu His Arg Gln Arg Ala Leu Ser Ala				
	510	515	520	
gtc agt atc tta acc atc acc atg cag gaa caa gaa aaa tcc cag gag				1635
Val Ser Ile Leu Thr Ile Thr Met Gln Glu Gln Glu Lys Ser Gln Glu				
	525	530	535	
cct tgt ttc ccg tgt ggg aaa aac ttg gca tcc aag tac ctg gtg tgg				1683
Pro Cys Phe Pro Cys Gly Lys Asn Leu Ala Ser Lys Tyr Leu Val Trp				
	540	545	550	555
gaa tgt agc cct ccg tgg ctg tgc ata aag aag gtc ctg cag act atc				1731
Glu Cys Ser Pro Pro Trp Leu Cys Ile Lys Lys Val Leu Gln Thr Ile				
	560	565	570	
atg aca gac ccc ttc act gag ctg gcc atc acc atc tgc atc atc gtc				1779
Met Thr Asp Pro Phe Thr Glu Leu Ala Ile Thr Ile Cys Ile Ile Val				
	575	580	585	
aat act gtc ttc ttg gcc atg gaa cac cac aat atg gat aac tct tta				1827
Asn Thr Val Phe Leu Ala Met Glu His His Asn Met Asp Asn Ser Leu				
	590	595	600	
aaa gac ata ctg aaa ata gga aac tgg gtt ttc act gga att ttc ata				1875
Lys Asp Ile Leu Lys Ile Gly Asn Trp Val Phe Thr Gly Ile Phe Ile				

605	610	615	
gcg gaa atg tgt ctc aag atc att gcg cta gac cct tac cac tac ttc			1923
Ala Glu Met Cys Leu Lys Ile Ile Ala Leu Asp Pro Tyr His Tyr Phe			
620	625	630	635
cgg cac ggc tgg aac atc ttt gac agc att gtg gcc ctt gtg agt ctc			1971
Arg His Gly Trp Asn Ile Phe Asp Ser Ile Val Ala Leu Val Ser Leu			
640	645		650
gct gac gtg ctc ttc cac aaa ctg tct aaa aac ctc tcc ttc ttg gct			2019
Ala Asp Val Leu Phe His Lys Leu Ser Lys Asn Leu Ser Phe Leu Ala			
655	660		665
tcc ctc aga gtg ctg agg gtc ttc aag tta gcc aaa tcc tgg ccc aca			2067
Ser Leu Arg Val Leu Arg Val Phe Lys Leu Ala Lys Ser Trp Pro Thr			
670	675		680
tta aac act ctc att aag atc atc ggc cac tcc gtg ggt gcg ctc gga			2115
Leu Asn Thr Leu Ile Lys Ile Ile Gly His Ser Val Gly Ala Leu Gly			
685	690		695
aac ctg act gtg gtc cta acg atc gtg gtc ttc atc ttt tcc gtg gtt			2163
Asn Leu Thr Val Val Leu Thr Ile Val Val Phe Ile Phe Ser Val Val			
700	705	710	715
ggc atg cgg ctc ttt ggt gcc aag ttt aac aag act tgc tcc acc tct			2211
Gly Met Arg Leu Phe Gly Ala Lys Phe Asn Lys Thr Cys Ser Thr Ser			
720	725		730
ccg gag tcc ctc cgg cgc tgg cac atg ggt gat ttc tac cat tcc ttc			2259
Pro Glu Ser Leu Arg Arg Trp His Met Gly Asp Phe Tyr His Ser Phe			
735	740		745
ctg gtg gtg ttc cgc atc ctc tgt ggg gag tgg atc gag aac atg tgg			2307
Leu Val Val Phe Arg Ile Leu Cys Gly Glu Trp Ile Glu Asn Met Trp			
750	755		760
gaa tgc atg cag gag atg gaa ggc tcc ccg ctg tgt gtc atc gtc ttt			2355
Glu Cys Met Gln Glu Met Glu Gly Ser Pro Leu Cys Val Ile Val Phe			
765	770		775
gtg ctg atc atg gtg gtc ggg aag ctc gtg gtg ctt aac ctc ttc att			2403
Val Leu Ile Met Val Val Gly Lys Leu Val Val Leu Asn Leu Phe Ile			
780	785	790	795
gcc ttg ctg ctc aat tcc ttc agc aat gag gaa aag gat ggg aac cca			2451
Ala Leu Leu Leu Asn Ser Phe Ser Asn Glu Glu Lys Asp Gly Asn Pro			
800	805		810
gaa gga gag acc agg aaa acc aaa gtg cag cta gcc ctg gat cgg ttc			2499
Glu Gly Glu Thr Arg Lys Thr Lys Val Gln Leu Ala Leu Asp Arg Phe			
815	820		825
agc cga gcg ttc tac ttc atg gcg cgc gct ctt cag aat ttc tgt tgc			2547
Ser Arg Ala Phe Tyr Phe Met Ala Arg Ala Leu Gln Asn Phe Cys Cys			

830	835	840	
aag aga tgc agg agg caa aac tgc cca aag cca aat gag gca aca gaa Lys Arg Cys Arg Arg Gln Asn Ser Pro Lys Pro Asn Glu Ala Thr Glu 845 850 855			2595
agc ttt gct ggt gag agt aga gac aca gcc acc ctg gat aca agg tcc Ser Phe Ala Gly Glu Ser Arg Asp Thr Ala Thr Leu Asp Thr Arg Ser 860 865 870 875			2643
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gct cca ctg gcc cca ctg gca aaa gaa gag gac gat atg gaa tgt tgt Ala Pro Leu Ala Pro Leu Ala Lys Glu Glu Asp Asp Met Glu Cys Cys 895 900 905			2739
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gcc tgt gac ctc cct ctg aag acc aag cgg ctc ccc agc cca gat gac Ala Cys Asp Leu Pro Leu Lys Thr Lys Arg Leu Pro Ser Pro Asp Asp 925 930 935			2835
cac ggg gtt gaa atg gaa gtg ttt tcc gaa gaa gat ccg aat tta acc His Gly Val Glu Met Glu Val Phe Ser Glu Glu Asp Pro Asn Leu Thr 940 945 950 955			2883
ata cag agt gct cga aag aag tct gat gcg gca agc atg ctc tca gaa Ile Gln Ser Ala Arg Lys Lys Ser Asp Ala Ala Ser Met Leu Ser Glu 960 965 970			2931
tgc agc aca ata gac ctg aat gat atc ttt aga aat tta cag aaa aca Cys Ser Thr Ile Asp Leu Asn Asp Ile Phe Arg Asn Leu Gln Lys Thr 975 980 985			2979
gtt tcc ccc caa aag caa cca gat cga tgc ttt ccc aag ggc ctc agt Val Ser Pro Gln Lys Gln Pro Asp Arg Cys Phe Pro Lys Gly Leu Ser 990 995 1000			3027
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tgg ttt gag agc ttc ata att ttt gtc atc ctg ctg agc agc gga gca Trp Phe Glu Ser Phe Ile Ile Phe Val Ile Leu Leu Ser Ser Gly Ala 1040 1045 1050			3171
ctg ata ttc gaa gat gtc aat ctt ccc agc cgg ccc caa gtt gaa aaa Leu Ile Phe Glu Asp Val Asn Leu Pro Ser Arg Pro Gln Val Glu Lys			3219

1055	1060	1065	
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atg att ttg aag tgg gtg gcc ttt gga ttc cgg aag tat ttc acc agt Met Ile Leu Lys Trp Val Ala Phe Gly Phe Arg Lys Tyr Phe Thr Ser 1085	1090	1095	3315
gcc tgg tgc tgg ctc gat ttc ctc att gtg gtg gtg tct gtg ctc agc Ala Trp Cys Trp Leu Asp Phe Leu Ile Val Val Val Ser Val Leu Ser 1100	1105	1110	3363
ctc acg aac tta cca aac ttg aag tcc ttc cgg aat ctg cga gcg ctg Leu Thr Asn Leu Pro Asn Leu Lys Ser Phe Arg Asn Leu Arg Ala Leu 1120	1125	1130	3411
aga cct ctg cgg gca ctg tct cag ttt gaa gga atg aag gtt gtt gtc Arg Pro Leu Arg Ala Leu Ser Gln Phe Glu Gly Met Lys Val Val Val 1135	1140	1145	3459
aat gcc ctc atg agt gcc ata cct gcc atc ctc aat gtc ttg ctg gtc Asn Ala Leu Met Ser Ala Ile Pro Ala Ile Leu Asn Val Leu Leu Val 1150	1155	1160	3507
tgc ctc att ttc tgg ctc ata ttt tgt atc ctg gga gta aat ttt ttt Cys Leu Ile Phe Trp Leu Ile Phe Cys Ile Leu Gly Val Asn Phe Phe 1165	1170	1175	3555
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Phe Asn Gln Gln Gln Lys Lys Leu Gly Gly Gln Asp Ile Phe Met Thr			
1295	1300	1305	
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Glu Glu Gln Lys Lys Tyr Tyr Asn Ala Met Lys Lys Leu Gly Thr Lys			
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aag cct caa aag ccc atc cca agg ccc ctg aac aaa tgt caa gcc ttc			4035
Lys Pro Gln Lys Pro Ile Pro Arg Pro Leu Asn Lys Cys Gln Ala Phe			
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Val Phe Asp Leu Val Thr Ser Gln Val Phe Asp Val Ile Ile Leu Gly			
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ctt att gtc aca aac atg att atc atg atg gct gaa tct gaa ggc cag			4131
Leu Ile Val Thr Asn Met Ile Ile Met Met Ala Glu Ser Glu Gly Gln			
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ccc aac gaa gtg aag aaa atc ttt gat att ctc aac ata gtc ttc gtg			4179
Pro Asn Glu Val Lys Lys Ile Phe Asp Ile Leu Asn Ile Val Phe Val			
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Val Ile Phe Thr Val Glu Cys Leu Ile Lys Val Phe Ala Leu Arg Gln			
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cac tac ttc acc aat ggc tgg aac tta ttt gat tgt gtg gtc gtg gtt			4275
His Tyr Phe Thr Asn Gly Trp Asn Leu Phe Asp Cys Val Val Val Val			
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Leu Ser Ile Ile Ser Thr Leu Val Ser Gly Leu Glu Asn Ser Asn Val			
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ttc ccg ccc aca ctc ttc agg att gtc cgc ttg gct cgg atc ggt cga			4371
Phe Pro Pro Thr Leu Phe Arg Ile Val Arg Leu Ala Arg Ile Gly Arg			
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Ile Leu Arg Leu Val Arg Ala Ala Arg Gly Ile Arg Thr Leu Leu Phe			
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gcg ttg atg atg tct ctc ccc tct ctc ttc aac att ggt ctg ctt ctc			4467
Ala Leu Met Met Ser Leu Pro Ser Leu Phe Asn Ile Gly Leu Leu Leu			
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Phe Leu Val Met Phe Ile Tyr Ala Ile Phe Gly Met Asn Trp Phe Ser			
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Lys Val Lys Arg Gly Ser Gly Ile Asp Asp Ile Phe Asn Phe Asp Thr			

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Trp Asp Ala Leu Leu Asn Pro Met Leu Glu Ser Lys Ala Ser Cys Asn				
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Ser Ser Ser Gln Glu Ser Cys Gln Gln Pro Gln Ile Ala Ile Val Tyr				
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Asp Pro Leu Gly Glu Asp Asp Phe Glu Ile Phe Tyr Glu Ile Trp Glu				
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Lys Phe Asp Pro Glu Ala Thr Gln Phe Ile Gln Tyr Ser Ser Leu Ser				
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Asp Phe Ala Asp Ala Leu Pro Glu Pro Leu Arg Val Ala Lys Pro Asn				
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Arg Phe Gln Phe Leu Met Met Asp Leu Pro Met Val Met Gly Asp Arg				
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Leu His Cys Met Asp Val Leu Phe Ala Phe Thr Thr Arg Val Leu Gly				
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Asn Ser Ser Gly Leu Asp Thr Met Lys Ala Met Met Glu Glu Lys Phe				
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Met Glu Ala Asn Pro Phe Lys Lys Leu Tyr Glu Pro Ile Val Thr Thr				
1695	1700	1705		
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Thr Lys Arg Lys Glu Glu Glu Cys Ala Ala Val Ile Gln Arg Ala				
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Tyr Arg Arg His Met Glu Lys Met Ile Lys Leu Lys Leu Lys Gly Arg				

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 Ile Gln Lys Glu Lys Lys Lys Ser Lys Asp Lys Ala Ala Thr Glu Pro  
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 Gln Pro Arg Pro Gln Leu Asp Leu Lys Ala Ser Arg Lys Leu Pro Lys  
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 Leu Tyr Gly Asp Val Pro Pro Asp Leu Ile Ala Lys Pro Leu Glu Asp  
 65                      70                      75                      80  
 Leu Asp Pro Phe Tyr Lys Asp His Lys Thr Phe Met Val Leu Asn Lys  
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 Lys Arg Thr Ile Tyr Arg Phe Ser Ala Lys Arg Ala Leu Phe Ile Leu  
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Gly Pro Phe Asn Pro Ile Arg Ser Phe Met Ile Arg Ile Ser Val His  
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 Arg Val Leu Arg Ala Leu Lys Ala Ile Ser Val Ile Ser Gly Leu Lys  
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Lys Met Phe Gln Glu Ala Gln Gln Leu Leu Arg Glu Glu Lys Glu Ala  
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Lys Asn Val Ala Ala Glu Ile Glu Ala Lys Glu Lys Met Phe Gln Glu	
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Thr Cys Tyr Gln Ile Val Lys His Ser Trp Phe Glu Ser Phe Ile Ile	
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 820 825 830  
 Val Asp Lys Arg Lys Pro Pro Trp Val Ile Trp Trp Asn Leu Arg Lys  
 835 840 845  
 Thr Cys Tyr Gln Ile Val Lys His Ser Trp Phe Glu Ser Phe Ile Ile  
 850 855 860  
 Phe Val Ile Leu Leu Ser Ser Gly Ala Leu Ile Phe Glu Asp Val His  
 865 870 875 880  
 Leu Glu Asn Gln Pro Lys Ile Gln Glu Leu Leu Asn Cys Thr Asp Ile  
 885 890 895  
 Ile Phe Thr His Ile Phe Ile Leu Glu Met Val Leu Lys Trp Val Ala  
 900 905 910  
 Phe Gly Phe Gly Lys Tyr Phe Thr Ser Ala Trp Cys Cys Leu Asp Phe  
 915 920 925  
 Ile Ile Val Ile Val Ser Val Thr Thr Leu Ile Asn Leu Met Glu Leu  
 930 935 940  
 Lys Ser Phe Arg Thr Leu Arg Ala Leu Arg Pro Leu Arg Ala Leu Ser  
 945 950 955 960  
 Gln Phe Glu Gly Met Lys Val Val Val Asn Ala Leu Ile Gly Ala Ile  
 965 970 975  
 Pro Ala Ile Leu Asn Val Leu Leu Val Cys Leu Ile Phe Trp Leu Val  
 980 985 990  
 Phe Cys Ile Leu Gly Val Tyr Phe Phe Ser Gly Lys Phe Gly Lys Cys  
 995 1000 1005  
 Ile Asn Gly Thr Asp Ser Val Ile Asn Tyr Thr Ile Ile Thr Asn Lys  
 1010 1015 1020  
 Ser Gln Cys Glu Ser Gly Asn Phe Ser Trp Ile Asn Gln Lys Val Asn  
 1025 1030 1035 1040  
 Phe Asp Asn Val Gly Asn Ala Tyr Leu Ala Leu Leu Gln Val Ala Thr  
 1045 1050 1055  
 Phe Lys Gly Trp Met Asp Ile Ile Tyr Ala Ala Val Asp Ser Thr Glu  
 1060 1065 1070

Lys Glu Gln Gln Pro Glu Phe Glu Ser Asn Ser Leu Gly Tyr Ile Tyr  
           1075                          1080                          1085  
  
 Phe Val Val Phe Ile Ile Phe Gly Ser Phe Phe Thr Leu Asn Leu Phe  
           1090                          1095                          1100  
  
 Ile Gly Val Ile Ile Asp Asn Phe Asn Gln Gln Gln Lys Lys Leu Gly  
  1105                          1110                          1115                          1120  
  
 Gly Gln Asp Ile Phe Met Thr Glu Glu Gln Lys Lys Tyr Tyr Asn Ala  
                           1125                          1130                          1135  
  
 Met Lys Lys Leu Gly Ser Lys Lys Pro Gln Lys Pro Ile Pro Arg Pro  
           1140                          1145                          1150  
  
 Leu Asn Lys Cys Gln Gly Leu Val Phe Asp Ile Val Thr Ser Gln Ile  
           1155                          1160                          1165  
  
 Phe Asp Ile Ile Ile Ile Ser Leu Ile Ile Leu Asn Met Ile Ser Met  
  1170                          1175                          1180  
  
 Met Ala Glu Ser Tyr Asn Gln Pro Lys Ala Met Lys Ser Ile Leu Asp  
  1185                          1190                          1195                          1200  
  
 His Leu Asn Trp Val Phe Val Val Ile Phe Thr Leu Glu Cys Leu Ile  
                           1205                          1210                          1215  
  
 Lys Ile Phe Ala Leu Arg Gln Tyr Tyr Phe Thr Asn Gly Trp Asn Leu  
           1220                          1225                          1230  
  
 Phe

&lt;210&gt; 8

&lt;211&gt; 1243

&lt;212&gt; PRT

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;223&gt; partial human NaN amino acid seq.

&lt;400&gt; 8

Ser Ile Val Ile Gly Ile Ala Ile Val Ser Tyr Ile Pro Gly Ile Thr  
   1                          5                          10                          15  
  
 Ile Lys Leu Leu Pro Leu Arg Thr Phe Arg Val Phe Arg Ala Leu Lys  
           20                          25                          30  
  
 Ala Ile Ser Val Val Ser Arg Leu Lys Val Ile Val Gly Ala Leu Leu  
           35                          40                          45  
  
 Arg Ser Val Lys Lys Leu Val Asn Val Ile Ile Leu Thr Phe Phe Cys  
   50                          55                          60

Leu Ser Ile Phe Ala Leu Val Gly Gln Gln Leu Phe Met Gly Ser Leu  
 65 70 75 80  
 Asn Leu Lys Cys Ile Ser Arg Asp Cys Lys Asn Ile Ser Asn Pro Glu  
 85 90 95  
 Ala Tyr Asp His Cys Phe Glu Lys Lys Glu Asn Ser Pro Glu Phe Lys  
 100 105 110  
 Met Cys Gly Ile Trp Met Gly Asn Ser Ala Cys Ser Ile Gln Tyr Glu  
 115 120 125  
 Cys Lys His Thr Lys Ile Asn Pro Asp Tyr Asn Tyr Thr Asn Phe Asp  
 130 135 140  
 Asn Phe Gly Trp Ser Phe Leu Ala Met Phe Arg Leu Met Thr Gln Asp  
 145 150 155 160  
 Ser Trp Glu Lys Leu Tyr Gln Gln Thr Leu Arg Thr Thr Gly Leu Tyr  
 165 170 175  
 Ser Val Phe Phe Phe Ile Val Val Ile Phe Leu Gly Ser Phe Tyr Leu  
 180 185 190  
 Ile Asn Leu Thr Leu Ala Val Val Thr Met Ala Tyr Glu Glu Gln Asn  
 195 200 205  
 Lys Asn Val Ala Ala Glu Ile Glu Ala Lys Glu Lys Met Phe Gln Glu  
 210 215 220  
 Ala Gln Gln Leu Leu Lys Glu Glu Lys Glu Ala Leu Val Ala Met Gly  
 225 230 235 240  
 Ile Asp Arg Ser Ser Leu Thr Ser Leu Glu Thr Ser Tyr Phe Thr Pro  
 245 250 255  
 Lys Lys Arg Lys Leu Phe Gly Asn Lys Lys Arg Lys Ser Phe Phe Leu  
 260 265 270  
 Arg Glu Ser Gly Lys Asp Gln Pro Pro Gly Ser Asp Ser Asp Glu Asp  
 275 280 285  
 Cys Gln Lys Lys Pro Gln Leu Leu Glu Gln Thr Lys Arg Leu Ser Gln  
 290 295 300  
 Asn Leu Ser Leu Asp His Phe Asp Glu His Gly Asp Pro Leu Gln Arg  
 305 310 315 320  
 Gln Arg Ala Leu Ser Ala Val Ser Ile Leu Thr Ile Thr Met Lys Glu  
 325 330 335  
 Gln Glu Lys Ser Gln Glu Pro Cys Leu Pro Cys Gly Glu Asn Leu Ala  
 340 345 350  
 Ser Lys Tyr Leu Val Trp Asn Cys Cys Pro Gln Trp Leu Cys Val Lys  
 355 360 365

Lys Val Leu Arg Thr Val Met Thr Asp Pro Phe Thr Glu Leu Ala Ile  
 370 375 380  
 Thr Ile Cys Ile Ile Ile Asn Thr Val Phe Leu Ala Met Glu His His  
 385 390 395 400  
 Lys Met Glu Ala Ser Phe Glu Lys Met Leu Asn Ile Gly Asn Leu Val  
 405 410 415  
 Phe Thr Ser Ile Phe Ile Ala Glu Met Cys Leu Lys Ile Ile Ala Leu  
 420 425 430  
 Asp Pro Tyr His Tyr Phe Arg Arg Gly Trp Asn Ile Phe Asp Ser Ile  
 435 440 445  
 Val Ala Leu Leu Ser Phe Ala Asp Val Met Asn Cys Val Leu Gln Lys  
 450 455 460  
 Arg Ser Trp Pro Phe Leu Arg Ser Phe Arg Val Leu Arg Val Phe Lys  
 465 470 475 480  
 Leu Ala Lys Ser Trp Pro Thr Leu Asn Thr Leu Ile Lys Ile Ile Gly  
 485 490 495  
 Asn Ser Val Gly Ala Leu Gly Ser Leu Thr Val Val Leu Val Ile Val  
 500 505 510  
 Ile Phe Ile Phe Ser Val Val Gly Met Gln Leu Phe Gly Arg Ser Phe  
 515 520 525  
 Asn Ser Gln Lys Ser Pro Lys Leu Cys Asn Pro Thr Gly Pro Thr Val  
 530 535 540  
 Ser Cys Leu Arg His Trp His Met Gly Asp Phe Trp His Ser Phe Leu  
 545 550 555 560  
 Val Val Phe Arg Ile Leu Cys Gly Glu Trp Ile Glu Asn Met Trp Glu  
 565 570 575  
 Cys Met Gln Glu Ala Asn Ala Ser Ser Ser Leu Cys Val Ile Val Phe  
 580 585 590  
 Ile Leu Ile Thr Val Ile Gly Lys Leu Val Val Leu Asn Leu Phe Ile  
 595 600 605  
 Ala Leu Leu Leu Asn Ser Phe Ser Asn Glu Glu Arg Asn Gly Asn Leu  
 610 615 620  
 Glu Gly Glu Ala Arg Lys Thr Lys Val Gln Leu Ala Leu Asp Arg Phe  
 625 630 635 640  
 Arg Arg Ala Phe Cys Phe Val Arg His Thr Leu Glu His Phe Cys His  
 645 650 655  
 Lys Trp Cys Arg Lys Gln Asn Leu Pro Gln Gln Lys Glu Val Ala Gly  
 660 665 670

Gly Cys Ala Ala Gln Ser Lys Asp Ile Ile Pro Leu Val Met Glu Met  
 675 680 685  
 Lys Arg Gly Ser Glu Thr Gln Glu Glu Leu Gly Ile Leu Thr Ser Val  
 690 695 700  
 Pro Lys Thr Leu Gly Val Arg His Asp Trp Thr Trp Leu Ala Pro Leu  
 705 710 715 720  
 Ala Glu Glu Glu Asp Asp Val Glu Phe Ser Gly Glu Asp Asn Ala Gln  
 725 730 735  
 Arg Ile Thr Gln Pro Glu Pro Glu Gln Gln Ala Tyr Glu Leu His Gln  
 740 745 750  
 Glu Asn Lys Lys Pro Thr Ser Gln Arg Val Gln Ser Val Glu Ile Asp  
 755 760 765  
 Met Phe Ser Glu Asp Glu Pro His Leu Thr Ile Gln Asp Pro Arg Lys  
 770 775 780  
 Lys Ser Asp Val Thr Ser Ile Leu Ser Glu Cys Ser Thr Ile Asp Leu  
 785 790 795 800  
 Gln Asp Gly Phe Gly Trp Leu Pro Glu Met Val Pro Lys Lys Gln Pro  
 805 810 815  
 Glu Arg Cys Leu Pro Lys Gly Phe Gly Cys Cys Phe Pro Cys Cys Ser  
 820 825 830  
 Val Asp Lys Arg Lys Pro Pro Trp Val Ile Trp Trp Asn Leu Arg Lys  
 835 840 845  
 Thr Cys Tyr Gln Ile Val Lys His Ser Trp Phe Glu Ser Phe Ile Ile  
 850 855 860  
 Phe Val Ile Leu Leu Ser Ser Gly Ala Leu Ile Phe Glu Asp Val His  
 865 870 875 880  
 Leu Glu Asn Gln Pro Lys Ile Gln Glu Leu Leu Asn Cys Thr Asp Ile  
 885 890 895  
 Ile Phe Thr His Ile Phe Ile Leu Glu Met Val Leu Lys Trp Val Ala  
 900 905 910  
 Phe Gly Phe Gly Lys Tyr Phe Thr Ser Ala Trp Cys Cys Leu Asp Phe  
 915 920 925  
 Ile Ile Val Ile Val Ser Val Thr Thr Leu Ile Asn Leu Met Glu Leu  
 930 935 940  
 Lys Ser Phe Arg Thr Leu Arg Ala Leu Arg Pro Leu Arg Ala Leu Ser  
 945 950 955 960  
 Gln Phe Glu Gly Met Lys Val Val Val Asn Ala Leu Ile Gly Ala Ile  
 965 970 975

Pro Ala Ile Leu Asn Val Leu Leu Val Cys Leu Ile Phe Trp Leu Val  
                   980                                  985                                  990  
 Phe Cys Ile Leu Gly Val Tyr Phe Phe Ser Gly Lys Phe Gly Lys Cys  
                   995                                  1000                                  1005  
 Ile Asn Gly Thr Asp Ser Val Ile Asn Tyr Thr Ile Ile Thr Asn Lys  
                   1010                                  1015                                  1020  
 Ser Gln Cys Glu Ser Gly Asn Phe Ser Trp Ile Asn Gln Lys Val Asn  
                   1025                                  1030                                  1035                                  1040  
 Phe Asp Asn Val Gly Asn Ala Tyr Leu Ala Leu Leu Gln Val Ala Thr  
                   1045                                  1050                                  1055  
 Phe Lys Gly Trp Met Asp Ile Ile Tyr Ala Ala Val Asp Ser Thr Glu  
                   1060                                  1065                                  1070  
 Lys Glu Gln Gln Pro Glu Phe Glu Ser Asn Ser Leu Gly Tyr Ile Tyr  
                   1075                                  1080                                  1085  
 Phe Val Val Phe Ile Ile Phe Gly Ser Phe Phe Thr Leu Asn Leu Phe  
                   1090                                  1095                                  1100  
 Ile Gly Val Ile Ile Asp Asn Phe Asn Gln Gln Gln Lys Lys Leu Gly  
                   1105                                  1110                                  1115                                  1120  
 Gly Gln Asp Ile Phe Met Thr Glu Glu Gln Lys Lys Tyr Tyr Asn Ala  
                   1125                                  1130                                  1135  
 Met Lys Lys Leu Gly Ser Lys Lys Pro Gln Lys Pro Ile Pro Arg Pro  
                   1140                                  1145                                  1150  
 Leu Asn Lys Cys Gln Gly Leu Val Phe Asp Ile Val Thr Ser Gln Ile  
                   1155                                  1160                                  1165  
 Phe Asp Ile Ile Ile Ile Ser Leu Ile Ile Leu Asn Met Ile Ser Met  
                   1170                                  1175                                  1180  
 Met Ala Glu Ser Tyr Asn Gln Pro Lys Ala Met Lys Ser Ile Leu Asp  
                   1185                                  1190                                  1195                                  1200  
 His Leu Asn Trp Val Phe Val Val Ile Phe Thr Leu Glu Cys Leu Ile  
                   1205                                  1210                                  1215  
 Lys Ile Phe Ala Leu Arg Gln Tyr Tyr Phe Thr Asn Gly Trp Asn Leu  
                   1220                                  1225                                  1230  
 Phe Asp Cys Val Val Val Leu Leu Ser Ile Val  
                   1235                                  1240

&lt;210&gt; 9

&lt;211&gt; 20

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

<220>  
<221> variation  
<222> (6)  
<223> r = a or g

<220>  
<223> Description of Artificial Sequence: rat NaN  
forward primer no. 1

<400> 9  
gaccertgga attggttgga 20

<210> 10  
<211> 20  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: rat NaN  
forward primer no. 2

<400> 10  
aatccctgga attggttgga 20

<210> 11  
<211> 20  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: rat NaN  
forward primer no. 3

<400> 11  
gaccctgga actggttaga 20

<210> 12  
<211> 20  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: rat NaN  
forward primer no. 4

<400> 12  
gatctttgga actggcttga 20

<210> 13  
<211> 21  
<212> DNA  
<213> Artificial Sequence



&lt;220&gt;

<223> Description of Artificial Sequence: rat Nan  
forward primer no. 5

&lt;400&gt; 13

aacatagtgc tggagttcag g

21

&lt;210&gt; 14

&lt;211&gt; 21

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

<223> Description of Artificial Sequence: rat NaN  
forward primer no. 6

&lt;400&gt; 14

gtggcctttg gattccggag g

21

&lt;210&gt; 15

&lt;211&gt; 24

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

<223> Description of Artificial Sequence: rat NaN  
reverse primer no. 1

&lt;400&gt; 15

caagaaggcc cagctgaagg tgtc

24

&lt;210&gt; 16

&lt;211&gt; 24

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

<223> Description of Artificial Sequence: rat NaN  
reverse primer no. 2

&lt;400&gt; 16

gaggaatgcc. cacgcaaagg aatc

24

&lt;210&gt; 17

&lt;211&gt; 24

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

<223> Description of Artificial Sequence: rat NaN  
reverse primer no. 3

<400> 17  
aagaaggac cagccaaagt tgtc 24

<210> 18  
<211> 20  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: rat NaN  
reverse primer no. 4, y = c or t, r = a or g, n =  
a or c or g or t, w = a or t

<400> 18  
acytccatrc anwccacat 20

<210> 19  
<211> 20  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: rat NaN  
reverse primer no. 5, r = a or g

<400> 19  
agraartcna gccarcacca 20

<210> 20  
<211> 19  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: rat NaN  
reverse primer no. 6

<400> 20  
tctgctgccg agccaggta 19

<210> 21  
<211> 20  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: rat NaN  
reverse primer no. 7

<400> 21  
ctgagataac tgaaatcgcc 20

<210> 22  
<211> 27  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: primer,  
Marathon AP-1

<400> 22  
ccatcctaatacgcactcactatagggc 27

<210> 23  
<211> 23  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: primer,  
Marathon AP-2

<400> 23  
actcactatagggctcgagcggc 23

<210> 24  
<211> 23  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: mouse NaN  
forward primer

<400> 24  
ccctgctgcgctcgggtgaagaag 23

<210> 25  
<211> 20  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: mouse NaN  
reverse primer

<400> 25  
gacaaagtagatcccagagg 20

<210> 26  
<211> 17  
<212> DNA  
<213> Artificial Sequence

&lt;220&gt;

<223> Description of Artificial Sequence: human NaN  
forward primer

&lt;400&gt; 26

ctcagtagtt ggcattgc

17

&lt;210&gt; 27

&lt;211&gt; 24

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

<223> Description of Artificial Sequence: human NaN  
reverse primer

&lt;400&gt; 27

ggaaagaagc acgaccacac agtc

24

&lt;210&gt; 28

&lt;211&gt; 94

&lt;212&gt; PRT

&lt;213&gt; Rattus norvegicus

&lt;220&gt;

&lt;223&gt; C-terminal truncated rat NaN

&lt;400&gt; 28

Ala Ala Gly Gln Ala Met Arg Lys Gln Gly Asp Ile Leu Gly Pro Asn  
1 5 10 15Ile His Gln Phe Ser Gln Ser Ser Glu Thr Pro Phe Leu Gly Cys Pro  
20 25 30Gln Gln Arg Thr Cys Val Ser Phe Val Arg Pro Gln Arg Val Leu Arg  
35 40 45Val Pro Trp Phe Pro Ala Trp Arg Thr Val Thr Phe Leu Ser Arg Pro  
50 55 60Arg Ser Ser Glu Ser Ser Ala Trp Leu Gly Leu Val Glu Ser Ser Gly  
65 70 75 80Trp Ser Gly Leu Pro Gly Glu Ser Gly Pro Ser Ser Leu Leu  
85 90

&lt;210&gt; 29

&lt;211&gt; 211

&lt;212&gt; DNA

&lt;213&gt; Mus musculus

&lt;400&gt; 29

agttaaatgt tgagtgaatt gtggtggtga tttccactt gaggcctttg tgttaaagcc 60

```

caatgtgtgt gtgtgtgtgt gtgtgtgtgt gtgtgtgtgt gtgtgtgtgt gtgtgtgggt 120
gggggggtgt ggcagagtct ggtattggtt aggtgagagc aatcccagaa cgtccacctg 180
ctcttccatt ttattaatca ggcaggcctc t 211

```

```

<210> 30
<211> 242
<212> DNA
<213> Mus musculus

```

```

<400> 30
gtaagccact ggctcttaac taaaatgctc gttggcatta gaacatttct gagctggggg 60
ggtgggtgtg gtggtggtgg tgggtggtgg ggtggtggtg gtggtggtgg tgatgggtgg 120
ggtggaggtg gnggtggagg tgggtggctg ggtggtggng gtggtggtgg tgggtggangt 180
ggangtgggt gcgtgggtgg gngngtgggt gtggaggtgg tggctgtggt ggtngtgggt 240
gc 242

```

```

<210> 31
<211> 200
<212> DNA
<213> Mus musculus

```

```

<400> 31
tgtgcatgct tgattcccag ctcttatggt ctgattactc ggtccttagg agcaaggcca 60
gactgtccac cctgacacac acacacacac acacacacac acacacacac acacacacac 120
acagtgtaga gaattacctc attcttggag tttctctgga aaaggaatgt ctcaaagcca 180
agttcacaga gcaacagctg 200

```

```

<210> 32
<211> 181
<212> DNA
<213> Mus musculus

```

```

<400> 32
tgttagaaac tctaagacaa tgaagcacca tgctggaaat aagagcacia actcttttctt 60
catgcattac cactgcttg tgetttcacc ttagtgctcg tgctctctct ttctctctct 120
ctctctctct ctctctctct ctctctctct ctctctctct ctctctcttt tttttttttt 180
t 181

```

```

<210> 33
<211> 128
<212> DNA
<213> Mus musculus

```

```

<400> 33
cacacacaca cacacacaca cacacacaca cacacacaca gagaaacact gtgcagtc 60
tacatatata gataaatata tcttaaaaaa agaaccatgt gattgagtta taaaatattc 120
caacttat 128

```

```

<210> 34
<211> 200
<212> DNA

```

<213> Mus musculus

<400> 34

```
agggtcatttc ctctgcagtg tgcttggcag gaaaaacttc ctggctattc aagtcagtgc 60
cctgcttgat catccatgta tcacacacac acaaaacaaa caaacaaca aacaaaaccc 120
tggggaagaa ggaagaggtt aagcacatag gcagagagca gccaggctga ctcagagcaa 180
acacctgatc attcttccat                                     200
```

<210> 35

<211> 158

<212> DNA

<213> Mus musculus

<400> 35

```
gtgctgggat caaaggcgtg egccgccacc acgcccggcc cctttttatg tttcaaattt 60
acttttatca tgtgcaegtg tgtgggtgcg tgcattgtgtg tgcgtgcgtg tgcgtgtgng 120
tgtgngtgtg tgtgtgtgtg tgtgtgtgtg tgtgtgtg                                     158
```

<210> 36

<211> 113

<212> DNA

<213> Mus musculus

<400> 36

```
cacacacaca cacacacaca cacacacaca cacacacaca cacacacaca cacacacttg 60
catctttgag ttaattggat aggctgagtc ttacaccgga atcatactgt tgc                                     113
```

<210> 37

<211> 200

<212> DNA

<213> Mus musculus

<400> 37

```
ccaatgagag actcttgtct caaaaaagcc atgggtgtcca gatcctgagg aataacacct 60
aagaatgtgc tctggcctga aaacacacac acacacacac acacacacac acacacacac 120
agttttatatt atttatataa aaaaatatgt ctctaggcat tgctgaaatg tctcctacag 180
gattaagtca accagagcca                                     200
```

<210> 38

<211> 7

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: protein seq.  
basis for rat NaN reverse primers

<220>

<221> VARIANT

<222> (3)

<223> Xaa = Val or Asp

1/28  
**FIG. 1A**

Nucleotide sequence of rat NaN. Translation initiation begins  
at position 41 (ATG). Reading frame ends at position 5336 (TGA).

```

1  ACGGTGCCCT GATCCTCTGT ACCAGGAAGA CAGGGTGAAG ATGGAGGAGA
51  GGTACTACCC GGTGATCTTC CCGGACGAGC GGAATTTCCG CCCCTTCACT
101 TCCGACTCTC TGGCTGCCAT AGAGAAGCGG ATTGCTATCC AAAAGGAGAG
151 GAAGAAGTCC AAAGACAAGG CGGCAGCTGA GCCCCAGCCT CGGCCTCAGC
201 TTGACCTAAA GGCCTCCAGG AAGTTACCTA AGCTTTATGG TGACATTCCC
251 CCTGAGCTTG TAGCGAAGCC TCTGGAAGAC CTGGACCCAT TCTACAAAGA
301 CCATAAGACA TTCATGGTGT TGAACAAGAA GAGAACAATT TATCGCTTCA
351 GCGCCAAGCG GGCCTTGTTT ATTCTGGGGC CTTTAAATCC CCTCAGAAGC
401 TTAATGATTC GTATCTCTGT CCATTCAGTC TTTAGCATGT TCATCATCTG
451 CACGGTGATC ATCAACTGTA TGTTCATGGC GAATTCATG GAGAGAAGTT
501 TCGACAACGA CATTCCCGAA TACGTCTTCA TTGGGATTTA TATTTTAGAA
551 GCTGTGATTA AAATATTGGC AAGAGGCTTC ATTGTGGATG AGTTTTCTTT
601 CCTCCGAGAT CCGTGGAAC TGGCTGGACTT CATTTGCATT GGAACAGCGA
651 TCGCAACTTG TTTTCCGGGC AGCCAAGTCA ATCTTTCAGC TCTTCGTACC
701 TTCCGAGTGT TCAGAGCTCT GAAGGCGATT TCAGTTATCT CAGGTCTGAA
751 GGTCATCGTA GGTGCCCTGC TGCCTCGGT GAAGAAGCTG GTAGACGTGA
801 TGGTCCTCAC TCTCTCTGC CTCAGCATCT TTGCCCTGGT CGGTCAGCAG
851 CTGTTTCATG GAATTCCTGAA CCAGAAGTGT ATTAAGCACA ACTGTGGCCC
901 CAACCTGCA TCCAACAAGG ATTGTTTTGA AAAGGAAAAA GATAGCGAAG
951 ACTTCATAAT GTGTGGTACC TGGCTCGGCA GCAGACCCTG TCCCAATGGT
1001 TCTACGTGCG ATAAAACCAC ATTGAACCCA GACAATAATT ATACAAAGTT
1051 TGACAACTTT GGCTGGTCCT TTCTCGCCAT GTTCCGGGTT ATGACTCAAG
1101 ACTCCTGGGA GAGGCTTTAC CGACAGATCC TGCAGGACCTC TGGGATCTAC
1151 TTTGTCTTCT TCTTCGTGGT GGTTCATCTT CTGGGCTCCT TCTACCTGCT
1201 TAACCTAACC CTGGCTGTTG TCACCATGGC TTATGAAGAA CAGAACAGAA
1251 ATGTAGCTGC TGAGACAGAG GCCAAGGAGA AAATGTTTCA GGAAGCCCAG
1301 CAGCTGTAA GGGAGGAGAA GGAGGCTCTG GTTGCCATGG GAATTGACAG
1351 AAGTTCCCTT AATTCCCTTC AAGCTTCATC CTTTCCCGG AAGAAGAGGA
1401 AGTTTTTCGG TAGTAAGACA AGAAAGTCCT TCTTTATGAG AGGGTCCAAG
1451 ACGGCCCAAG CCTCAGCGTC TGATTCAGAG GACGATGCCT CTAAAAATCC
1501 ACAGCTCCTT GAGCAGACCA AACGACTGTC CCAGAACTTG CCAGTGATC
1551 TCTTTGATGA GCACGTGGAC CCCCTCCACA GGCAGAGAGC GCTGAGCGCT
1601 GTCAGTATCT TAACCATCAC CATGCAGGAA CAAGAAAAAT TCCAGGAGCC
1651 TTGTTTCCCA TGTGGGAAAA ATTTGGCCTC TAAGTACCTG GTGTGGGACT

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**FIG. 1B**

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1701 GTAGCCCTCA ATGGCTGTGC ATAAAGAAGG TCCTGCGGAC CATCATGACG  
 1751 GATCCCTTTA CTGAGCTGGC CATCACCATC TGCATCATCA TCAATACCGT  
 1801 TTTCTTAGCC GTGGAGCACC ACAACATGGA TGACAACCTTA AAGACCATAC  
 1851 TGAAAATAGG AAACCTGGGT TTTACGGGAA TTTTCATAGC GGAAATGTGT  
 1901 CTCAAGATCA TCGCGCTCGA CCCTTACCAC TACTTCCGGC ACGGCTGGAA  
 1951 TGTTTTTGAC AGCATCGTGG CCCTCCTGAG TCTCGCTGAT GTGCTNTACA  
 2001 ACACACTGTC TGATAACAAT AGGTCTTTCT TGGCTTCCCT CAGAGTGCTG  
 2051 AGGGTCTTCA AGTTAGCCAA ATCCTGGCCC ACGTTAAACA CTCTCATTA  
 2101 GATCATCGGC CACTCCGTGG GCGCGCTTGG AAACCTGACT GTGGTCTTGA  
 2151 CTATCGTGGT CTTTCATCTTT TCTGTGGTGG GCATGCGGCT CTTGCGCACC  
 2201 AAGTTTAACA AGACCGCCTA CGCCACCCAG GAGCGGCCCA GGCGGCGCTG  
 2251 GCACATGGAT AATTTCTACC ACTCCTTCCT GGTGGTGTTC CGCATCCTCT  
 2301 GTGGGGAATG GATCGAGAAC ATGTGGGGCT GCATGCAGGA TATGGACGGC  
 2351 TCCCCGTGTG GCATCATTTG CTTTGTCTTG ATAATGGTGA TCGGGAAGCT  
 2401 TGTGGTGCTT AACCTCTTCA TTGCCTTGCT GCTCAATTCC TTCAGCAATG  
 2451 AGGAGAAGGA TGGGAGCCTG GAAGGAGAGA CCAGGAAAAC CAAAGTGCAG  
 2501 CTAGCCCTGG ATCGGTTCCT CCGGGCCTTC TCCTTCATGC TGCACGCTCT  
 2551 TCAGAGTTTT TGTGCAAGA AATGCAGGAG GAAAACTCG CCAAAGCCAA  
 2601 AAGAGACAAC AGAAAGCTTT GCTGGTGAGA ATAAAGACTC AATCCTCCCG  
 2651 GATGCGAGGC CCTGGAAGGA GTATGATACA GACATGGCTT TGTACACTGG  
 2701 ACAGGCCGGG GCTCCGCTGG CCCCCTCGC AGAGGTAGAG GACGATGTGG  
 2751 AATATGTGGG TGAAGGCGGT GCCCTACCCA CCTCACAACA TAGTGCTGGA  
 2801 GTTCAGGCCG GTGACCTCCC TCCAGAGACC AAGCAGCTCA CTAGCCCGGA  
 2851 TGACCAAGGG GTTGAAATGG AAGTATTTTC TGAAGAAGAT CTGCATTTAA  
 2901 GCATACAGAG TCCTCGAAAG AAGCTGACG CAGTGAGCAT GCTCTCGGAA  
 2951 TGCAGCACAA TTGACCTGAA TGATATCTTT AGAAATTTAC AGAAAACAGT  
 3001 TTCCCCCAA AAGCAGCCAG ATAGATGCTT TCCCAAGGGC CTTAGTTGTC  
 3051 ACTTCTATG CCACAAAACA GACAAGAGAA AGTCCCCCTG GGTCCCTGTGG  
 3101 TGGAACATTC GGAAAACCTG CTACCAAATC GTGAAGCACA GCTGGTTTGA  
 3151 GAGTTTCATA ATCTTTGTTA TTCTGCTGAG CAGTGGAGCG CTGATATTTG  
 3201 AAGATGTCAA TCTCCCCAGC CGGCCCCAAG TTGAGAAATT ACTAAGGTGT  
 3251 ACCGATAATA TTTTCACATT TATTTTCCTC CTGGAAATGA TCCTGAAGTG  
 3301 GGTGGCCTTT GGATTCCGGA GGTATTTTAC CAGTGCCTGG TGCTGGCTTG  
 3351 ATTTCCTCAT TGTGGTGGTG TCTGTGCTCA GTCTCATGAA TCTACCAAGC  
 3401 TTGAAGTCCT TCCGGACTCT GCGGGCCCTG AGACCTCTGC GGGCGCTGTC  
 3451 CCAGTTTGAA GGAATGAAGG TTGTCGTCTA CGCCCTGATC AGCGCCATAC  
 3501 CTGCCATTC TCAATGTCTG CTGGTCTGCC TCATTTTCTG GCTCGTATTT  
 3551 TGTATCTTGG GAGTAAATTT ATTTTCTGGG AAGTTTGGA GGTGCATTA  
 3601 CGGGACAGAC ATAAATATGT ATTTGGATT TACCGAAGTT CCGAACCAG



**FIG. 1C**

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3651 GCCAATGTAA CATTAGTAAT TACTCGTGGA AGGTCCCGCA GGTCAACTTT  
 3701 GACAACGTGG GGAATGCCTA TCTCGCCCTG CTGCAAGTGG CAACCTATAA  
 3751 GGGCTGGCTG GAAATCATGA ATGCTGCTGT CGATTCCAGA GAGAAAGACG  
 3801 AGCAGCCGGA CTTTGAGGCG AACCTCTACG CGTATCTCTA CTTTGTTGGTT  
 3851 TTTATCATCT TCGGCTCCTT CTATTACCCTG AACCTCTTTA TCGGTGTTAT  
 3901 TATTGACAAC TTCAATCAGC AGCAGAAAAA GTTAGGTGGC CAAGACATTT  
 3951 TTATGACAGA AGAACAGAAG AAATATTACA ATGCAATGAA AAAGTTAGGA  
 4001 ACCAAGAAAC CTCAAAAGCC CATCCCAAGG CCCCTGAACA ANTGTCAAGC  
 4051 CTTTGTGTTC GACCTGGTCA CAAGCCATGT CTTTGACGTC ATCATCTCTGG  
 4101 GTCTTATTGT CTAAATATG ATTATCATGA TGGCTGAATC TGCCGACCAG  
 4151 CCCAAAGATG TGAAGAAAAC CTTTGATATC CTCAACATAG CCTTCGTGGT  
 4201 CATCTTTACC ATAGAGTGTC TCATCAAAGT CTTTGCTTTG AGGCAACACT  
 4251 ACTTCACCAA TGGCTGGAAC TTATTTGATT GTGTGGTCGT GGTTCCTTCT  
 4301 ATCATTAGTA CCCTGGTTTC CCGCTTGGAG GACAGTGACA TTTCTTTCCC  
 4351 GCCCACGCTC TTCAGAGTCG TCCGCTTGGC TCGGATTGGT CGAATCCTCA  
 4401 GGCTGGTCCG GGCTGCCCCG GGAATCAGGA CCCTCCTCTT TGCTTTGATG  
 4451 ATGTCTCTCC CCTCTCTCTT CAACATCGGT CTGCTGCTCT TCCTGGTGAT  
 4501 GTTCATTTAC GCCATCTTTG GGATGAGCTG GTTTTCCAAA GTGAAGAAGG  
 4551 GCTCCGGGAT CGACGACATC TTCAACTTCG AGACCTTTAC GGGCAGCATG  
 4601 CTGTGCCTCT TCCAGATAAC CACTTCGGCT GGCTGGGATA CCCTCCTCAA  
 4651 CCCCATGCTG GAGGCAAAAG AACACTGCAA CTCCTCCTCC CAAGACAGCT  
 4701 GTCAGCAGCC GCAGATAGCC GTCGCTACT TCGTCAGTTA CATCATCATC  
 4751 TCCTTCCTCA TCGTGGTCAA CATGTACATC GCTGTGATCC TCGAGAACTT  
 4801 CAACACAGCC ACGGAGGAGA GCGAGGACCC TCTGGGAGAG GACGACTTTG  
 4851 AAATCTTCTA TGAGGTCTGG GAGAAGTTTG ACCCCGAGGC GTCGCAGTTC  
 4901 ATCCAGTATT CGGCCCTCTC TGACTTTGCG GACGCCCTGC CGGAGCCGTT  
 4951 GCGTGTGGCC AAGCCGAATA AGTTTCAGTT TCTAGTGATG GACTTGCCCA  
 5001 TGGTGATGGG CGACCGCCTC CATTCGATGG ATGTTCTCTT TGCTTTCACT  
 5051 ACCAGGGTCC TCGGGGACTC CAGCGCCTTG GATACCATGA AAACCATGAT  
 5101 GGAGGAGAAG TTTATGGAGG CCAACCTTTT TAAGAAGCTC TACGAGCCCA  
 5151 TAGTCACCAC CACCAAGAGG AAGGAGGAGG AGCAAGGCGC CGCCGTCATC  
 5201 CAGAGGGCCT ACCGGAACA CATGGAGAAG ATGGTCAAAC TGAGGCTGAA  
 5251 GGACAGGTCA AGTTCATCGC ACCAGGTGTT TTGCAATGGA GACTTGTTCA  
 5301 GCTTGATGTG GGCCAAGGTC AAGGTTTACA ATGACTGAAC CCTCATCTCC  
 5351 ACCCTTACCT CACTGCCTCA CAGCTTAGCC TCCAGCCTCT GCGGAGCAGG  
 5401 CGGCAGACTC ACTGAACACA GGCCGTTCGA TCTGTGTTTT TGGCTGAACG  
 5451 AGGTGACAGG TTGGCGTCCA TTTTAAATG ACTCTTGGA AGATTTTCATG  
 5501 TAGAGAGATG TTAGAAGGGA CTGCAAAGGA CACCGACCAT AACGGAAGGC  
 5551 CTGGAGGACA GTCCAACTTA CATAAAGATG AGAAACAAGA AGGAAAGATC

***FIG. 1D***

5601 CCAGGAAAAC TTCAGATTGT GTTCTCAGTA CATCCCCAA TGTGTCTGTT  
5651 CGGTGTTTTG AGTATGTGAC CTGCCACATG TAGCTCTTTT TTGCATGTAC  
5701 GTCAAAACCC TGCAGTAAGT TAATAGCTTG CTACGGGTGT TCCTACCAGC  
5751 ATCACAGAAT TGGGTGTATG ACTCAAACCT AAAAGCATGA CTCTGACTTG  
5801 TCAGTCAGCA CCCCRACTTT CAGACGCTCC AATCTCTGTC CCAGGTGTCT  
5851 AACGAATAAA TAGGTAAAAG AAAAA

**FIG. 2A**

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Predicted amino acid sequence of rat NaN (1765 a.a).

1 MEERYYPVIF PDERNFRPFT SDSLAAIEKR IAIQKERKKS KDKAAAEPQP  
 51 RPQLDLKASR KLPKLYGDIP PELVAKPLED LDPFYKDHKT FMVLNKKRTI  
 DI-S1  
 101 YRFSAKRALF ILGPFNPLRS LMIRISVHSV FSMFIICTVI INCMFMANS  
 DI-S2 DI-S3  
 151 ERSFDNDIPE YVFIGIYILE AVIKILARGF IVDEFSFLRD PWNWLD FIVI  
 DI-S4  
 201 GTAIATCFPG SQVNLSALRT FRVFRALKAI SVISGLKVIV GALLRSVKKL  
 DI-S5  
 251 VDV~~M~~VLTLEFC LSIFALVGQO LFMGILNQKC IKHNCGPNPA SNKDCFEKEK  
 DI-SS1  
 301 DSEDFIMCGT WLGSRPCPNG STCDKTTLNP DNNYTKF~~DNF~~ GWSFLAMFRV  
 DI-SS2 DI-S6  
 351 MTOD~~S~~WERLY RQILRTSGIY FVFFFVVVIF LGSFYLLNLT LAVVTMAYEE  
 401 QNRNVAAETE AKEKMFQEAQ QLLREEKEAL VAMGIDRSSL NSLQASSFSP  
 451 KKRKFFGSKT RKSFFMRGSK TAQASASDSE DDASKNPQLL EQTKRLSQNL  
 501 PVDLFDEHVD PLHRQRALSA VSILTITMQE QEKFQEPFCF CGKNLASKYL  
 DII-S1  
 551 VWDCSPQWLC IKKVLRTIMT DPFTELAITI CIIINTVFLA VEHNMDDNL  
 DII-S2 DII-S3  
 601 KTILKIGNWV FTGIFIAEMC LKIIALDPYH YFRHGWNVFD SIVALLSLAD  
 DII-S4  
 651 VLNTLSDNN RSFLASLRVL RVFKIAKSWP TLNTLIKIIG HSVGALGNLT  
 DII-S5 DII-SS1  
 701 VVLTIVVFIF SVVGMRLFGT KFNKTAYATQ ERPRRRWHMD NEYHSFLVVF  
 DII-SS2 DII-S6  
 751 RILC~~G~~EWIEN MWGCMQDMDG SPLCIIVFVL IMVIGKLVVL NLFIALLLNS

**FIG. 2B**

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801 FSNEEKDGSL EGETRKTQVQ LALDRFRRAF SFMLHALQSF CCKKCRRKNS  
851 PKPKETTESF AGENKDSILP DARPWKEYDT DMALYTGQAG APLAPLAEVE  
  
901 DDVEYCGEGG ALPTSQHSAG VQAGDLPPET KQLTSPDDQG VEMEVFSEED  
  
951 LHLSIQSPRK KSDAVSMLSE CSTIDLNDIF RNLQKTVSPK KQPDRCFPKG  
DIII-S1  
1001 LSCHFLCHKT DKRKSPWVLW WNIRKTCYQI VKHSWFESFI IFVILLSSGA  
DIII-S2  
1051 LIFEDVNLPS RPQVEKLLRC TDNIFTFIFL LEMILKWVAF GERRYFTSAW  
DIII-S3 DIII-S4  
1101 CWLDFLIVVV SVLSLMNLPS LKSFRTLRL RPLRALSOFE GMKVVVYALI  
DIII-S5  
1151 SAIPAILNVL LVCLIFWLVE CILGVNLESG KFGRCINGTD INMYLDFTEV  
DIII-SS1 DIII-SS2  
1201 PNRSQCNISN YSWKVPQVNF DNVGNAYLAL LOVATYKGWL EIMNAAVDSR  
DIII-S6  
1251 EKDEQPDFEA NLYAYLYFVV FIIFGSFETL NLEFIGVIIDN FNQQQKKLGG  
DIV-S1  
1301 QDIFMTEEQK KYYNAMKKLG TKKPQKPIPR PLNRCQAFVF DLVTSHVFDV  
  
1351 IILGLIVLNM IIMMAESADQ PKDVKKTFDI LNIAFVVIFT IECLIKVFAL  
DIV-S4  
1401 RQHYFTNGWN LFDCVVVVLS IISTLVSRL E DSDISFPPTL FRVRLARIG  
DIV-S5  
1451 RILRLVRAAR GIRTLLEFALM MSLPSLFNIG LLLFLVMFIY AIFGMSWFSK  
DIV-SS1 DIV-SS2  
1501 VKKGSGIDDI FNFETFTGSM LCLFOITTS AWDTLLNPML EAKEHCNSSS  
DIV-S6  
1551 QDSCQQPOIA VVYFVSYIII SFLIVNMYI AVILENFNTA TEESEDPLGE  
  
1601 DDFEIFYEVW EKFDPEASQF IQYSALSDFA DALPEPLRVA KPNKFQFLVM  
  
1651 DLPMVMGDRL HCMDVLFAFT TRVLGDSSGL DTMKTMMEEK FMEANPFKKL

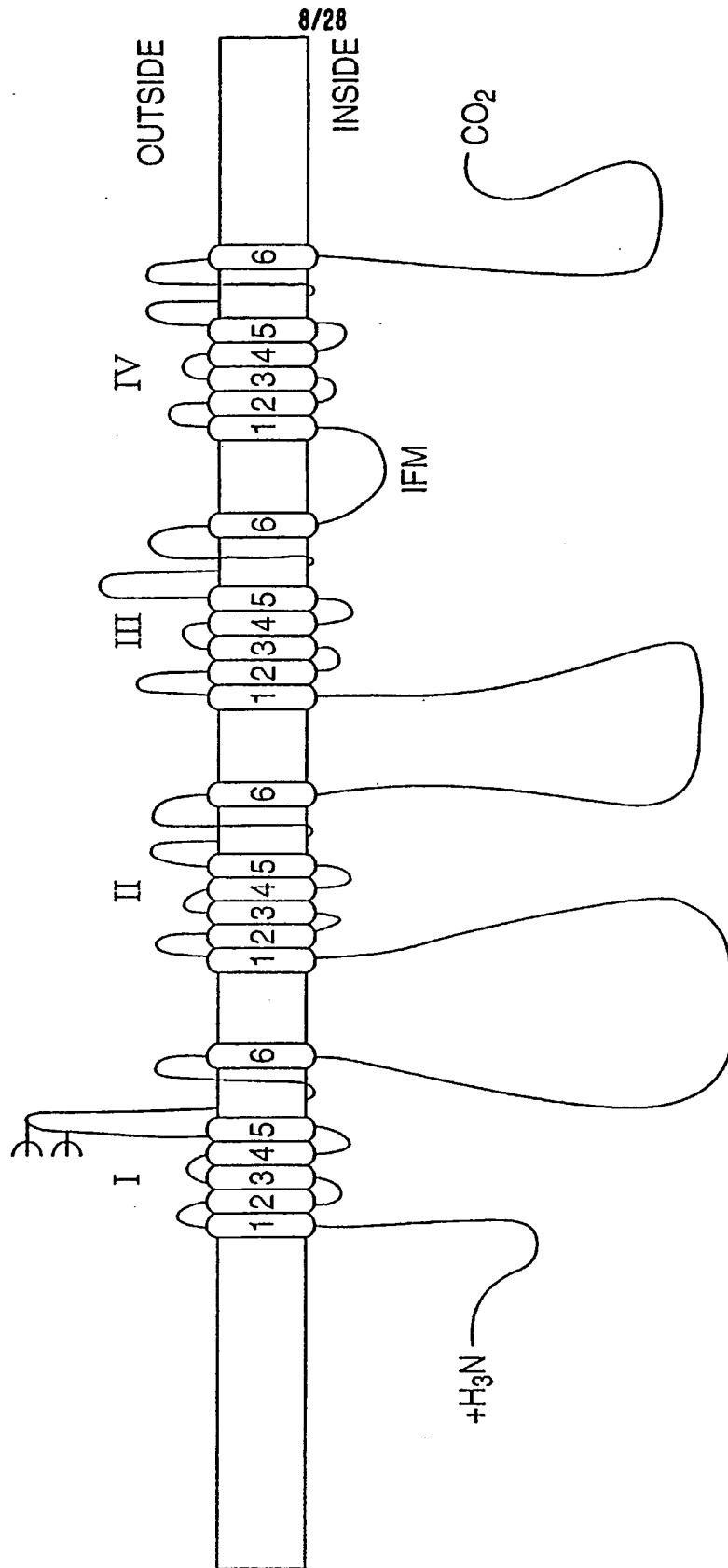
*FIG. 2C*

1701 YEPIVTTTKR KEEEQGAIVI QRAYRKHMEK MVKLRLKDRS SSSHQVFCNG

1751 DLSSLDVAKV KVHND\*

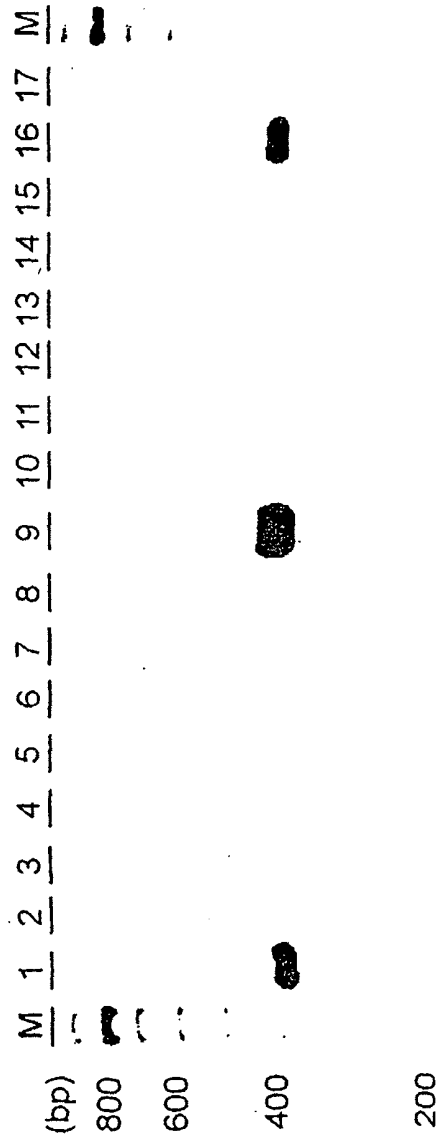
**FIG. 3**

VOLTAGE-GATED SODIUM CHANNEL  $\alpha$  SUBUNIT



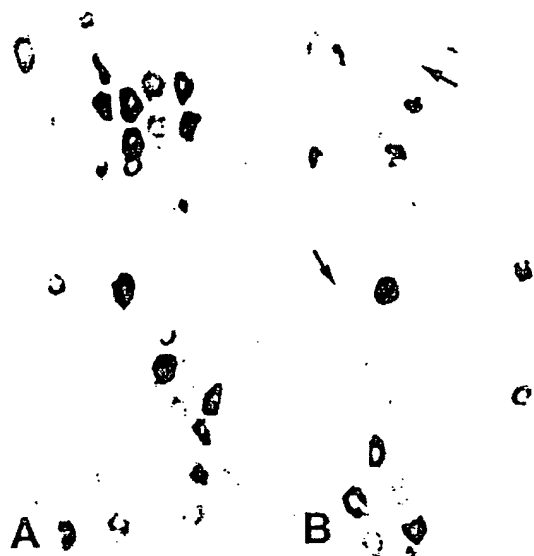
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**FIG. 4**



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**FIG. 5**



C — D

E F —



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**FIG. 6**

RESTRICTION ENZYME ANALYSIS OF  $\alpha$ -SUBUNIT PCR PRODUCTS FROM DOMAIN I USING THE FOLLOWING PRIMERS: NACHD1A.1-4 (FORWARD PRIMERS) AND NAAGEN.REV1-3 (REVERSE PRIMERS).

Generic Primer pair	F1 R1	F2 R1	F1 R1	F1/F3 R1	F1 R1	F1/F3 R1	F1 R1	F1/F3 R1	F1 R1	F2 R2	F4 R3	F2 R3
	$\alpha$ I 558 bp	$\alpha$ II 561 bp	$\alpha$ III 561 bp	$\alpha$ VI 507 bp	$\alpha$ PN1 501 bp	$\alpha$ RH1 518 bp	$\alpha$ u1 602 bp	$\alpha$ SNS 479 bp	$\alpha$ NaG 501 bp	$\alpha$ NaN 468 bp		
EcoR V	+	-	-	-	-	-	-	-	-	-	-	-
EcoNI	-	+	-	-	-	-	-	-	-	-	-	-
Ava I	-	-	+	-	-	-	-	-	-	-	-	-
Sph I	-	-	-	+	-	-	-	-	-	-	-	-
Bam H I	-	-	-	-	+	-	-	+	-	-	-	-
Acc I	-	-	-	-	-	+	-	-	-	-	+	185, 283
Ngo M I	-	-	-	-	-	-	+	-	+	-	+	-
Afl II	-	-	-	-	-	-	-	+	-	+	-	-
Xba I	-	-	-	-	-	-	-	-	+	-	+	-
EcoR I	-	-	-	-	-	-	-	-	-	-	-	+

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**FIG. 7A-1**

Sequence of the mouse NaN cDNA.

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1      TCTGAGCCAA GGGTGAAGAT GGAGGAGAGG TACTATCCAG TGATCTTCCC AGACGAGAGG
61     AATTTCCGCC CCTTCACTTT CGACTCTTTG GCTGCAATAG AGAAGCGGAT CACCATCCAA
121    AAGGAGAAGA AGAAATCCAA AGACAAGGCA GCAACTGAGC CCCAGCCTCG GCCTCAGCTC
181    GACCTAAAGG CCTCCAGGAA GTTACCTAAG CTCTATGGCG ACGTTCCCCC TGACCTTATA
241    GCGAAGCCCC TGGAAGATCT GGACCCATTT TACAAAGACC ATAAGACATT CATGGTATTG
301    AACAGAAGA GAACAATCTA TCGCTTCAGC GCCAAGAGGG CCTTGTTTCAT TCTGGGGCCT
361    TTTAATCCCA TCAGAAGCTT CATGATTCGC ATCTCTGTCC ATTCAGTCTT CAGCATGTTC
421    ATTATCTGCA CAGTGATCAT CAACTGTATG TTCATGGCTA ATAATTCTTC TGTGGACAGT
481    CGTCCTAGCA GTAACATTCC CGAATACGTC TTCATTGGGA TTTATGTTTT AGAAGCTGTG
541    ATTAAAATAT TGGCAAGAGG CTTCAATTGT GATGAGTTTT CCTACCTCCG AGATCCTTGG
601    AACTGGCTGG ACTTCATTGT CATCGGAACA GCGATAGCGC CTGTGTTTTCT CGGTAACAAA
661    GTCAATAATC TTTCCACTCT ACGTACCTTC CGAGTGTGGA GAGCTCTGAA AGCCATTTCT
721    GTAATCTCAG GTCTGAAGGT CATCGTGGGT GCCCTGCTGC GCTCCGTGAA GAAGCTAGTG
781    GACGTGATGG TCCTCACTCT CTTTTGCCTC AGCATCTTTG CCCTGGTTGG TCAGCAGCTC
841    TTCATGGGAA TTCTGAGCCA GAAATGTATT AAGGACGACT GTGGCCCTAA CGCTTTTTCC
901    AACAAAGGATT GCTTTGTAAA AGAAAATGAT AGCGAGGACT TCATAATGTG TGGCAACTGG
961    CTCGGCAGAA GATCCTGCCC CGATGGTTCC ACGTGCAATA AAACCACATT TAACCCAGAT
1021   TATAATTATA CAACTTTGA CAGCTTTGGC TGGTCTTTTC TCGCCATGTT CCGGGTTATG
1081   ACTCAAGACT CCTGGGAGAA GCTTTATCGA CAGATCCTTC GCACCTCCGG GATCTACTTT
1141   GTCTTCTTCT TCGTGGTCGT CATCTTCCTG GGCTCTTTCT ACCTGCTTAA CTTAACCCCTG
1201   GCTGTGCTCA CCATGGCTTA CGAGGAACAG AACAGAAATG TCGCTGCCGA GACAGAGGCC
1261   AAGGAGAAGA TGTTTCAGGA AGCCCAGCAG CTGTTGAGGG AGGAAAAGGA GGCTCTGGTT
1321   GCCATGGGAA TTGACAGAAC TTCCCTTAAT TCCCTCCAAG CTTGCTCCTT TTCCCCAAAG
1381   AAGAGGAAGT TTTTGGCAG TAAGACAAGA AAGTCCTTCT TTATGAGAGG GTCCAAGACA
1441   GCCCAGCCT CAGCGTCCGA TTCAGAGGAC GATGCCTCTA AAAACCCACA ACTCCTTGAG
1501   CAAACAAAAC GACTATCCCA GAACTTGCCC GTAGAACTCT TTGATGAGCA CGTGGACCCC
1561   CTCCATAGGC AGAGAGCGCT GAGTGCCGTC AGTATCTTAA CCATCACCAT GCAGGAACAA
1621   GAAAAATCCC AGGAGCCTTG TTTCCCGTGT GGGAAAAACT TGGCATCCAA GTACCTGGTG
1681   TGGGAATGTA GCCCTCCGTG GCTGTGCATA AAGAAGGTCC TGCAGACTAT CATGACAGAC
1741   CCCTTCACTG AGCTGGCCAT CACCATCTGC ATCATCGTCA AACTGTCTT CTTGGCCATG
1801   GAACACCACA ATATGGATAA CTCTTTAAAA GACATACTGA AAATAGGAAA CTGGGTTTTCT
1861   ACTGGAATTT TCATAGCGGA AATGTGTCTC AAGATCATTG CGCTAGACCC TTACCACTAC
1921   TTCCGGCACG GCTGGAACAT CTTTGACAGC ATTGTGGCCC TTGTGAGTCT CGCTGACGTG

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**FIG. 7A-2**

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1981 CTCTTCCACA AACTGTCTAA AAACCTCTCC TTCTTGGCTT CCCTCAGAGT GCTGAGGGTC
2041 TTCAAGTTAG CCAAATCCTG GCCCACATTA AACACTCTCA TTAAGATCAT CGGCCACTCC
2101 GTGGGTGCGC TCGGAAACCT GACTGTGGTC CTAACGATCG TGGTCTTCAT CTTTCCGTG
2161 GTTGGCATGC GGCTCTTTGG TGCCAAGTTT AACAAGACTT GCTCCACCTC TCCGGAGTCC
2221 CTCCGGCGCT GGCACATGGG TGATTTCTAC CATTCTTCC TGGTGGTGTT CCGCATCCTC
2281 TGTGGGGAGT GGATCGAGAA CATGTGGGAA TGCATGCAGG AGATGGAAGG CTCCCCGTG
2341 TGTGTCATCG TCTTTGTGCT GATCATGGTG GTCGGGAAGC TCGTGGTGCT TAACCTCTTC
2401 ATTGCCTTGC TGCTCAATTC CTTCAGCAAT GAGGAAAAGG ATGGGAACCC AGAAGGAGAG
2461 ACCAGGAAAA CCAAAGTGCA GCTAGCCCTG GATCGGTTCA GCCGAGCGTT CTAATTATG
2521 GCGCGCGCTC TTCAGAATTT CTGTTGCAAG AGATGCAGGA GGCAAACTC GCCAAAGCCA
2581 AATGAGGCAA CAGAAAGCTT TGCTGGTGAG AGTAGAGACA CAGCCACCCT GGATACAAGG
2641 TCCTGGAAGG AGTATGATTC AGAAATGACT CTGTACACTG GGCAGGCCGG GGCTCCACTG
2701 GCCCCACTGG CAAAAGAAGA GGACGATATG GAATGTTGTG GTGAATGTGA TGCCTCACCT
2761 ACCTCACAGC CTAGTGAGGA AGCTCAGGCC TGTGACCTCC CTCTGAAGAC CAAGCGGCTC
2821 CCCAGCCCAQ ATGACCACGG GGTGAAATG GAAGTGTGTTT CCGAAGAAGA TCCGAATTTA
2881 ACCATACAGA GTGCTCGAAA GAAGTCTGAT GCGGCAAGCA TGCTCTCAGA ATGCAGCACA
2941 ATAGACCTGA ATGATATCTT TAGAAATTTA CAGAAAACAG TTTCCCCCA AAAGCAACCA
3001 GATCGATGCT TTCCCAAGGG CCTCAGTTGT ATCTTTCTAT GTTGCAAAAC AATCAAAAAA
3061 AAGTCCCCCT GGTCTCTGTG GTGGAATCTT CGGAAAACCT GCTACCAAT CGTGAAGCAT
3121 AGCTGTTTG AGAGCTTCAT AATTTTGTG ATCCTGCTGA GCAGCGGAGC ACTGATATTC
3181 GAAGATGTCA ATCTTCCCAQ CCGGCCCAA GTTGA AAAAT TACTGAAGTG TACCGATAAT
3241 ATTTTCACAT TTATTTTCT CCTGGAATG ATTTTGAAGT GGGTGGCCTT TGGATTCCGG
3301 AAGTATTTCA CCAGTGCTG GTGCTGGCTC GATTTCTCTA TTGTGGTGGT GTCTGTGCTC
3361 AGCCTCACGA ACTTACCAA CTTGAAGTCC TTCCGGAATC TGCGAGCGCT GAGACCTCTG
3421 CGGGCACTGT CTCAGTTTGA AGGAATGAAG GTTGTGTGCA ATGCCCTCAT GAGTGCCATA
3481 CCTGCCATCC TCAATGTCTT GCTGGTCTGC CTCATTTTCT GGCTCATATT TTGTATCCTG
3541 GGAGTAAATT TTTTCTGGA GAAGTTTGA AGATGCATTA ATGGAACAGA CATAAATAAA
3601 TATTTCAACG CTCCAATGT TCCAAACCAA AGCCAATGTT TAGTTAGTAA TTACACGTGG
3661 AAAGTCCCGA ATGTCAACTT TGACAACGTG GGGAAATGCCT ACCTTGCCCT GCTGCAAGTG
3721 GCGACCTATA AGGGCTGGCT GGACATTATG AATGCAGCTG TTGATTCCAG AGGGAAAGAT
3781 GAGCAGCCGG CTTTGAGGC GAATCTATAC GCATACCTTT ACTTCGTGGT TTTATCATC
3841 TTCGGCTCAT TCTTACCCT GAACCTCTT ATCGGTGTTA TTATTGACAA CTTCATCAG
3901 CAGCAGAAAA AGTTAGGTGG CCAAGACATT TTTATGACAG AAGAACAGAA GAAATATTAC
3961 AATGCAATGA AAAAGTTAGG AACCAAGAAG CCTCAAAAGC CCATCCCAAG GCCCCTGAAC
4021 AAATGTCAAG CCTTCGTGTT CGATTTGGTC ACAAGCCAGG TCTTTGACGT CATCATCTG
4081 GGTCTTATTG TCACAAACAT GATTATCATG ATGGCTGAAT CTGAAGGCCA GCCCAACGAA
4141 GTGAAGAAAA TCTTTGATAT TCTCAACATA GTCTTCGTGG TCATCTTTAC CGTAGAGTGT
4201 CTCATCAAAG TCTTTGCTTT GAGGCAACAC TACTTCACCA ATGGCTGGAA CTTATTTGAT

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**FIG. 7A-3**

4261 TGTGTGGTCG TGGTTCTTTC CATCATTAGT ACCTTGTTT CTGGCTTGA GAACAGCAAC  
4321 GTCTTCCCGC CCACACTCTT CAGGATTGTC CGCTTGGCTC GGATCGGTCG AATCCTCAGA  
4381 CTGGTCCGGG CGGCTCGAGG AATCAGGACA CTCCTTTTCG CGTTGATGAT GTCTCTCCCC  
4441 TCTCTCTTCA ACATTGGTCT GCTTCTCTTT CTGGTGATGT TCATTTATGC CATCTTTGGG  
4501 ATGAACTGGT TTTCCAAAGT GAAGAGAGGC TCTGGGATTG ATGACATCTT CAACTTTGAC  
4561 ACTTTCTCGG GCAGCATGCT CTGCCTCTTC CAGATAACCA CTTCAGCCGG CTGGGATGCT  
4621 CTCCTCAACC CCATGCTGGA ATCAAAAGCC TCTTGCAATT CCTCCTCCCA AGAGAGCTGT  
4681 CAGCAGCCGC AGATAGCCAT AGTCTACTTC GTCAGCTACA TCATCATCTC CTTTCTCATT  
4741 GTGGTTAACA TGTACATAGC TGTGATTCTA GAGAACTTCA ACACAGCCAC AGAGGAGAGC  
4801 GAGGACCCCC TGGGCGAAGA CGACTTTGAG ATCTTCTATG AGATCTGGGA GAAAGTTGAC  
4861 CCCGAAGCAA CACAGTTCAT CCAGTACTCA TCCCTCTCTG ACTTCGCCGA CGCCCTGCCC  
4921 GAGCCGTTGC GTGTGGCCAA GCCCAACAGG TTTCAGTTTC TCATGATGGA CTGCCCCATG  
4981 GTGATGGGTG ATCGCCTCCA TTGCATGAT GTTCTCTTTG CTTTCACCAC CAGGGTCCTC  
5041 GGGAACTCCA GCGGCTTGA TACCATGAAA GCCATGATGG AGGAGAAGTT CATGGAGGCC  
5101 AATCCTTTCA AGAAGTTGTA CGAGCCCAT TGCACCACCA CAAAGAGGAA GGAGGAGGAG  
5161 GAATGTGCCG CTGTCATCCA GAGGCGCTAC CGGAGACACA TGGAGAAGAT GATCAAGCTG  
5221 AAGCTGAAAG GCAGGTCAAG TTCATCGCTC CAGGTGTTTT GCAATGGAGA CTGTCTAGC  
5281 TTGGATGTGC CCAAGATCAA GGTTCATTGT GACTGAAACC CCCACCTGCA CGCCTACCTC  
5341 ACAGCCTCAC AGCTCAGCCC CCAGCCTCTG GCGAACCAAGC GCGGACTCA CCGAACAGGC  
5401 CGTTCAACTT GTTTTTTTGG GTGAAAGAGG TGATAGGTTG GTGTCCATTT TTAATGATT  
5461 CTTGGAAAGA TTGAACGTCG GAACATGTTA GAAAGGACTG CCAAGGACAT CCACAGTAAC  
5521 GGAAGGCCTG AAGGACAGTT CAAATTATGT AAAGAAACGA GAAGGAAAGG TCACATGTCT  
5581 GTTCAGTTTT AAGTATGTGA CCTGCCACAT GTAGCTCCTT TGCATGTTAA GTGAGAAGTC  
5641 AAAACCCTGC CATAAGTAAA TAGCTTTGTT GCAGGTGTTT CTACCAGTGC TCCGGATTTG  
5701 GGTGTATGGC TCAAACCTGA AAGCATGACT CTGACTTGTC AGCACCCCAA CTTTCAGAAG  
5761 CTCTGATCTC TGTCCTAGGT GTTTGACAAA TAAATACATA AAANAAAAAA AAAAAAAAAA  
5821 AA

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**FIG. 7B-1**

Protein sequence of mNaN

Molecular Weight 201451.00 Daltons

1765 Amino Acids

198 Strongly Basic(+) Amino Acids (K,R)

177 Strongly Acidic(-) Amino Acids (D,E)

712 Hydrophobic Amino Acids (A,I,L,F,W,V)

453 Polar Amino Acids (N,C,Q,S,T,Y)

8.260 Isoelectric Point

22.540 Charge at PH 7.0

```

1  MEERYYPVIF PDERNFRPFT FDSLAAIEKR ITIQEKKKKS KDKAATEPQP RPQLDLKASR
21  KLPKLYGDVP PDLIAKPLED LDPFYKDHKT FMVLNKKRTI YRFSAKRALF ILQPFNPIRS
121 FMIRISVHSV FSMFIICTVI INCMEMANNS SVDSRPSSNI PRYVFIGIYV LEAVIKILAR
      DI-S1                               DI-S2
181 GFIVDEFSYL RDPHNWLDEI VIGTAIAPCF LGNKVMNLST LRTERVLRAL KAISVISGLK
      DI-S3                               DI-S4
241 VIVGALLRSV KKLVDVMVLT LECLSIKALV GOOLEFMGILS QKCINDDCGP NAFSNKDCPV
      DI-S5
301 KENDSEDFIM CGNWLGRASC PDGSTCNKTT FNPDYNYTNF DSEGHSPILAM FRVMTQDSWE
      DI-SS1 DI-SS2
361 KLYRQILRTS GIYFVEFFVY VIPLGSEYLL NLTAVVTMA YEEQNRNVAA ETEAKEKMFQ
      DI-S6
421 EAQQLLREEK EALVAMQIDR TSLNSLQASS FSPKKRKFFG SKTRKSFFMR GSKTARASAS
481 DSEDDASKNP QLLBQTKRLS QNLPVELFDE HVDPLHRQRA LSAVSILTIT MQEQEKSQEP
541 CFPCGKNLAS KYLVWECSPF WLCIKKVLQT INTDPTELA ITICIIVNTV FLAMEHHNMD
      DII-S1
601 NSLKDILKIG NHVETGIFIA EMCLKIIALD PYHYFRHGWN IEDSIVALVS LADVLEHKLK
      DII-S2                               DII-S3
661 KNLSPLASLR VLRVFKLAKS WPTLNTLIKI IGHSVGALGN LTVVLTIVVF IFSVVGMRLE
      DII-S4                               DII-S5
721 GAKFNKTCST SPESLRRWHM GDEYHSFLVV FRILCGEWIE NMWECMQEME GSPLCVIVEV
      DII-SS1 DII-SS2

```

**FIG. 7B-2**

**SUBSTITUTE SHEET (RULE 26)**

**FIG. 8A-1** Partial Human NaN Nucleotide Sequence

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TCCATTGTCATTGGAATAGCGATTGTGTCATATATCCAGGAATCACCATCAAACTATTGCCCC  
TGCGTACCTTCCGTGTGTTGAGAGCTTTGAAAGCAATTCAGTAGTTTCACGTCTGAAGGTCAT  
CGTGGGGGCCTTGCTACGCTCTGTGAAGAAGCTGGTCAACGTGATTATCCTCACCTTCTTTTGC  
CTCAGCATCTTTGCCCTGGTAGGTGAGCAGCTCTTCATGGGAAGTCTGAACCTGAAATGCATCT  
CGAGGGACTGTAAAAATATCAGTAACCCGGAAGCTTATGACCATTGCTTTGAAAAGAAAAGAAA  
ATTACCTGAATTCAAAATGTGTGGCATCTGGATGGGTAACAGTGCCTGTTCCATACAATATGA  
ATGTAAGCACACCAAAATTAATCCTGACTATAATTATACGAATTTTGACAACCTTTGGCTGGTCT  
TTTCTTGCCATGTTCCGGCTGATGACCCAAGATTCTGGGAGAAGCTTTATCAACAGACCCTGC  
GTAATACTGGGCTCTACTCAGTCTTCTTCTTCATTGTGGTCATTTTCTGGGCTCCTTCTACCTGA  
TTAACTTAACCTGGCTGTTGTTACCATGGCATATGAGGAGCAGAACAAGAATGTAGCTGCAG  
AGATAGAGGGCCAAGGAAAAAGATGTTTCAGGAAGCCGAGCAGCTGTTAAAGGAGGAAAAAGGAG  
GCTCTGGTTGCCATGGGAATTGACAGAAAGTTCACTTACTTCCCTTGAAAACATCATATTTTACCC  
CAAAAAAGAGAAAAGCTCTTTGGTAATAAGAAAAGGAAGTCCTTCTTTTGAGAGAGTCTGGGA  
AAGACCAGCCTCCTGGGTCAGATTCTGATGAAGATTGCCAAAAAAGCCACAGCTCCTAGAGC  
AAACCAAAACGACTGTCCAGAACTCTATCAVTGGACCACTTTGATGAGCATGGAGATCCTCTCCA  
AAGGCAGAGAGCACTGAGTGCTGTCAGCATCCTCACCATCACCATGAAGGAACAAGAAAAATC  
ACAAGAGCCTTGTCTCCCTTGTGGAGAAAACTGGCATCCAAGTACCTCGTGTGGAACTGTTGC  
CCCCAGTGGCTGTGCGTTAAGAAGGTCCTGAGAACTGTGATGACTGACCCGTTTACTGAGCTGG  
CCATCACCATCTGCATCATCATCAACACTGTCTTCTTGGCCATGGAGCATCACAAGATGGAGGC  
CAGTTTTGAGAAGATGTTGAATATAGGGAAATTTGGTTTTCACTAGCATTTTTATAGCAGAAATG  
TGCCTAAAAATCATTGCGCTCGATCCCTACCACTACTTTCGCGGAGGCTGGAACATTTTTGACA  
GCATTGTTGCTCTTCTGAGTTTTGCAGATGTAATGAACTGTGTACTTCAAAAGAGAAGCTGGCC  
ATTCTTGCGTTCCCTCAGAGTGCTCAGGGTCTTCAAGTTAGCCAAATCCTGGCCAACTTTGAAC  
ACACTAATTAAGATAATCGGCAACTCTGTCGGAGCCCTTGGAAAGCCTGACTGTGGTCTGGTCA  
TTGTGATCTTTATTTTCTCAGTAGTTGGCATGCAGCTTTTTGGCCGTAGCTTCAATTCCCAAAAG  
AGTCCAAAACTCTGTAACCCGACAGGCCCGACAGTCTCATGTTTACGGCACTGGCAGATGGGG  
GATTTCTGGCACTCCTTCTAGTGGTATTCCGCATCCTCTGCGGGGAATGGATCGAAAAATATGT  
GGGAATGTATGCAAGAAGCGAATGCATCATCATATTGTGTGTTATTGTCTTCATATTGATCAC  
GGTGATAGGAAAACTTGTGGTGCTCAACCTCTTCATTGCCTTACTGCTCAATTCCTTTAGCAAT  
GAGGAAAAGAAATGGAACTTAGAAGGAGAGGCCAGGAAAAGTAAAGTCCAGTTAGCACTGGA  
TCGATTCCGCGGGGCTTTTTGTTTTGTGAGACACACTCTTGAGCATTTCTGTACAAAGTGGTGCA  
GGAAGCAAACTTACCACAGCAAAAAAGAGGTGGCAGGAGGCTGTGCTGCACAAAGCAAAAGAC  
ATCATTCCCCTGGTCATGGAGATGAAAAGGGGCTCAGAGACCCAGGAGGAGCTTGGTATACTA  
ACCTCTGTACCAAAGACCCTGGGCGTCAGGCATGATTGGACTTGGTTGGCACCACTTGGCGAG  
GAGGAAGATGACGTTGAATTTCTGGTGAAGATAATGCACAGCGCATCACACAACCTGAGCCT  
GAACAACAGGCCTATGAGCTCCATCAGGAGAACAAGAAGCCCAAGAGCCAGAGAGTTCAAAAG  
TGTGGAAATTGACATGTTCTCTGAAGATGAGCCTCATCTGACCATAACAGGATCCCCGAAAGAA  
GTCTGATGTTACCAGTATACTATCAGAAATGTAGCACCATTGATCTTCAGGATGGCTTTGGATGG  
TTACCTGAGATGGTTCCCAAAAGCAACCAGAGAGATGTTTGCCCAAGGCTTTGGTTGCTGCT  
TTCCATGCTGTAGCGTGGACAAGAGAAAGCCTCCCTGGGTCAATTTGGTGGAACCTGCGGAAAA

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*FIG. 8A-2*

CCTGCTACCAAATAGTGAAACACAGCTGGTTTGAGAGCTTTATTATCTTTGTGATTCTGCTGAG  
CAGTGGGGCACTGATATTTGAAGATGTTACCTTGAGAACCAACCCAAAATCCAAGAATTACT  
AAATTGTACTGACATTATTTTACACATATTTTATCCTGGAGATGGTACTAAAATGGGTAGCC  
TTCGGATTTGAAAAGTATTTACCAAGTGCCTGGTGCTGCCTTGATTTCATCATTGTGATTGTCTC  
TGTGACCAACCTCATTAACCTAATGGAATTGAAGTCCTTCCGGACTCTACGAGCACTGAGGCCT  
CTTCGTGCGCTGTCCCAGTTTGAAGGAATGAAGGTGGTGGTCAATGCTCTCATAGGTGCCATAC  
CTGCCATTCTGAATGTTTTGCTTGTCTGCCTCATTTTCTGGCTCGTATTTTGTATTCTGGGAGTAT  
ACTTCTTTTCTGGAAAATTTGGGAAATGCATTAATGGAACAGACTCAGTTATAAATTATACCAT  
CATTACAAATAAAAAGTCAATGTGAAAAGTGGCAATTTCTCTTGGATCAACCAGAAAGTCAACTTT  
GACAATGTGGGAAATGCTTACCTCGCTCTGCTGCAAGTGGCAACATTTAAGGGCTGGATGGAT  
ATTATATATGCAGCTGTTGATTCCACAGAGAAAAGAACAACAGCCAGAGTTTGAGAGCAATTCA  
CTCGGTTACATTTACTTCGTAGTCTTTATCATCTTTGGCTCATTCTTCACTCTGAATCTCTTCATT  
GGCGTTATCATTGACAACCTCAACCAACAGCAGAAAAAGTTAGGTGGCCAAGACATTTTTATG  
ACAGAAGAACAGAAGAAATACTATAATGCAATGAAAAAATTAGGATCCAAAAAACCTCAAAA  
ACCCATTCCACGGCCTCTGAACAAATGTCAAGGTCTCGTGTTTCGACATAGTCACAAGCCAGATC  
TTTGACATCATCATCATAAGTCTCATTATCCTAAACATGATTAGCATGATGGCTGAATCATACA  
ACCAACCCAAAGCCATGAAATCCATCCTTGACCATCTCAACTGGGTCTTTGTGGTCATCTTTAC  
GTTAGAATGTCTCATCAAAATCTTTGCTTTGAGGCAATACTACTTCACCAATGGCTGGAATTTA  
TTTGA



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**FIG. 8B**

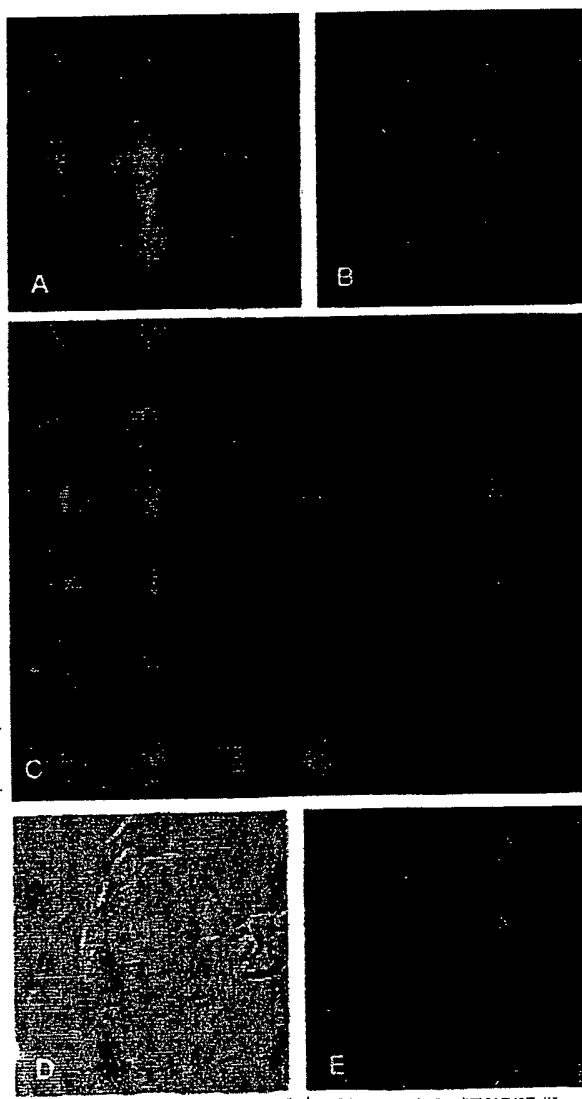
## Partial Human NaNAmino Acid Sequence

SI VIGIAIVSYI PGITIKLLPL RTFRVFRALK AISVVSRLKV IVGALLRSVK KLVNVITLTF  
FCLSIFALVG QQLFMGSLNL KCISRDCKNI SNPEAYDHCF EKKENSPEFK MCGIWMGNSA  
CSIQYECKHT KINPDYNYTN FDNFGWSFLA MFRLMTQDSW EKLYQQTLRT TGLYSVFFFI  
VVIFLGSFYI INLTAVVTM AYEEQNKNVA AEIEAKEKMF QEAQQLKEE KEALVAMGID  
RSSLTSLSTS YFTPKRKLK GNKKRKSFFL RESGKDQPPG SDSDEDCQKK PQLLEQTKRL  
SQNLSLDHFD EHGDPQRQR ALSAVSILTI TMKEQEKSQE PCLPCGENLA SKYLVWNCCP  
QWLCVKKVLR TVMTDPFTEL AITICIIINT VFLAMEHHKM EASF EKMLNI GNLVFTSIFI  
AEMCLKIAL DPYHYFRGW NIFDSIVALL SFADVMNCVL QKRSWPFLRS  
FRVLRVFKLAKSWPTLNTLI KIIGNSVGAL GSLTVVLVTV IFISVVGMMQ LFGRSFNSQK  
SPKLCNPTGP TVSCLRHWHM GDFWHSFLVV FRILCGEWIE NMWECMQEAN ASSSLCVTVF  
ILITVIGKLV VLNLFIALLL NSFSNEERNG NLEGEARKTK VQLALDRFRR AFCFVRHTLE  
HFCHKWCRKQ NLPQQKEVAG GCAAQSKDII PLVMEMKRGS ETQEELGILT SVPKTLGVRH  
DWTWLAPLAE EEDDVEFSGE DNAQRITQPE PEQQA YELHQ ENKKPTSQRVQSVEIDMFSE  
DEPHLTIQDP RKKSDVTSIL SECSTIDLQD GFGWLPEMVP KKQPERCLPK GFGCCFPCCS  
VDKRKPPWVI WWNLRKTCYQ IVKHSWFESF IIFVILLSSG ALIFEDVHLE NQPKIQELN  
CTDIIFTHIF ILEMVLKWVA FGFGKYFTSA WCCLDFIIVI VSVTTLINLM ELKSFRTLRA  
LRPLRALSQF EGMKVVVNAL IGAIPAILNV LLVCLIFWLV FCILGVYFFS GKFGKCIINGT  
DSVINYTIIT NKSQCESGNF SWINQKVNFD NVGNAYLALL QVATFKGWMD IYAAVDSTE  
KEQQPEFESN SLGYTYFVVF IIFGSFFTLN LFIGVIIDNF NQQQKKLGGQ DIFMTEEQKK  
YYNAMKKLGS KKPQKPIPRP LNKCCQLVFD IVTSQIFDII IISLIILNMI SMMAESYNQP  
KAMKSILDHL NWVFVVIIFTL ECLIKIFALR QYFTNGWNL FDCVVVLLSIV

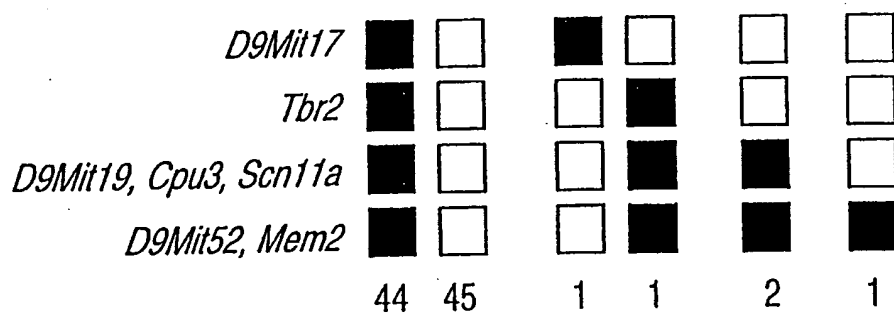
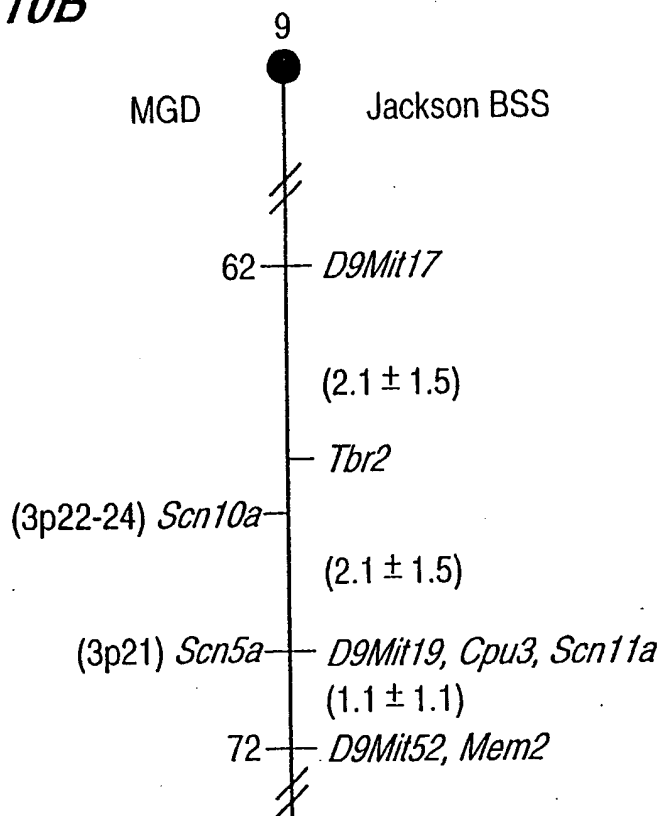
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**FIG. 9**

NaN immunostaining in DRG neurons



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**FIG. 10A****FIG. 10B**

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**FIG. 11A**

Sequence of human NaN cDNA. Open reading frame (cdc)  
Extends from position 31 (ATG) to the termination codon TGA at  
Position 5400.

```
1  ATCTGCTCAA GCCAGGAATC TCGGGTGAAG ATGGATGACA GATGCTACCC
51  AGTAATCTTT CCAGATGAGC GGAATTTCCG CCCCTTCACT TCCGACTCTC
101 TGGCTGCAAT TGAGAAGCGG ATTGCCATCC AAAAGGAGAA AAAGAAGTCT
151 AAAGACCAGA CAGGAGAAGT ACCCCAGCCT CGGCCTCAGC TTGACCTAAA
201 GGCCTCCAGG AAGTTGCCCCA AGCTCTATGG CGACATTCCT CGTGAGCTCA
251 TAGGAAAGCC TCTGGAAGAC TTGGACCCAT TCTACCGAAA TCATAAGACA
301 TTTATGGTGT TAAACAGAAA GAGGACAATC TACCGCTTCA GTGCCAAGCA
351 TGCCTTGTTT ATTTTGGGC CTTTCAATTC AATCAGAAGT TTAGCCATTA
401 GAGTCTCAGT CCATTCATTG TTCAGCATGT TCATTATCGG CACCGTTATC
451 ATCAACTGCG TGTTTCATGGC TACAGGGCCT GCTAAAAACA GCAACAGTAA
501 CAATACTGAC ATTGCAGAGT GTGTCTTCAC TGGGATTTAT ATTTTGAAG
551 CTTTGATTAA AATATTGGCA AGAGGTTTCA TTCTGGATGA GTTTTCTTTC
601 CTTGAGATC CATGGAAGT GCTGGACTCC ATTGTCATTG GAATAGCGAT
651 TGTGTCATAT ATTCCAGGAA TCACCATCAA ACTATTGCCC CTGCGTACCT
701 TCCGTGTGTT CAGAGCTTTG AAAGCAATTT CAGTAGTTTC ACGTCTGAAG
751 GTCATCGTGG GGGCCTTGCT ACGCTCTGTG AAGAAGCTGG TCAACGTGAT
801 TATCCTCACC TTCTTTTGCC TCAGCATCTT TGCCCTGGTA GGTCAGCAGC
851 TCTTCATGGG AAGTCTGAAC CTGAAATGCA TCTCGAGGGA CTGTAAAAAT
901 ATCAGTAACC CGGAAGCTTA TGACCATTGC TTTGAAAAGA AAGAAAATTC
951 ACCTGAATTC AAAATGTGTG GCATCTGGAT GGGTAACAGT GCCTGTTCCA
1001 TACAATATGA ATGTAAGCAC ACCAAAATTA ATCCTGACTA TAATTATACG
1051 AATTTTGACA ACTTTGGCTG GTCTTTTCTT GCCATGTTCC GGCTGATGAC
1101 CCAAGATTCC TGGGAGAAGC TTTATCAACA GACCCTGCGT ACTACTGGGC
1151 TCTACTCAGT CTTCTTCTTC ATTGTGGTCA TTTTCTGGG CTCCTTCTAC
1201 CTGATTAAC TAACCCTGGC TGTTGTTACC ATGGCATATG AGGAGCAGAA
```

**FIG. 11A-2** 23/28

1251 CAAGAATGTA GCTGCAGAGA TAGAGGCCAA GGAAAAGATG TTTCAGGAAG  
1301 CCCAGCAGCT GTTAAAGGAG GAAAAGGAGG CTCTGGTTGC CATGGGAATT  
1351 GACAGAAGTT CACTTACTTC CCTTGAAACA TCATATTTTA CCCCCAAAAA  
1401 GAGAAAGCTC TTTGGTAATA AGAAAAGGAA GTCCTTCTTT TTGAGAGAGT  
1451 CTGGGAAAGA CCAGCCTCCT GGGTCAGATT CTGATGAAGA TTGCCAAAAA  
1501 AAGCCACAGC TCCTAGAGCA AACCAAACGA CTGTCCCAGA ATCTATCACT  
1551 GGACCACTTT GATGAGCATG GAGATCCTCT CCAAAGGCAG AGAGCACTGA  
1601 GTGCTGTCAG CATCCTCACC ATCACCATGA AGGAACAAGA AAAATCACAA  
1651 GAGCCTTGTC TCCCTTGTGG AGAAAACCTG GCATCCAAGT ACCTCGTGTG  
1701 GAACTGTTGC CCCAGTGGC TGTGCGTTAA GAAGGTCCTG AGAACTGTGA  
1751 TGA CTGACCC GTTTACTGAG CTGGCCATCA CCATCTGCAT CATCATCAAC  
1801 ACTGTCTTCT TGGCCATGGA GCATGACAAG ATGGAGGCCA GTTTTGAGAA  
1851 GATGTTGAAT ATAGGGAATT TGGTTTTTAC TAGCATTTTT ATAGCAGAAA  
1901 TGTGCCTAAA AATCATTGCG CTCGATCCCT ACCACTACTT TCGCCGAGGC  
1951 TGGAACATTT TTGACAGCAT TGTGCTCTT CTGAGTTTTG CAGATGTAAT  
2001 GAACTGTGTA CTTCAAAAGA GAAGCTGGCC ATTCTTGCGT TCCTTCAGAG  
2051 TGCTCAGGGT CTTCAAGTTA GCCAAATCCT GGCCAACCTT GAACACACTA  
2101 ATTAAGATAA TCGGCAACTC TGTCGGAGCC CTTGGAAGCC TGACTGTGGT  
2151 CCTGGTCATT GTGATCTTTA TTTTCTCAGT AGTTGGCATG CAGCTTTTTG  
2201 GCCGTAGCTT CAATCCCAA AAGAGTCCAA AACTCTGTAA CCCGACAGGC  
2251 CCGACAGTCT CATGTTTACG GCACTGGCAC ATGGGGGATT TCTGGCACTC  
2301 CTTCTAGTG GTATTCCGCA TCCTCTGCGG GGAATGGATC GAAAATATGT  
2351 GGAATGTAT GCAAGAAGCG AATGCATCAT CATCATTGTG TGTTATTGTC  
2401 TTCATATTGA TCACGGTGAT AGGAAAACCT GTGGTGCTCA ACCTCTTCAT  
2451 TGCCTTACTG CTCAATTCCT TTAGCAATGA GGAAAGAAAT GGAAACTTAG  
2501 AAGGAGAGGC CAGGAAAACT AAAGTCCAGT TAGCACTGGA TCGATTCCGC  
2551 CGGGCTTTTT GTTTGTGAG ACACACTCTT GAGCATTTCT GTCACAAGTG

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**FIG. 11A-3**

2601 GTGCAGGAAG CAAACTTAC CACAGCAAAA AGAGGTGGCA GGAGGCTGTG  
2651 CTGCACAAAG CAAAGACATC ATTCCCCTGG TCATGGAGAT GAAAAGGGGC  
2701 TCAGAGACCC AGGAGGAGCT TGGTATACTA ACCTCTGTAC CAAAGACCCT  
2751 GGGCGTCAGG CATGATTGGA CTTGGTTGGC ACCACTTGCG GAGGAGGAAG  
2801 ATGACGTTGA ATTTTCTGGT GAAGATAATG CACAGCGCAT CACACAACCT  
2851 GAGCCTGAAC AACAGGCCTA TGAGCTCCAT CAGGAGAACA AGAAGCCCAC  
2901 GAGCCAGAGA GTTCAAAGTG TGGAAATTGA CATGTTCTCT GAAGATGAGC  
2951 CTCATCTGAC CATAcAGGAT CCCCgAAAGA AGTCTGATGT TACCAGTATA  
3001 CTATCAGAAT GTAGCACCAT TGATCTTCAG GATGGCTTTG GATGGTTACC  
3051 TGAGATGGTT CCCAAAAAGC AACCAGAGAG ATGTTTGCCC AAAGGCTTTG  
3101 GTTGCTGCTT TCCATGCTGT AGCGTGGACA AGAGAAAGCC TCCCTGGGTC  
3151 ATTTGGTGGA ACCTGCGGAA AACCTGCTAC CAAATAGTGA AACACAGCTG  
3201 GTTTGAGAGC TTTATTATCT TTGTGATTCT GCTGAGCAGT GGGGCACTGA  
3251 TATTTGAAGA TGTTCACTT GAGAACCAAC CCAAAATCCA AGAATTACTA  
3301 AATTGTACTG ACATTATTTT TACACATATT TTTATCCTGG AGATGGTACT  
3351 AAAATGGGTA GCCTTCGGAT TTGGAAGTA TTTCAACAGT GCCTGGTGCT  
3401 GCCTTGATTT CATCATTGTG ATTGTCTCTG TGACCACCCT CATTAACTTA  
3451 ATGGAATTGA AGTCCTCCG GACTCTACGA GCACTGAGGC CTCTTCGTGC  
3501 GCTGTCCCAG TTTGAAGGAA TGAAGGTGGT GGTCAATGCT CTCATAGGTG  
3551 CCATACCTGC CATTCTGAAT GTTTTGCTTG TCTGCCTCAT TTTCTGGCTC  
3601 GTATTTTGTA TTCTGGGAGT ATACTTCTTT TCTGGAAAAT TTGGGAAATG  
3651 CATTAAATGA ACAGACTCAG TTATAAATTA TACCATCATT ACAAATAAAA  
3701 GTCAATGTGA AAGTGGCAAT TTCTCTTGA TCAACCAGAA AGTCAACTTT  
3751 GACAATGTGG GAAATGCTTA CCTCGCTCTG CTGCAAGTGG CAACATTTAA  
3801 GGGCTGGATG GATATTATAT ATGCAGCTGT TGATTCCACA GAGAAAGAAC  
3851 AACAGCCAGA GTTTGAGAGC AATTCACCTG GTTACATTTA CTTCGTAGTC  
3901 TTTATCATCT TTGGCTCATT CTTCACTCTG AATCTCTTCA TTGGCGTTAT  
3951 CATTGACAAC TTCAACCAAC AGCAGAAAAA GTTAGGTGGC CAAGACATTT

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**FIG. 11A-4**

4001 TTATGACAGA AGAACAGAAG AAATACTATA ATGCAATGAA AAAATTAGGA  
4051 TCCAAAAAAC CTCAAAAACC CATTCCACGG CCTCTGAACA AATGTCAAGG  
4101 TCTCGTGTTT GACATAGTCA CAAGCCAGAT CTTTGACATC ATCATCATAA  
4151 GTCTCATTAT CCTAAACATG ATTAGCATGA TGGCTGAATC ATACAACCAA  
4201 CCCAAAGCCA TGAAATCCAT CCTTGACCAT CTCAACTGGG TCTTTGTGGT  
4251 CATCTTTACG TTAGAATGTC TCATCAAAAT CTTTGCTTTG AGGCAATACT  
4301 ACTTCACCAA TGGCTGGAAT TTATTTGACT GTGTGGTCGT GCTTCTTTCC  
4351 ATTGTTAGTA CAATGATTTC TACCTTGAA AATCAGGAGC ACATTCCTTT  
4401 CCCTCCGACG CTCTTCAGAA TTGTCCGCTT GGCTCGGATT GGCCGAATCC  
4451 TGAGGCTTGT CCGGGCTGCA CGAGGAATCA GGAATCTCCT CTTTGCTCTG  
4501 ATGATGTCGC TTCCTTCTCT GTTCAACATT GGTCTTCTAC TCTTTCTGAT  
4551 TATGTTTATC TATGCCATTG TGGGTATGAA CTGGTTTTCC AAAGTGAATC  
4601 CAGAGTCTGG AATCGATGAC ATATTCAACT TCAAGACTTT TGCCAGCAGC  
4651 ATGCTCTGTC TCTTCCAGAT AAGCACATCA GCAGGTTGGG ATTCCCTGCT  
4701 CAGCCCCATG CTGCGATCAA AAGAATCATG TAACTCTTCC TCAGAAAACCT  
4751 GCCACCTCCC TGGCATAGCC ACATCCTACT TTGTCAGTTA CATTATCATC  
4801 TCCTTTCTCA TTGTTGTCAA CATGTACATT GCTGTGATTT TAGAGAACTT  
4851 CAATACAGCC ACTGAAGAAA GTGAGGACCC TTTGGGTGAA GATGACTTTG  
4901 ACATATTTTA TGAAGTGTGG GAAAAGTTTG ACCCAGAAGC AACACAATTT  
4951 ATCAAATATT CTGCCCTTTC TGACTTTGCT GATGCCTTGC CTGAGCCTTT  
5001 GCGTGTCGCA AAGCCAAATA AATATCAATT TCTAGTAATG GACTTGCCCA  
5051 TGGTGAGTGA AGATCGCCTC CACTGCATGG ATATTCTTTT CGCCTTCACC  
5101 GCTAGGGTAC TCGGTGGCTC TGATGGCCTA GATAGTATGA AAGCAATGAT  
5151 GGAAGAGAAG TTCATGGAAG CCAATCCTCT CAAGAAGTTG TATGAACCCA  
5201 TAGTCACCAC CACCAAGAGA AAGGAAGAGG AAAGAGGTGC TGCTATTATT  
5251 CAAAAGGCCT TTCGAAAGTA CATGATGAAG GTGACCAAGG GTGACCAAGG  
5301 TGACCAAAAT GACTTGGAAG ACGGGCCTCA TTCACCACTC CAGACTCTTT

**FIG. IIA-5**

5351 GCAATGGAGA CTTGTCTAGC TTTGGGGTGG CCAAGGGCAA GGTCCACTGT  
5401 GACTGAGCCC TCACCTCCAC GCCTACCTCA TAGCTTCACA GCCTTGCCTT  
5451 CAGCCTCTGA GCTCCAGGGG TCAGCAGCTT AGTGTATCAA CAGGGAGTGG  
5501 ATTCACCAAA TTAGCCATTC CATTTTCTTT TCTGGCTAAA ATAAATGATA  
5551 TTTCAATTTC ATTTTAAATG ATACTTACAG AGATATAAGA TAAGGCTACT  
5601 TGACAACCAG TGGTACTATT ATAATAAGGA AGAAGACACC AGGAAGGACT  
5651 GTAAAAGGAC ATACCAATTT TAGGATTGAA ATAGTTCAGG CCGGGCGCAG  
5701 TGGCTCATGC CTGTAATCCC AGCACTTTGA GAGGCCAAGG CAGGTGGATC  
5751 ACGAGGTCAA GAGATCGAGA CCATCCTGGC CAACATGATG AAACTCCGTC  
5801 TCTCTAAAAA TACAAAAATT AGCTGGGCAT GGTGGCGTGC GCCTGTAGTC  
5851 CCACTACTTG



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**FIG. 11B**

Sequence of human NaN.

1 MDDRCYPVIF PDERNFRPFT SDSLAAIEKR IAIQKEKKKS KDQTGEVPQP  
51 RPQLDLKASR KLPKLYGDIP RELIGKPLED LDPFYRNHKT FMVLNRKRTI  
101 YRFSAKHALF IFGPFNSIRS LAIRSVHSL FSMFIIGTVI INCVFMTATGP  
151 AKNSNSNNTD IAECVFTGIY IFEALIKILA RGFILDEFSS LRDPWNWLDS  
201 IVIGIAIVSY IPGITIKLLP LRTFRVFRAL KAISVVSRLK VIVGALLRSV  
251 KKLNVNIILT FFCLSIFALV GQQLFMGSLN LKCISRDCKN ISNPEAYDHC  
301 FEKKENSPEF KMCGIWMGNS ACSIQYECKH TKINPDYNYT NFDNFGWSFL  
351 AMFRLMTQDS WEKLYQOTLR TTGLYSVFFF IVVIFLGSFY LINLTLAVVT  
401 MAYEEQNKNV AAEIEAKEKM FQEAQQLKE EKEALVAMGI DRSSLTSLET  
451 SYFTPKKRKL FGNKKRKSFF LRESGKDQPP GSDSDEDCQK KPQLLEQTKR  
501 LSQNLSLDHF DEHGDPLQRQ RALSAVSILT ITMKEQESQ EPCLPCGENL  
551 ASKYLWNCC PQWLCVKKVL RTVMTDPFTE LAITICIIIN TVFLAMEHHK  
601 MEASF EKMLN IGNLVFTSIF IAEMCLKIIA LDPYHYFRRG WNIFDSIVAL  
651 LSFADVMNCV LQKRSWPFLR SFRVLRVFKL AKSWPTLNTL IKIIGNSVGA  
701 LGSLTVLVI VIFIFSVVGM QLFGRSFNSQ KSPKLCNPTG PTVSCLRHHW  
751 MGDFWHSFLV VFRILCGEWI ENMWEQMSEA NASSSLCVIV FILITVIGKL  
801 VVLNLFIAL LNSFSNEERN GNLEGEARKT KVQLALDRFR RAFCFVRHTL  
851 EHFCHKWCRK QNLPQQKEVA GGCAAQSKDI IPLVMEMKRG SETQEELGIL  
901 TSVPKTLGVR HDWTWLAPLA EEEDDVEFSG EDNAQRITQP EPEQQAYELH  
951 QENKKPTSQR VQSVEIDMFS EDEPHLTIQD PRKKS DVTSI LSECSTIDLQ  
1001 DGFGWLPEMV PPKQPERCLP KGFGCCFPCC SVDKRKPPWV IWWNLRKTCY  
1051 QIVKHSWFES FIIFVILLSS GALIFEDVHL ENQPKIQELL NCTDIIFTHI  
1101 FILEMVLKWW AFGFGKYFTS AWCLDFIIV IVSVTTLINL MELKSFRTLR  
1151 ALRPLRALSQ FEGMKVVVNA LIGAIPAILN VLLVCLIFWL VFCILGVYFF

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*FIG. 11B-2*

1201 SGKFGKACING TDSVINYTII TNKSQCESGN FSWINQKVN F DNVGNAYLAL  
1251 LQVATFKGWM DIIYAAVDST EKEQQPEFES NSLGYIYFVV FIIFGSFFTL  
1301 NLFIGVIIDN FNQQQKKLGG QDIFMTEEQK KYYNAMKKLG SKKPQKPIPR  
1351 PLNKCQGLVF DIVTSQIFDI IIISLIILNM ISMMAESYNQ PKAMKSILDH  
1401 LNWVFWVIFT LECLIKIFAL RQYYFTNGWN LFDCVVVLLS IVSTMISTLE  
1451 NQEHIPFPPT LFRIVRLARI GRILRLVRAA RGIRTLLFAL MMSLPSLFNI  
1501 GLLLFLIMFI YAILGMNWFS KVPESGIDD IFNFKTFASS MLCLFQISTS  
1551 AGWDSLSPM LRSKESCNSS SENCHLPGIA TSYFVSYIII SFLIVNMYI  
1601 AVILENFNTA TEESEDPLGE DDFDIFYEVW EKFDPEATQF IKYSALSDFA  
1651 DALPEPLRVA KPNKYQFLVM DLPVSEDRL HCMDILFAFT ARVLGGSDDL  
1701 DSMKAMMEK FMEANPLKKL YEPIVTTTKR KEEERGAAII QKAFRKYMMK  
1751 VTKGDQGDQN DLNGPHSPL QTLNCGDLSS FGVAKGVHC D.

&lt;400&gt; 38

Met Trp Xaa Cys Met Glu Val

1

5

&lt;210&gt; 39

&lt;211&gt; 22

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

<223> Description of Artificial Sequence: rat NaN  
forward primer

&lt;400&gt; 39

ccctgctgcg ctcggtgaag aa

22

&lt;210&gt; 40

&lt;211&gt; 20

&lt;212&gt; PRT

&lt;213&gt; Artificial Sequence

&lt;220&gt;

<223> Description of Artificial Sequence: a.a. seq.  
used to derive epitope for polyclonal antibody

&lt;400&gt; 40

Cys Gly Pro Asn Pro Ala Ser Asn Lys Asp Cys Phe Glu Lys Glu Lys

1

5

10

15

Asp Ser Glu Asp

20

&lt;210&gt; 41

&lt;211&gt; 5860

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; CDS

&lt;222&gt; (31)..(5403)

&lt;223&gt; full length cDNA sequence for human NaN

&lt;400&gt; 41

atctgctcaa gccaggaatc tcgggtgaag atg gat gac aga tgc tac cca gta 54

Met Asp Asp Arg Cys Tyr Pro Val

1

5

atc ttt cca gat gag cgg aat ttc cgc ccc ttc act tcc gac tct ctg 102

Ile Phe Pro Asp Glu Arg Asn Phe Arg Pro Phe Thr Ser Asp Ser Leu

10

15

20

gct gca att gag aag cgg att gcc atc caa aag gag aaa aag aag tct 150

Ala Ala Ile Glu Lys Arg Ile Ala Ile Gln Lys Glu Lys Lys Lys Ser

25	30	35	40	
aaa gac cag aca gga gaa gta ccc cag cct cgg cct cag ctt gac cta				198
Lys Asp Gln Thr Gly Glu Val Pro Gln Pro Arg Pro Gln Leu Asp Leu	45	50	55	
aag gcc tcc agg aag ttg ccc aag ctc tat ggc gac att cct cgt gag				246
Lys Ala Ser Arg Lys Leu Pro Lys Leu Tyr Gly Asp Ile Pro Arg Glu	60	65	70	
ctc ata gga aag cct ctg gaa gac ttg gac cca ttc tac cga aat cat				294
Leu Ile Gly Lys Pro Leu Glu Asp Leu Asp Pro Phe Tyr Arg Asn His	75	80	85	
aag aca ttt atg gtg tta aac aga aag agg aca atc tac cgc ttc agt				342
Lys Thr Phe Met Val Leu Asn Arg Lys Arg Thr Ile Tyr Arg Phe Ser	90	95	100	
gcc aag cat gcc ttg ttc att ttt ggg cct ttc aat tca atc aga agt				390
Ala Lys His Ala Leu Phe Ile Phe Gly Pro Phe Asn Ser Ile Arg Ser	105	110	115	120
tta gcc att aga gtc tca gtc cat tca ttg ttc agc atg ttc att atc				438
Leu Ala Ile Arg Val Ser Val His Ser Leu Phe Ser Met Phe Ile Ile	125	130	135	
ggc acc gtt atc atc aac tgc gtg ttc atg gct aca ggg cct gct aaa				486
Gly Thr Val Ile Ile Asn Cys Val Phe Met Ala Thr Gly Pro Ala Lys	140	145	150	
aac agc aac agt aac aat act gac att gca gag tgt gtc ttc act ggg				534
Asn Ser Asn Ser Asn Asn Thr Asp Ile Ala Glu Cys Val Phe Thr Gly	155	160	165	
att tat att ttt gaa gct ttg att aaa ata ttg gca aga ggt ttc att				582
Ile Tyr Ile Phe Glu Ala Leu Ile Lys Ile Leu Ala Arg Gly Phe Ile	170	175	180	
ctg gat gag ttt tct ttc ctt cga gat cca tgg aac tgg ctg gac tcc				630
Leu Asp Glu Phe Ser Phe Leu Arg Asp Pro Trp Asn Trp Leu Asp Ser	185	190	195	200
att gtc att gga ata gcg att gtg tca tat att cca gga atc acc atc				678
Ile Val Ile Gly Ile Ala Ile Val Ser Tyr Ile Pro Gly Ile Thr Ile	205	210	215	
aaa cta ttg ccc ctg cgt acc ttc cgt gtg ttc aga gct ttg aaa gca				726
Lys Leu Leu Pro Leu Arg Thr Phe Arg Val Phe Arg Ala Leu Lys Ala	220	225	230	
att tca gta gtt tca cgt ctg aag gtc atc gtg ggg gcc ttg cta cgc				774
Ile Ser Val Val Ser Arg Leu Lys Val Ile Val Gly Ala Leu Leu Arg	235	240	245	
tct gtg aag aag ctg gtc aac gtg att atc ctc acc ttc ttt tgc ctc				822
Ser Val Lys Lys Leu Val Asn Val Ile Ile Leu Thr Phe Phe Cys Leu				

250	255	260	
agc atc ttt gcc ctg gta ggt cag cag ctc ttc atg gga agt ctg aac			870
Ser Ile Phe Ala Leu Val Gly Gln Gln Leu Phe Met Gly Ser Leu Asn			
265	270	275	280
ctg aaa tgc atc tcg agg gac tgt aaa aat atc agt aac ccg gaa gct			918
Leu Lys Cys Ile Ser Arg Asp Cys Lys Asn Ile Ser Asn Pro Glu Ala			
	285	290	295
tat gac cat tgc ttt gaa aag aaa gaa aat tca cct gaa ttc aaa atg			966
Tyr Asp His Cys Phe Glu Lys Lys Glu Asn Ser Pro Glu Phe Lys Met			
	300	305	310
tgt ggc atc tgg atg ggt aac agt gcc tgt tcc ata caa tat gaa tgt			1014
Cys Gly Ile Trp Met Gly Asn Ser Ala Cys Ser Ile Gln Tyr Glu Cys			
	315	320	325
aag cac acc aaa att aat cct gac tat aat tat acg aat ttt gac aac			1062
Lys His Thr Lys Ile Asn Pro Asp Tyr Asn Tyr Thr Asn Phe Asp Asn			
	330	335	340
ttt ggc tgg tct ttt ctt gcc atg ttc cgg ctg atg acc caa gat tcc			1110
Phe Gly Trp Ser Phe Leu Ala Met Phe Arg Leu Met Thr Gln Asp Ser			
	345	350	355
tgg gag aag ctt tat caa cag acc ctg cgt act act ggg ctc tac tca			1158
Trp Glu Lys Leu Tyr Gln Gln Thr Leu Arg Thr Thr Gly Leu Tyr Ser			
	365	370	375
gtc ttc ttc ttc att gtg gtc att ttc ctg ggc tcc ttc tac ctg att			1206
Val Phe Phe Phe Ile Val Val Ile Phe Leu Gly Ser Phe Tyr Leu Ile			
	380	385	390
aac tta acc ctg gct gtt gtt acc atg gca tat gag gag cag aac aag			1254
Asn Leu Thr Leu Ala Val Val Thr Met Ala Tyr Glu Glu Gln Asn Lys			
	395	400	405
aat gta gct gca gag ata gag gcc aag gaa aag atg ttt cag gaa gcc			1302
Asn Val Ala Ala Glu Ile Glu Ala Lys Glu Lys Met Phe Gln Glu Ala			
	410	415	420
cag cag ctg tta aag gag gaa aag gag gct ctg gtt gcc atg gga att			1350
Gln Gln Leu Leu Lys Glu Glu Lys Glu Ala Leu Val Ala Met Gly Ile			
	425	430	435
gac aga agt tca ctt act tcc ctt gaa aca tca tat ttt acc cca aaa			1398
Asp Arg Ser Ser Leu Thr Ser Leu Glu Thr Ser Tyr Phe Thr Pro Lys			
	445	450	455
aag aga aag ctc ttt ggt aat aag aaa agg aag tcc ttc ttt ttg aga			1446
Lys Arg Lys Leu Phe Gly Asn Lys Lys Arg Lys Ser Phe Phe Leu Arg			
	460	465	470
gag tct ggg aaa gac cag cct cct ggg tca gat tct gat gaa gat tgc			1494
Glu Ser Gly Lys Asp Gln Pro Pro Gly Ser Asp Ser Asp Glu Asp Cys			

475	480	485	
caa aaa aag cca cag ctc cta gag caa acc aaa cga ctg tcc cag aat			1542
Gln Lys Lys Pro Gln Leu Leu Glu Gln Thr Lys Arg Leu Ser Gln Asn			
490	495	500	
cta tca ctg gac cac ttt gat gag cat gga gat cct ctc caa agg cag			1590
Leu Ser Leu Asp His Phe Asp Glu His Gly Asp Pro Leu Gln Arg Gln			
505	510	515	520
aga gca ctg agt gct gtc agc atc ctc acc atc acc atg aag gaa caa			1638
Arg Ala Leu Ser Ala Val Ser Ile Leu Thr Ile Thr Met Lys Glu Gln			
	525	530	535
gaa aaa tca caa gag cct tgt ctc cct tgt gga gaa aac ctg gca tcc			1686
Glu Lys Ser Gln Glu Pro Cys Leu Pro Cys Gly Glu Asn Leu Ala Ser			
	540	545	550
aag tac ctc gtg tgg aac tgt tgc ccc cag tgg ctg tgc gtt aag aag			1734
Lys Tyr Leu Val Trp Asn Cys Cys Pro Gln Trp Leu Cys Val Lys Lys			
	555	560	565
gtc ctg aga act gtg atg act gac ccg ttt act gag ctg gcc atc acc			1782
Val Leu Arg Thr Val Met Thr Asp Pro Phe Thr Glu Leu Ala Ile Thr			
	570	575	580
atc tgc atc atc atc aac act gtc ttc ttg gcc atg gag cat cac aag			1830
Ile Cys Ile Ile Ile Asn Thr Val Phe Leu Ala Met Glu His His Lys			
	585	590	595
atg gag gcc agt ttt gag aag atg ttg aat ata ggg aat ttg gtt ttc			1878
Met Glu Ala Ser Phe Glu Lys Met Leu Asn Ile Gly Asn Leu Val Phe			
	605	610	615
act agc att ttt ata gca gaa atg tgc cta aaa atc att gcg ctc gat			1926
Thr Ser Ile Phe Ile Ala Glu Met Cys Leu Lys Ile Ile Ala Leu Asp			
	620	625	630
ccc tac cac tac ttt cgc cga ggc tgg aac att ttt gac agc att gtt			1974
Pro Tyr His Tyr Phe Arg Arg Gly Trp Asn Ile Phe Asp Ser Ile Val			
	635	640	645
gct ctt ctg agt ttt gca gat gta atg aac tgt gta ctt caa aag aga			2022
Ala Leu Leu Ser Phe Ala Asp Val Met Asn Cys Val Leu Gln Lys Arg			
	650	655	660
agc tgg cca ttc ttg cgt tcc ttc aga gtg ctc agg gtc ttc aag tta			2070
Ser Trp Pro Phe Leu Arg Ser Phe Arg Val Leu Arg Val Phe Lys Leu			
	665	670	675
gcc aaa tcc tgg cca act ttg aac aca cta att aag ata atc ggc aac			2118
Ala Lys Ser Trp Pro Thr Leu Asn Thr Leu Ile Lys Ile Ile Gly Asn			
	685	690	695
tct gtc gga gcc ctt gga agc ctg act gtg gtc ctg gtc att gtg atc			2166
Ser Val Gly Ala Leu Gly Ser Leu Thr Val Val Leu Val Ile Val Ile			

700	705	710	
ttt att ttc tca gta gtt ggc atg cag ctt ttt ggc cgt agc ttc aat	2214		
Phe Ile Phe Ser Val Val Gly Met Gln Leu Phe Gly Arg Ser Phe Asn			
715 720 725			
tcc caa aag agt cca aaa ctc tgt aac ccg aca ggc ccg aca gtc tca	2262		
Ser Gln Lys Ser Pro Lys Leu Cys Asn Pro Thr Gly Pro Thr Val Ser			
730 735 740			
tgt tta cgg cac tgg cac atg ggg gat ttc tgg cac tcc ttc cta gtg	2310		
Cys Leu Arg His Trp His Met Gly Asp Phe Trp His Ser Phe Leu Val			
745 750 755 760			
gta ttc cgc atc ctc tgc ggg gaa tgg atc gaa aat atg tgg gaa tgt	2358		
Val Phe Arg Ile Leu Cys Gly Glu Trp Ile Glu Asn Met Trp Glu Cys			
765 770 775			
atg caa gaa gcg aat gca tca tca tca ttg tgt gtt att gtc ttc ata	2406		
Met Gln Glu Ala Asn Ala Ser Ser Ser Leu Cys Val Ile Val Phe Ile			
780 785 790			
ttg atc acg gtg ata gga aaa ctt gtg gtg ctc aac ctc ttc att gcc	2454		
Leu Ile Thr Val Ile Gly Lys Leu Val Val Leu Asn Leu Phe Ile Ala			
795 800 805			
tta ctg ctc aat tcc ttt agc aat gag gaa aga aat gga aac tta gaa	2502		
Leu Leu Leu Asn Ser Phe Ser Asn Glu Glu Arg Asn Gly Asn Leu Glu			
810 815 820			
gga gag gcc agg aaa act aaa gtc cag tta gca ctg gat cga ttc cgc	2550		
Gly Glu Ala Arg Lys Thr Lys Val Gln Leu Ala Leu Asp Arg Phe Arg			
825 830 835 840			
cgg gct ttt tgt ttt gtg aga cac act ctt gag cat ttc tgt cac aag	2598		
Arg Ala Phe Cys Phe Val Arg His Thr Leu Glu His Phe Cys His Lys			
845 850 855			
tgg tgc agg aag caa aac tta cca cag caa aaa gag gtg gca gga ggc	2646		
Trp Cys Arg Lys Gln Asn Leu Pro Gln Gln Lys Glu Val Ala Gly Gly			
860 865 870			
tgt gct gca caa agc aaa gac atc att ccc ctg gtc atg gag atg aaa	2694		
Cys Ala Ala Gln Ser Lys Asp Ile Ile Pro Leu Val Met Glu Met Lys			
875 880 885			
agg ggc tca gag acc cag gag gag ctt ggt ata cta acc tct gta cca	2742		
Arg Gly Ser Glu Thr Gln Glu Glu Leu Gly Ile Leu Thr Ser Val Pro			
890 895 900			
aag acc ctg ggc gtc agg cat gat tgg act tgg ttg gca cca ctt gcg	2790		
Lys Thr Leu Gly Val Arg His Asp Trp Thr Trp Leu Ala Pro Leu Ala			
905 910 915 920			
gag gag gaa gat gac gtt gaa ttt tct ggt gaa gat aat gca cag cgc	2838		
Glu Glu Glu Asp Asp Val Glu Phe Ser Gly Glu Asp Asn Ala Gln Arg			

	925	930	935	
atc aca caa cct gag cct gaa caa cag gcc tat gag ctc cat cag gag				2886
Ile Thr Gln Pro Glu Pro Glu Gln Gln Ala Tyr Glu Leu His Gln Glu				
	940	945	950	
aac aag aag ccc acg agc cag aga gtt caa agt gtg gaa att gac atg				2934
Asn Lys Lys Pro Thr Ser Gln Arg Val Gln Ser Val Glu Ile Asp Met				
	955	960	965	
ttc tct gaa gat gag cct cat ctg acc ata cag gat ccc cga aag aag				2982
Phe Ser Glu Asp Glu Pro His Leu Thr Ile Gln Asp Pro Arg Lys Lys				
	970	975	980	
tct gat gtt acc agt ata cta tca gaa tgt agc acc att gat ctt cag				3030
Ser Asp Val Thr Ser Ile Leu Ser Glu Cys Ser Thr Ile Asp Leu Gln				
	985	990	995	1000
gat ggc ttt gga tgg tta cct gag atg gtt ccc aaa aag caa cca gag				3078
Asp Gly Phe Gly Trp Leu Pro Glu Met Val Pro Lys Lys Gln Pro Glu				
	1005	1010	1015	
aga tgt ttg ccc aaa ggc ttt ggt tgc tgc ttt cca tgc tgt agc gtg				3126
Arg Cys Leu Pro Lys Gly Phe Gly Cys Cys Phe Pro Cys Cys Ser Val				
	1020	1025	1030	
gac aag aga aag cct ccc tgg gtc att tgg tgg aac ctg cgg aaa acc				3174
Asp Lys Arg Lys Pro Pro Trp Val Ile Trp Trp Asn Leu Arg Lys Thr				
	1035	1040	1045	
tgc tac caa ata gtg aaa cac agc tgg ttt gag agc ttt att atc ttt				3222
Cys Tyr Gln Ile Val Lys His Ser Trp Phe Glu Ser Phe Ile Ile Phe				
	1050	1055	1060	
gtg att ctg ctg agc agt ggg gca ctg ata ttt gaa gat gtt cac ctt				3270
Val Ile Leu Leu Ser Ser Gly Ala Leu Ile Phe Glu Asp Val His Leu				
	1065	1070	1075	1080
gag aac caa ccc aaa atc caa gaa tta cta aat tgt act gac att att				3318
Glu Asn Gln Pro Lys Ile Gln Glu Leu Leu Asn Cys Thr Asp Ile Ile				
	1085	1090	1095	
ttt aca cat att ttt atc ctg gag atg gta cta aaa tgg gta gcc ttc				3366
Phe Thr His Ile Phe Ile Leu Glu Met Val Leu Lys Trp Val Ala Phe				
	1100	1105	1110	
gga ttt gga aag tat ttc acc agt gcc tgg tgc tgc ctt gat ttc atc				3414
Gly Phe Gly Lys Tyr Phe Thr Ser Ala Trp Cys Cys Leu Asp Phe Ile				
	1115	1120	1125	
att gtg att gtc tct gtg acc acc ctc att aac tta atg gaa ttg aag				3462
Ile Val Ile Val Ser Val Thr Thr Leu Ile Asn Leu Met Glu Leu Lys				
	1130	1135	1140	
tcc ttc cgg act cta cga gca ctg agg cct ctt cgt gcg ctg tcc cag				3510
Ser Phe Arg Thr Leu Arg Ala Leu Arg Pro Leu Arg Ala Leu Ser Gln				



1145	1150	1155	1160	
ttt gaa gga atg aag gtg gtg gtc aat gct ctc ata ggt gcc ata cct				3558
Phe Glu Gly Met Lys Val Val Val Asn Ala Leu Ile Gly Ala Ile Pro				
	1165	1170	1175	
gcc att ctg aat gtt ttg ctt gtc tgc ctc att ttc tgg ctc gta ttt				3606
Ala Ile Leu Asn Val Leu Leu Val Cys Leu Ile Phe Trp Leu Val Phe				
	1180	1185	1190	
tgt att ctg gga gta tac ttc ttt tct gga aaa ttt ggg aaa tgc att				3654
Cys Ile Leu Gly Val Tyr Phe Phe Ser Gly Lys Phe Gly Lys Cys Ile				
	1195	1200	1205	
aat gga aca gac tca gtt ata aat tat acc atc att aca aat aaa agt				3702
Asn Gly Thr Asp Ser Val Ile Asn Tyr Thr Ile Ile Thr Asn Lys Ser				
	1210	1215	1220	
caa tgt gaa agt ggc aat ttc tct tgg atc aac cag aaa gtc aac ttt				3750
Gln Cys Glu Ser Gly Asn Phe Ser Trp Ile Asn Gln Lys Val Asn Phe				
	1225	1230	1235	1240
gac aat gtg gga aat gct tac ctc gct ctg ctg caa gtg gca aca ttt				3798
Asp Asn Val Gly Asn Ala Tyr Leu Ala Leu Leu Gln Val Ala Thr Phe				
	1245	1250	1255	
aag ggc tgg atg gat att ata tat gca gct gtt gat tcc aca gag aaa				3846
Lys Gly Trp Met Asp Ile Ile Tyr Ala Ala Val Asp Ser Thr Glu Lys				
	1260	1265	1270	
gaa caa cag cca gag ttt gag agc aat tca ctc ggt tac att tac ttc				3894
Glu Gln Gln Pro Glu Phe Glu Ser Asn Ser Leu Gly Tyr Ile Tyr Phe				
	1275	1280	1285	
gta gtc ttt atc atc ttt ggc tca ttc ttc act ctg aat ctc ttc att				3942
Val Val Phe Ile Ile Phe Gly Ser Phe Phe Thr Leu Asn Leu Phe Ile				
	1290	1295	1300	
ggc gtt atc att gac aac ttc aac caa cag cag aaa aag tta ggt ggc				3990
Gly Val Ile Ile Asp Asn Phe Asn Gln Gln Gln Lys Lys Leu Gly Gly				
	1305	1310	1315	1320
caa gac att ttt atg aca gaa gaa cag aag aaa tac tat aat gca atg				4038
Gln Asp Ile Phe Met Thr Glu Glu Gln Lys Lys Tyr Tyr Asn Ala Met				
	1325	1330	1335	
aaa aaa tta gga tcc aaa aaa cct caa aaa ccc att cca cgg cct ctg				4086
Lys Lys Leu Gly Ser Lys Lys Pro Gln Lys Pro Ile Pro Arg Pro Leu				
	1340	1345	1350	
aac aaa tgt caa ggt ctc gtg ttc gac ata gtc aca agc cag atc ttt				4134
Asn Lys Cys Gln Gly Leu Val Phe Asp Ile Val Thr Ser Gln Ile Phe				
	1355	1360	1365	
gac atc atc atc ata agt ctc att atc cta aac atg att agc atg atg				4182
Asp Ile Ile Ile Ile Ser Leu Ile Ile Leu Asn Met Ile Ser Met Met				

1370	1375	1380	
gct gaa tca tac aac caa ccc aaa gcc atg aaa tcc atc ctt gac cat			4230
Ala Glu Ser Tyr Asn Gln Pro Lys Ala Met Lys Ser Ile Leu Asp His			
1385	1390	1395	1400
ctc aac tgg gtc ttt gtg gtc atc ttt acg tta gaa tgt ctc atc aaa			4278
Leu Asn Trp Val Phe Val Val Ile Phe Thr Leu Glu Cys Leu Ile Lys			
	1405	1410	1415
atc ttt gct ttg agg caa tac tac ttc acc aat ggc tgg aat tta ttt			4326
Ile Phe Ala Leu Arg Gln Tyr Tyr Phe Thr Asn Gly Trp Asn Leu Phe			
	1420	1425	1430
gac tgt gtg gtc gtg ctt ctt tcc att gtt agt aca atg att tct acc			4374
Asp Cys Val Val Val Leu Leu Ser Ile Val Ser Thr Met Ile Ser Thr			
	1435	1440	1445
ttg gaa aat cag gag cac att cct ttc cct ccg acg ctc ttc aga att			4422
Leu Glu Asn Gln Glu His Ile Pro Phe Pro Pro Thr Leu Phe Arg Ile			
	1450	1455	1460
gtc cgc ttg gct cgg att ggc cga atc ctg agg ctt gtc cgg gct gca			4470
Val Arg Leu Ala Arg Ile Gly Arg Ile Leu Arg Leu Val Arg Ala Ala			
	1465	1470	1475
cga gga atc agg act ctc ctc ttt gct ctg atg atg tcg ctt cct tct			4518
Arg Gly Ile Arg Thr Leu Leu Phe Ala Leu Met Met Ser Leu Pro Ser			
	1485	1490	1495
ctg ttc aac att ggt ctt cta ctc ttt ctg att atg ttt atc tat gcc			4566
Leu Phe Asn Ile Gly Leu Leu Leu Phe Leu Ile Met Phe Ile Tyr Ala			
	1500	1505	1510
att ctg ggt atg aac tgg ttt tcc aaa gtg aat cca gag tct gga atc			4614
Ile Leu Gly Met Asn Trp Phe Ser Lys Val Asn Pro Glu Ser Gly Ile			
	1515	1520	1525
gat gac ata ttc aac ttc aag act ttt gcc agc agc atg ctc tgt ctc			4662
Asp Asp Ile Phe Asn Phe Lys Thr Phe Ala Ser Ser Met Leu Cys Leu			
	1530	1535	1540
ttc cag ata agc aca tca gca ggt tgg gat tcc ctg ctc agc ccc atg			4710
Phe Gln Ile Ser Thr Ser Ala Gly Trp Asp Ser Leu Leu Ser Pro Met			
	1545	1550	1555
ctg cga tca aaa gaa tca tgt aac tct tcc tca gaa aac tgc cac ctc			4758
Leu Arg Ser Lys Glu Ser Cys Asn Ser Ser Ser Glu Asn Cys His Leu			
	1565	1570	1575
cct ggc ata gcc aca tcc tac ttt gtc agt tac att atc atc tcc ttt			4806
Pro Gly Ile Ala Thr Ser Tyr Phe Val Ser Tyr Ile Ile Ile Ser Phe			
	1580	1585	1590
ctc att gtt gtc aac atg tac att gct gtg att tta gag aac ttc aat			4854
Leu Ile Val Val Asn Met Tyr Ile Ala Val Ile Leu Glu Asn Phe Asn			

1595	1600	1605	
aca gcc act gaa gaa agt gag gac cct ttg ggt gaa gat gac ttt gac			4902
Thr Ala Thr Glu Glu Ser Glu Asp Pro Leu Gly Glu Asp Asp Phe Asp			
1610	1615	1620	
ata ttt tat gaa gtg tgg gaa aag ttt gac cca gaa gca aca caa ttt			4950
Ile Phe Tyr Glu Val Trp Glu Lys Phe Asp Pro Glu Ala Thr Gln Phe			
1625	1630	1635	1640
atc aaa tat tct gcc ctt tct gac ttt gct gat gcc ttg cct gag cct			4998
Ile Lys Tyr Ser Ala Leu Ser Asp Phe Ala Asp Ala Leu Pro Glu Pro			
1645	1650	1655	
ttg cgt gtc gca aag cca aat aaa tat caa ttt cta gta atg gac ttg			5046
Leu Arg Val Ala Lys Pro Asn Lys Tyr Gln Phe Leu Val Met Asp Leu			
1660	1665	1670	
ccc atg gtg agt gaa gat cgc ctc cac tgc atg gat att ctt ttc gcc			5094
Pro Met Val Ser Glu Asp Arg Leu His Cys Met Asp Ile Leu Phe Ala			
1675	1680	1685	
ttc acc gct agg gta ctc ggt ggc tct gat ggc cta gat agt atg aaa			5142
Phe Thr Ala Arg Val Leu Gly Gly Ser Asp Gly Leu Asp Ser Met Lys			
1690	1695	1700	
gca atg atg gaa gag aag ttc atg gaa gcc aat cct ctc aag aag ttg			5190
Ala Met Met Glu Glu Lys Phe Met Glu Ala Asn Pro Leu Lys Lys Leu			
1705	1710	1715	1720
tat gaa ccc ata gtc acc acc acc aag aga aag gaa gag gaa aga ggt			5238
Tyr Glu Pro Ile Val Thr Thr Thr Lys Arg Lys Glu Glu Glu Arg Gly			
1725	1730	1735	
gct gct att att caa aag gcc ttt cga aag tac atg atg aag gtg acc			5286
Ala Ala Ile Ile Gln Lys Ala Phe Arg Lys Tyr Met Met Lys Val Thr			
1740	1745	1750	
aag ggt gac caa ggt gac caa aat gac ttg gaa aac ggg cct cat tca			5334
Lys Gly Asp Gln Gly Asp Gln Asn Asp Leu Glu Asn Gly Pro His Ser			
1755	1760	1765	
cca ctc cag act ctt tgc aat gga gac ttg tct agc ttt ggg gtg gcc			5382
Pro Leu Gln Thr Leu Cys Asn Gly Asp Leu Ser Ser Phe Gly Val Ala			
1770	1775	1780	
aag ggc aag gtc cac tgt gac tgagccctca cctccacgcc tacctcatag			5433
Lys Gly Lys Val His Cys Asp			
1785	1790		
cttcacagcc ttgccttcag cctctgagct ccaggggtca gcagcttagt gtatcaacag			5493
ggagtggatt caccaaatta gccattccat tttcttttct ggctaaaata aatgatattt			5553
caatttcatt ttaaatagata cttacagaga tataagataa ggctacttga caaccagtgg			5613

tactattata ataaggaaga agacaccagg aaggactgta aaaggacata ccaatttttag 5673  
 gattgaaata gttcaggccg ggcgcagtgg ctcatgcctg taatcccagc actttgagag 5733  
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<211> 1791

<212> PRT

<213> Homo sapiens

<400> 42

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Ile	Gln	Lys	Glu	Lys	Lys	Lys	Ser	Lys	Asp	Gln	Thr	Gly	Glu	Val	Pro	35	40	45	
Gln	Pro	Arg	Pro	Gln	Leu	Asp	Leu	Lys	Ala	Ser	Arg	Lys	Leu	Pro	Lys	50	55	60	
Leu	Tyr	Gly	Asp	Ile	Pro	Arg	Glu	Leu	Ile	Gly	Lys	Pro	Leu	Glu	Asp	65	70	75	80
Leu	Asp	Pro	Phe	Tyr	Arg	Asn	His	Lys	Thr	Phe	Met	Val	Leu	Asn	Arg	85	90	95	
Lys	Arg	Thr	Ile	Tyr	Arg	Phe	Ser	Ala	Lys	His	Ala	Leu	Phe	Ile	Phe	100	105	110	
Gly	Pro	Phe	Asn	Ser	Ile	Arg	Ser	Leu	Ala	Ile	Arg	Val	Ser	Val	His	115	120	125	
Ser	Leu	Phe	Ser	Met	Phe	Ile	Ile	Gly	Thr	Val	Ile	Ile	Asn	Cys	Val	130	135	140	
Phe	Met	Ala	Thr	Gly	Pro	Ala	Lys	Asn	Ser	Asn	Ser	Asn	Asn	Thr	Asp	145	150	155	160
Ile	Ala	Glu	Cys	Val	Phe	Thr	Gly	Ile	Tyr	Ile	Phe	Glu	Ala	Leu	Ile	165	170	175	
Lys	Ile	Leu	Ala	Arg	Gly	Phe	Ile	Leu	Asp	Glu	Phe	Ser	Phe	Leu	Arg	180	185	190	
Asp	Pro	Trp	Asn	Trp	Leu	Asp	Ser	Ile	Val	Ile	Gly	Ile	Ala	Ile	Val	195	200	205	

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 Arg Val Phe Arg Ala Leu Lys Ala Ile Ser Val Val Ser Arg Leu Lys  
 225 230 235 240  
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 Lys Arg Lys Ser Phe Phe Leu Arg Glu Ser Gly Lys Asp Gln Pro Pro  
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 Gly Ser Asp Ser Asp Glu Asp Cys Gln Lys Lys Pro Gln Leu Leu Glu  
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His Gly Asp Pro Leu Gln Arg Gln Arg Ala Leu Ser Ala Val Ser Ile  
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 Phe Leu Ala Met Glu His His Lys Met Glu Ala Ser Phe Glu Lys Met  
 595 600 605  
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 Cys Leu Lys Ile Ile Ala Leu Asp Pro Tyr His Tyr Phe Arg Arg Gly  
 625 630 635 640  
 Trp Asn Ile Phe Asp Ser Ile Val Ala Leu Leu Ser Phe Ala Asp Val  
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 Met Asn Cys Val Leu Gln Lys Arg Ser Trp Pro Phe Leu Arg Ser Phe  
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 Arg Val Leu Arg Val Phe Lys Leu Ala Lys Ser Trp Pro Thr Leu Asn  
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 Thr Leu Ile Lys Ile Ile Gly Asn Ser Val Gly Ala Leu Gly Ser Leu  
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 Gln Leu Phe Gly Arg Ser Phe Asn Ser Gln Lys Ser Pro Lys Leu Cys  
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 Val Val Leu Asn Leu Phe Ile Ala Leu Leu Leu Asn Ser Phe Ser Asn  
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<213> Artificial Sequence

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<210> 44

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<212> DNA

<213> Artificial Sequence

<220>

<223> human NaN reverse primer


<400> 44

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23

## INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US00/19342

<b>A. CLASSIFICATION OF SUBJECT MATTER</b> IPC(7) :C07K 14/705; C12N 5/10, 15/12, 15/63 US CL :435/320.1, 325; 530/350; 536/23.5 According to International Patent Classification (IPC) or to both national classification and IPC														
<b>B. FIELDS SEARCHED</b> Minimum documentation searched (classification system followed by classification symbols) U.S. : 435/320.1, 325; 530/350; 536/23.5  Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched  Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) BRS, STN, MEDLINE, GENESEQ, PIR, SWISS-PROT, STREMBL, GENEMBL search terms: sodium channel, tetrodotoxin														
<b>C. DOCUMENTS CONSIDERED TO BE RELEVANT</b>														
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.												
X - A	WO 97/01577 A1 (UNIVERSITY COLLEGE LONDON) 16 January 1997, see especially pages 85-93.	1-6, 8-9, 22, 25 ----- 7, 33-35												
<input type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.														
<table border="0"><tr><td>* Special categories of cited documents:</td><td>*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</td></tr><tr><td>*A* document defining the general state of the art which is not considered to be of particular relevance</td><td>*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone</td></tr><tr><td>*B* earlier document published on or after the international filing date</td><td>*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art</td></tr><tr><td>*L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</td><td>*G* document member of the same patent family</td></tr><tr><td>*O* document referring to an oral disclosure, use, exhibition or other means</td><td></td></tr><tr><td>*P* document published prior to the international filing date but later than the priority date claimed</td><td></td></tr></table>			* Special categories of cited documents:	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention	*A* document defining the general state of the art which is not considered to be of particular relevance	*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone	*B* earlier document published on or after the international filing date	*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art	*L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*G* document member of the same patent family	*O* document referring to an oral disclosure, use, exhibition or other means		*P* document published prior to the international filing date but later than the priority date claimed	
* Special categories of cited documents:	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention													
*A* document defining the general state of the art which is not considered to be of particular relevance	*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone													
*B* earlier document published on or after the international filing date	*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art													
*L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*G* document member of the same patent family													
*O* document referring to an oral disclosure, use, exhibition or other means														
*P* document published prior to the international filing date but later than the priority date claimed														
Date of the actual completion of the international search 10 SEPTEMBER 2000		Date of mailing of the international search report 02 NOV 2000												
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. (703) 305-3230		Authorized officer  MICHAEL PAK Telephone No. (703) 308-0196												

# INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US00/19342

## Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:  
because they relate to subject matter not required to be searched by this Authority, namely:
  
2. ☐ Claims Nos.:  
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
  
3. ☐ Claims Nos.:  
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

## Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Please See Extra Sheet.

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
  
4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:  
1-9, 22, 25, 33-35

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.  
☐ No protest accompanied the payment of additional search fees.

## INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US00/19342

### BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be searched, the appropriate additional search fees must be paid.

Group I, claim(s) 1-9, 22, 25, 33-35, drawn to an isolated nucleic acid molecule, a method of producing a transformed cell, a Na<sup>+</sup> channel and a method of producing recombinant NaN protein.

Group II, claim(s) 10-14, drawn to a method to identify an agent that modulates the activity of the Na<sup>+</sup> channel.

Group III, claim(s) 15, drawn to a method to identify an agent that modulates the transcription or translation of mRNA.

Group IV, claim(s) 16, drawn to a method to treat pain by administering an agent that modulates the activity of the Na<sup>+</sup> channel.

Group V, claim(s) 17, drawn to a method to treat pain by administering an agent that modulates the transcription or translation of mRNA.

Group VI, claim(s) 18, drawn to an isolated nucleic acid that is antisense.

Group VII, claim(s) 19, drawn to a scintigraphic method to image loci of pain generation.

Group VIII, claim(s) 20, drawn to a method to identify tissues by detecting NaN.

Group IX, claim(s) 21, drawn to a method to identify tissues by detecting mRNA.

Group X, claim(s) 23-24, drawn to an antibody.

Group XI, claim(s) 26-27 and 36-37, drawn to a therapeutic composition comprising an agent and a method of treatment using the agent of claim 26.

Group XII, claim(s) 28, drawn to a method to screen candidate compound for use in treating pain.

Group XIII, claim(s) 29-32, drawn to a chimeric NaN channel and the DNA encoding the chimeric NaN channel.

The inventions listed as Groups I-XIII do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features because the product of claim 1 is anticipated by UNIVERSITY COLLEGE LONDON (WO 97/01577A1 16 January 1997) and thus, does not share a special technical feature with any other group.

The products of Group VI, X-XI, and XIII does not share the same or corresponding special technical feature with Group I, because they are drawn to products having materially different structures and functions, and each defines a separate invention over the art.

The methods of Groups II-V, VII-IX, and XII do not share the same or corresponding special technical feature with Group I, because the methods have materially different process steps and are practiced for materially different purposes, and each defines a separate invention over the art.

Since Groups I-XIII do not share a special technical feature, unity of invention is lacking.